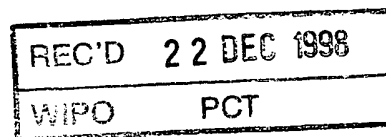




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AUSTRALIA

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COMMONWEALTH SCIENTIFIC AND INDUSTRIAL RESEARCH ORGANISATION

PROVISIONAL SPECIFICATION

Invention Title:

Receptor agonists and antagonists

The invention is described in the following statement:

RECEPTOR AGONISTS AND ANTAGONISTS

Field of the Invention

This invention relates to the field of receptor structure and
receptor/ligand interactions. In particular it relates to the field of using

- 5 receptor structure to predict the structure of related receptors and to use the determined structures and predicted structures to select and screen for agonists and antagonists of the polypeptide ligands.

Background of the Invention

- Insulin is the peptide hormone that regulates glucose uptake and
10 metabolism. The two types of diabetes are associated with either an inability to produce insulin because of destruction of the pancreatic islet cells (Homo-Delarche, F. & Boitard, C., 1996, Immunol. Today 10: 456-460) or poor glucose metabolism resulting from either insulin resistance at the target tissues, inadequate insulin secretion by the islets or faulty liver function (Taylor, S.
15 I., et al., 1994, Diabetes, 43: 735-740).

- Insulin-like growth factors-1 and 2 (IGF-1 and 2) are structurally related to insulin but are more important in tissue growth and development than in metabolism. They are primarily produced in the liver in response to growth hormone but are also produced in most other tissues where they
20 function as paracrine/autocrine regulators. The IGFs are strong mitogens and are involved in numerous physiological states and certain cancers (Baserga, R., 1996, TibTech 14: 150-152).

- Epidermal growth factor (EGF) is a small polypeptide cytokine that is unrelated to the insulin/IGF family. It stimulates marked proliferation of
25 epithelial tissues and is a member of a larger family of structurally related cytokines such as transforming growth factor α , amphiregulin, betacellulin, heparin-binding EGF and some viral gene products. Abnormal EGF family signalling is a characteristic of certain cancers (Soler, C. & Carpenter, G., 1994 In Nicola, N. (ed) Guidebook to Cytokines and Their receptors", Oxford
30 Univ. Press, Oxford, pp194-197; Walker, F. & Burgess, A. W., 1994, In Nicola, N. (ed) Guidebook to Cytokines and Their receptors", Oxford Univ. Press, Oxford, pp198-201).

- Each of these growth factors mediate their biological actions through binding to the corresponding receptor. The IR, IGF-1R and insulin receptor-related receptor (IRR), for which the ligand is not known, are closely related
35 to each other and are referred to as the insulin receptor subfamily. There is a

large body of information now available concerning the primary structure of these insulin receptor subfamily members (Ebina, Y., et al., 1985 *Cell* 40: 747-758; Ullrich, A., et al., 1985, *Nature* 313: 756-761; Ullrich, A. et al.,

1986, *EMBO J* 5: 2503-2512; Shier, P. & Watt, V. M., 1989, *J. Biol. Chem.* 264:

5 14605-14608) and the identification of some of their functional domains (for reviews see De Meyts, P. 1994, *Diabetologia* 37: 135-148; Lee, J. & Pilch, P. F. 1994 *Amer. J. Physiol.* 266: C319-C334.; Schaffer, L. 1994, *Eur. J. Biochem.* 221: 1127-1132). IGF-1R, IR and IRR are members of the tyrosine kinase receptor superfamily and are closely related to the epidermal growth factor
10 receptor (EGFR) subfamily, with which they share significant sequence identity in the extracellular region as well as in the cytoplasmic kinase domains (Ullrich, A. et al., 1984 *Nature* 309: 418-425; Ward, C. W. et al., 1995 *Proteins: Structure Function & Genetics* 22: 141-153). Both the insulin and
15 EGF receptor subfamilies have a similar arrangement of two homologous domains (L1 and L2) separated by a cys-rich region of approximately 160 amino acids containing 22-24 cys residues (Bajaj, M., et al., 1987 *Biochim. Biophys. Acta* 916: 220-226; Ward, C. W. et al., 1995 *Proteins: Structure Function & Genetics* 22: 141-153). The C-terminal portion of the IGF-1R ectodomain (residues 463 to 906) is comprised of four domains: a connecting
20 domain, two fibronectin type 3 (Fn3) repeats, and an insert domain (O'Bryan, J. P., et al., 1991 *Mol Cell Biol* 11: 5016-5031); the C-terminal portion of the EGFR ectodomain (residues 477-621) consists solely of a second cys-rich region containing 20 cys residues (Ullrich, A. et al., 1984, *Nature* 309: 418-425).

25 Little is known about the secondary, tertiary and quaternary structure of the ectodomains of these receptor subfamilies. Unlike the members of the EGFR subfamily which are transmembrane monomers which dimerise on binding ligand, the IR subfamily members are homodimers, held together by disulphide bonds. The extracellular region of the IR/IGF-1R/IRR monomers
30 contains an α -chain (\sim 703 to 735 amino acid residues) and 192-196 residues of the β -chain. There is a \sim 23 residue transmembrane segment, followed by the cytoplasmic portion (354 to 408 amino acids) which contains the catalytic tyrosine kinase domain flanked by juxtamembrane and C-tail regulatory regions and is responsible for mediating all receptor-specific
35 functions (White, M. F. & Kahn, C. R. 1994 *J. Biol. Chem.* 269: 1-4). Chemical analyses of the receptor suggest that the α -chains are linked to the β -chains

via a single disulphide bond with the IR dimer being formed by at least two α - α disulphide linkages (Finn, F. M., et al., 1990, Proc. Natl. Acad. Sci. 87: 419-423; Chiacchia, K. B., 1991, Biochem. Biophys. Res. Commun. 176, 1178-

1182; Schaffer, L. & Ljungqvist, L., 1992, Biochem. Biophys. Res. Comm. 189: 650-653; Sparrow, L. G., et al., 1997, J. Biol. Chem. 47: 29460-29467).

Although the 3D structures of the ligands EGF, TGF- α (Hommel, U., et al., 1992, J. Mol. Biol. 227:271-282), insulin (Dodson, E. J., et al., 1983, Biopolymers 22:281-291), IGF-1 (Sato, A., et al., 1993, Int J Peptide Protein Res 41:433-440) and IGF-2 (Torres, A. M., et al., 1995, J. Mol. Biol. 248:385-401) are known and numerous analytical and functional studies of ligand binding to EGFR (Soler, C. & Carpenter, G., 1994 In Nicola (ed) Guidebook to Cytokines and Their receptors", Oxford Univ. Press, Oxford, pp194-197), IGF-1R and IR (see De Meyts, P., 1994 Diabetologia, 37:135-148) have been carried out, the mechanisms of ligand binding and subsequent transmembrane signalling have not been resolved.

Ligand-induced, receptor-mediated phosphorylation is the signalling mechanism by which most cytokines, polypeptide hormones and membrane-anchored ligands exert their biological effects. The primary kinase may be part of the intracellular portion of the transmembrane receptor protein as in the tyrosine kinase receptors (for review see Yarden, Y., et al., 1988, Ann. Rev. Biochem. 57:443-478) or the Ser/Thr kinase receptors (Alevizopoulos, A. & Mermoud, N., 1997, BioEssays, 19:581-591) or be non-covalently associated with the cytoplasmic tail of the transmembrane protein(s) making up the receptor complex as in the case of the haemopoietic growth factor receptors (Stahl, N., et al., 1995, Science 267:1349-1353). The end result is the same, ligand binding leads to receptor dimerization or oligomerization or a conformational change in pre-existing receptor dimers or oligomers resulting in activation by transphosphorylation, of the covalently attached or non-covalently associated protein kinase domains (Hunter, T., 1995, Cell, 80:225-236).

Many oncogenes have been shown to be homologous to growth factors, growth factor receptors or molecules in the signal transduction pathways (Baserga, R., 1994 Cell, 79:927-930; Hunter, T., 1997 Cell, 88:333-346). One of the best examples is v-Erb (related to the EGFR). Since overexpression of a number of growth factor receptors results in ligand-dependent transformation an alternate strategy for oncogenes is to regulate

the expression of growth factor receptors or their ligands or to directly bind to the receptors to stimulate the same effect (Baserga, R., 1994 Cell, 79:927-930). Examples are v-Src, which activates IGF-1 R intracellularly; c-Myb, which transforms cells by enhancing the expression of IGF1R and SV40 T

5 antigen which interacts with the IGF-1R and enhances the secretion of IGF-1 (see Baserga, R., 1994 Cell, 79:927-930 for review). Cells in which the IGF-1 receptor has been knocked out cannot be transformed by SV40 T antigen. If oncogenes activate growth factors and their receptors then tumour suppressor genes should have the opposite effect. One good example of this
10 is WT1, the Wilm's tumour suppressor gene which suppresses the expression of IGF-1R (Drummond, J. A., et al., 1992, Science, 257:275-277). Cells that are driven to proliferate by oncogenes undergo massive apoptosis when growth factor receptors are ablated since unlike normal cells, they appear unable to withdraw from the cell-cycle and enter into the G0 phase (Baserga, R., 1994
15 Cell, 79:927-930).

The insulin-like growth factor-1 receptor (IGF-1R) is one of several growth-factor receptors that regulate the proliferation of mammalian cells. However, its ubiquitousness and certain unique aspects of its function make IGF-1R an ideal target for therapeutic interventions against abnormal growth,
20 with very little effect on normal cells (see Baserga, R., 1996 TIBTECH, 14:150-152). The receptor is activated by IGF1, IGF2 and insulin and plays a major role in cellular proliferation in at least three ways: it is essential for optimal growth of cells in vitro and in vivo; several cell types require IGF-1R to maintain the transformed state and activated IGF-1R has a protective effect
25 against apoptotic cell death (Baserga, R., 1996 TIBTECH, 14:150-152). These properties alone make it an ideal target for therapeutic interventions. Transgenic experiments have shown that IGF-1R is not an absolute requirement for cell growth but is essential for the establishment of the transformed state (Baserga, R., 1994 Cell, 79: 927-930). In several cases
30 (human glioblastoma, human melanoma; human breast carcinoma; human lung carcinoma; human ovarian carcinoma; human rhabdomyosarcoma; mouse melanoma, mouse leukaemia; rat glioblastoma; rat rhabdomyosarcoma; hamster mesothelioma) the transformed phenotype can be reversed by decreasing the expression of IGF-1R using antisense to IGF-1R
35 (Baserga, R., 1996 TIBTECH 14:150-152); or interfering with its function by antibodies to IGF-1R (human breast carcinoma; human rhabdomyosarcoma)

or by dominant negatives of IGF-1R (rat glioblastoma; Baserga, R., 1996 TIBTECH 14:150-152).

Three effects are observed when the function of IGF-1R is impaired:

-
- 5 tumour cells undergo massive apoptosis which results in inhibition of tumourogenesis; surviving tumour cells are eliminated by a specific immune response; and such a host response can cause a regression of an established wild-type tumour (Resnicoff, M., et al., 1995, Cancer Res. 54:2218-2222). These effects, plus the fact that interference of IGF-1R function has a limited effect on normal cells (partial inhibition of growth without apoptosis) makes
- 10 IGF-1R a unique target for therapeutic interventions (Baserga, R., 1996 TIBTECH 14:150-152). In addition IGF-1R is downstream of many other growth factor receptors, which makes it an even more generalised target. The implication of these findings is that if you can decrease the number of IGF-1 receptors on cells or antagonise their function then tumours cease to grow
- 15 and can be removed immunologically. These studies establish that IGF-1R antagonists will be extremely important therapeutically.

- Many cancer cells have constitutively active EGFR (Sandgreen, E. P., et al., 1990, Cell, 61:1121-135; Karnes, W. E. J., et al., 1992, Gastroenterology, 102:474-485) or other EGFR family members (Hines, N. E., 1993, Semin.
- 20 Cancer Biol. 4:19-26). Elevated levels of activated EGFR occur in bladder, breast, lung and brain tumours (Harris, A. L., et al., 1989, In Furth & Greaves (eds) The Molecular Diagnostics of human cancer. Cold Spring Harbor Lab. Press, CSH, NY, pp353-357). Antibodies to EGFR can inhibit ligand activation of EGFR (Sato, J. D., et al., 1983 Mol. Biol. Med. 1:511-529) and the growth of
- 25 many epithelial cell lines (Aboud-Pirak E., et al., 1988, J. Natl Cancer Inst. 85:1327-1331). Patients receiving repeated doses of a humanised chimeric anti-EGFR antibody showed signs of disease stabilization. The large doses required and the cost of production of humanised Mab is likely to limit the application of this type of therapy. These findings indicate that the
- 30 development of EGF antagonists will be attractive anticancer agents.

Summary of the Invention

- The present inventors have now obtained 3D structural information concerning the insulin-like growth factor receptor (IGF-1R) and the insulin receptor (IR) which provides a rational basis for the development of
- 35 antagonists and agonists of the polypeptide ligands for specific therapeutic applications. This information can be used to predict the structure of related

members of the insulin receptor family and epidermal growth factor family and to develop agonists and antagonists of their respective polypeptide ligands.

-
- Accordingly, in a first aspect the present invention provides a method
- 5 of screening for, or designing, an agonist of a ligand of an insulin receptor family member or EGF receptor family member which method includes
- (i) selecting or designing a substance which possesses stereochemical complementarity to a receptor site, wherein the receptor site is characterised by
- 10 (a) amino acids 1-462 of IGF-1R positioned at atomic coordinates substantially as shown in Figure 1 or a subset thereof; or
- (b) amino acids derived from an insulin receptor family member or EGF receptor family member which form an equivalent structure to the amino acids defined in paragraph (a); and
- 15 (ii) testing the substance for the ability to act as an agonist of the ligand of an insulin receptor family member or EGF receptor family member.

In a second aspect the present invention provides a method of screening for, or designing, an antagonist of a ligand of an insulin receptor family member or EGF receptor family member which method includes

- 20 (i) selecting or designing a substance which possesses stereochemical complementarity to a receptor site, wherein the receptor site is characterised by
- (a) amino acids 1-462 of IGF-1R positioned at atomic coordinates substantially as shown in Figure 1 or a subset thereof; or
- 25 (b) amino acids derived from an insulin receptor family member or an EGF receptor family member which form an equivalent structure to the amino acids defined in paragraph (a); and
- (ii) testing the substance for the ability to act as an antagonist of the ligand of an insulin receptor family member or EGF receptor family member.

30 The phrase "insulin receptor family" encompasses, for example, IGF-1R, IR and IRR. The phrase "EGF receptor family" encompasses for example, EGFR, ErbB2, ErbB3 and ErbB4. In general, insulin receptor family members and EGF receptor family members show similar domain arrangements and share significant sequence identity (preferably at least 20% identity between

35 the families and at least 40% identity within each family).

The receptor site defined in the first and second aspects of the present invention comprises the L1-cysteine rich-L2 region (residues 1-462) of the ectodomain of IGF-1R. ~~At the centre of this structure is a groove,~~

5 bounded by all three domains, of sufficient size to accommodate a ligand molecule. By "stereochemical complementarity" we mean that the biologically active substance or a portion thereof correlates, in the manner of the classic "lock-and-key" visualisation of ligand-receptor interaction, with the groove in the receptor site. Preferably, the stereochemical complementarity is such that the compound has a K_i for the receptor site of
 10 less than 10^{-6} M. More preferably, the K_i value is less than 10^{-8} M and more preferably less than 10^{-9} M.

In preferred embodiments of the first and second aspects of the present invention, the method further involves selecting or designing a substance which has portions that match residues positioned on the surface
 15 of the receptor site which faces the groove. By "match" we mean that the identified portions interact with the surface residues, for example, via hydrogen bonding or by enthalpy-reducing Van der Waals interactions which promote desolvation of the biologically active substance within the site, in such a way that retention of the biologically active substance within the
 20 groove is favoured energetically.

In a preferred embodiment of the first aspect of the present invention, the method includes screening for, or designing, a substance which possesses a stereochemistry and/or geometry which allows it to interact with both the L1 and L2 domains of the receptor site. As described above, the insulin
 25 receptor exists as homodimers held together by disulphide bonds. Electron microscopy studies described herein indicate that the insulin receptor monomers dimerise in nature in such a manner that the grooves of each monomer may face each other. Accordingly, the method of the first aspect of the present invention may involve screening for, or designing, a biologically
 30 active substance which interacts with the L1 domain of one monomer and the L2 domain of the other monomer.

In a third aspect the present invention provides a method of selecting or designing an agonist of a ligand of an insulin receptor family member or EGF receptor family member which method includes

35 (i) selecting or designing a substance which interacts with

(a) a fragment of IGF-1R characterised by amino acids 1-462 positioned at atomic coordinates substantially as shown in Figure 1 or a subset thereof; or

(b) a fragment derived from an insulin family receptor member
5 or EGF receptor family member which is equivalent to the fragment defined in paragraph (a);

wherein the interaction of the substance with the fragment alters the position of at least one of the L1, L2 or cys-rich domains of the fragment relative to the position of at least one of the other domains; and

10 (ii) testing the substance for the ability to act as an agonist of the ligand of an insulin receptor family member or EGF receptor family member.

In a preferred embodiment of the third aspect of the present invention the substance interacts with the fragment in the region of the L1 domain-cys rich domain interface, causing the L1 and cys-rich domains to
15 move away from each other. In a further preferred embodiment the substance interacts with the hinge region between the L2 domain and the cys-rich domain causing an alteration in the positions of the domains relative to each other. In a further preferred embodiment the substance interacts with the beta sheet of the L1 domain causing an alteration in the position of
20 the L1 domain relative to the position of the cys-rich domain or L2 domain.

In a fourth aspect the present invention provides an agonist of a ligand of an insulin receptor family member or EGF receptor family member obtained by a method according to the first or third aspects of the present invention.

25 In a fifth aspect the present invention provides an antagonist of ligand of an insulin receptor family member or EGF receptor family member obtained by a method according to the second aspect of the present invention.

The agonists or antagonists of the fourth and fifth aspects of the
30 present invention may be mutant insulin family member or EGF family member ligands where at least one mutation occurs in the region of the ligand which interacts with residues on the surface of the receptor site facing toward the groove. For example, the IGF-1 ligand has a predominance of basic residues in the C region which may interact with the acidic patch of the
35 cys-rich region near L1. An acidic patch on the other side of the ligand may interact with the patch of basic residues (residues 307-310) on the N-terminal

end of L2. Accordingly, mutants of IGF-1 which exhibit altered activity may be generated by introducing modifications in the C region of IGF-1 or residues in the acidic patch on the other side of the hormone.

5 In a sixth aspect the present invention provides a substance which possesses stereochemical complementarity to a receptor site, wherein the receptor site is characterised by

(a) amino acids 1-462 of IGF-1R positioned at atomic coordinates substantially as shown in Figure 1 or a subset thereof; or

10 (b) amino acids derived from an insulin receptor family member or an EGF receptor family member which form an equivalent structure to the amino acids defined in paragraph (a);

with the proviso that the substance is not a naturally occurring ligand of an insulin receptor family member or EGF receptor family member or a mutant thereof.

15 By "mutant" we mean a ligand which has been modified by one or more point mutations, insertions of amino acids or deletions of amino acids.

In a preferred embodiment of the sixth aspect of the present invention, the stereochemical complementarity is such that the compound has a K_i for the receptor site of less than 10^{-6} M. More preferably, the K_i value
20 is less than 10^{-8} M and more preferably less than 10^{-9} M.

In a seventh aspect the present invention provides a pharmaceutical composition for treatment of a disease associated with reduced activity of a ligand of an insulin receptor family member or EGF receptor family member which includes an agonist obtained by a method according to the first or
25 third aspects of the present invention and a pharmaceutically acceptable carrier or diluent.

In an eighth aspect the present invention provides a pharmaceutical composition for treatment of a disease associated with activity of a ligand of an insulin receptor family member or EGF receptor family member which
30 includes an antagonist obtained by a method according to the second aspect of the present invention and a pharmaceutically acceptable carrier or diluent.

In a ninth aspect the present invention provides a method of preventing or treating a disease associated with reduced activity of a ligand of an insulin receptor family member or EGF receptor family member which
35 method includes administering to a subject in need thereof an agonist

obtained by a method according to the first or third aspects of the present invention.

Diseases associated with reduced activity of a ligand of an insulin receptor family member or EGF receptor family member include diabetes, osteoporosis, nerve degeneration and a range of catabolic states.

In a tenth aspect the present invention provides a method of preventing or treating a disease associated with activity of a ligand of an insulin receptor family member or EGF receptor family member which method includes administering to a subject in need thereof an antagonist obtained by a method according to the second aspect of the present invention.

Diseases associated with activity of a ligand of an insulin receptor family member or EGF receptor family member include cancer, leukaemia and many types of tumour states including but not restricted to breast cancer, brain tumours, ovarian cancer, pancreatic tumours, lung cancer, melanoma, rhabdomyosarcoma, mesothelioma and glioblastoma.

Brief Description of the Drawings

Figure 1. IGF-1R residues 1-462, in terms of atomic coordinates refined to a resolution of 2.6 Å (average accuracy \approx 0.3Å). The coordinates are in relation to a Cartesian system of orthogonal axes.

Figure 2. Depiction of the residues lining the groove of the IGF-1R receptor fragment 1-462.

Figure 3. Gel filtration chromatography of affinity-purified IGF-1R/462 protein. The protein was purified on a Superdex S200 column (Pharmacia) fitted to a BioLogic L.C. system (Biorad), equilibrated and eluted at 0.8 ml/min with 40 mM Tris/150 mM NaCl/0.02% NaN₃ adjusted to pH 8.0. (a) Protein eluting in peak 1 contained aggregated IGF-1R/462 protein, peak 2 contained monomeric protein and peak 3 contained the c-myc undecapeptide used for elution from the Mab 9E10 immunoaffinity column. (b) Non-reduced SDS-PAGE of fraction 2 from IGF-1R/462 obtained following Superdex S200 (Fig.1a). Standard proteins are indicated.

Figure 4. Ion exchange chromatography of affinity-purified, truncated IGF-1R ectodomain. A mixture of gradient and isocratic elution chromatography was performed on a Resource Q column (Pharmacia) fitted to a BioLogic System (Biorad), using 20 mM Tris/pH 8.0 as buffer A and the same buffer

5 containing 1M NaCl as buffer B. Protein solution in TBSA was diluted at least 1:2 with water and loaded onto the column at 2 ml/min. Elution was monitored by absorbance (280 nm) and conductivity (mS/cm). Target protein (peak 2) eluted isocratically with 20 mM Tris/0.14 M NaCl pH 8.0. Inset: Isoelectric focusing gel (pH 3 - 7; Novex Australia Pty Ltd) of fraction 2. The
10 pI was estimated at 5.1 from standard proteins (not shown).

Figure 5. Gel filtration chromatography of affinity purified IR/485 protein.

Affinity-purified material at 1 mg/ml produced a dominant peak at apparent mass ~ 140 kDa (interpreted as a dimer) (a); whereas affinity-purified
15 material at 0.02 mg/ml produced a dominant peak at apparent mass ~ 85kDa (interpreted as a monomer) (b).

Figure 6. (a) SDS-PAGE of IR/485 following gel filtration chromatography.

The protein migrated as a single broad band of apparent molecular mass ~ 78
20 kDa (reduced - lane A) or ~ 68kDa (non-reduced - lane B). (b) Isoelectric focussing of the IR/485 protein. The IR/485 fragment reacted positively in an ELISA with Mab 83-7, gave a single sequence corresponding to the N-terminal 10 residues of IR, showing several isoforms on isoelectric focussing from pI6.0-6.8. The fragment was further purified by ion-exchange
25 chromatography on Uno Q (BioRad, USA), using stepwise isocratic elution with incremental changes in salt concentrations (see Figure 7). Fractions A and D were each enriched in a component isoform from the ladder of isoforms present in the unfractionated mixture. Both these fractions produced crystals, whereas no crystals were obtained from fractions B and C.

30

Figure 7. Purification of the IR/485 protein by ion-exchange chromatography on Uno Q (BioRad, USA), using stepwise isocratic elution with incremental changes in salt concentrations.

35 **Figure 8.** Polypeptide fold for residues 1-462 of IGF-1R. The L1 domain is at the top, viewed from the N-terminal end and L2 is at the bottom. The space

at the centre is of sufficient size to accommodate IGF-1. Helices are indicated by curled ribbon and b-strands by arrows. Cysteine side chains are drawn as ball-and-stick with lines showing disulfide bonds. The arrow points in the direction of view for Figure 9.

5

Figure 9. Amino acid sequences of IGF-1R and related proteins. a, L1 and L2 domains of human IGF-1R and IR are shown based on a sequence alignment for the two proteins and a structural alignment for the L1 and L2 domains. Positions showing conservation physico-chemical properties of amino acids are boxed, residues used in the structural alignment are shaded yellow and residues which form the Trp 176 pocket are in red. Secondary structure elements for L1 (above the sequences) and L2 (below) are indicated as cylinders for helices and arrows for b-strands. Strands are colour coded according to the b-sheet to which they belong. Disulfide bonds are also indicated. b, Cys-rich domains of human IGF-1R, IR and EGFR (domains 2 and 4) are aligned based on sequence and structural considerations. Secondary structural elements and disulfide bonds are indicated above the sequences. The dashed bond is only present in IR. Different types of disulfide bonded modules are labelled below the sequences as open, filled or broken lines. Boxed residues show conservation of physico-chemical properties and structurally conserved residues for modules 4-7 are shaded yellow. Residues from EGFR which do not conform to the pattern are shaded grey and the conserved Trp 176 and the semi-conserved Gln 182 are shaded red. This figure was prepared using ALSCRIPT (Barton, G. J., 1993, Prot. Engineering, 6:37-40).

Figure 10. Stereo view of a superposition of the L1 (white) and L2 (black) domains. Residues numbers above are for L1 and below for L2. The side chain of Trp 176 which protrudes into the core of L1 is drawn as ball-and-stick.

Figure 11. Schematic diagram showing the association of three β -finger motifs. β -strands are drawn as arrows and disulfide bonds as zigzags.

Figure 12. GRASP [Nicolls, A. et al., 1993, Biophys. J. 64, 166-170] surface diagram of the L1 domain of IGF-1R shown in a similar view to Figure 8. The

N-terminal β -strand is at the top. The mutation L87A [Nakae, J. et al., 1995, J. Biol. Chem. 270, 22017-22022] and four regions (residues 12-15, 34-44, 64-67 and 89-91 of IR) shown to be important in insulin binding to IR [Williams, P. F. et al., 1995, J. Biol. Chem. 270, 3012-3016] correspond to a patch of

5 residues on the large β -sheet. Residues numbers for IR/IGF-1R are given and residues are coloured according to the magnitude of $K_d(\text{mutant})/K_d(\text{wild type})$, red, > 40 ; orange, 10-40; yellow, 2.5-10; green, < 2.5 ; non-secreting, white; untested, blue. All mutants on the opposite face of the domain do not affect insulin affinity.

10

Figure 13: Sequence Alignment of hIGF-1R, hIR and hIRR Ectodomains. Derived by use of the PileUp program in the software package of the Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA. For assignment of homologous 3D structures see Figure 9.

15

Figure 14: Sequence Alignment of EGFR, ErbB2, ErbB3 and ErbB4 Ectodomains. Derived by use of the PileUp program in the software package of the Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA. For alignment on the IGF-1R fragment and assignment of homologous 20 3D structures, see Figure 9.

Figure 15 Sequence Alignment and Classification of the Disulphide-bonded Modules in the Cys-rich domains of IGF-1R, IR, IRR, EGFR, ErbB2, ErbB3 and ErbB4.

25

Figure 16. Gel filtration chromatography of insulin receptor ectodomain and MFab complexes. hIR -11 ectodomain dimer (5 - 20 mg) was complexed with MFab derivatives (15-25 mg each) of the anti-hIR antibodies 18-44, 83-7 and 83-14 (Soos et al., 1986). Elution profiles were generated from samples 30 loaded onto a Superdex S200 column (Pharmacia), connected to a BioLogic chromatography system (Biorad) and monitored at 280 nm. The column was eluted at 0.8 ml/min with 40 mM Tris/150 mM sodium chloride/0.02% sodium azide buffer adjusted to pH 8.0: Profile 0, hIR -11ectodomain, Profile 1, ectodomain mixed with MFab 18-44; Profile 2 , ectodomain mixed with 35 MFab18-44 and MFab 83-14; Profile 3, ectodomain mixed with MFab 18-44, MFab 83-14 and MFab 83-7. The apparent mass of each complex was

determined from a plot of the following standard proteins: thyroglobulin (660 kDa), ferritin (440 kDa), bovine gammaglobulin (158 kDa), bovine serum albumin (67 kDa), chicken ovalbumin (44 kDa) and equine myoglobin (17 kDa).

5

Figure 17. Micrographs of hIR and hIGF-1R ectodomains. (a) Undecorated hIR ectodomain dimer stained with methylamine tungstate showing parallel bars. (b) Undecorated hIR ectodomain dimer stained with uranyl formate, showing well-spaced parallel bars corresponding to the cartoon below. (c) Undecorated hIGF-1R ectodomain dimer stained with uranyl formate. Magnification bars for (a), (b) and (c) 50nm.

10

Figure 18. Micrographs of hIR and hIGF-1R ectodomains. (a) Thinly stained region of undecorated hIR ectodomain dimers in uranyl formate, showing U-shaped particles (circled) as well as parallel bars as in the cartoon below. (b) Undecorated hIGF-1R ectodomain dimer under similar staining conditions. Magnification bars 50 nm.

15

Figure 19. hIR ectodomain dimer complexed with MFab 83-7 and stained with KPT. Three projections can be recognised: circled particles have the Fab arms displaced either clockwise as in the cartoon below left, or anticlockwise as in the cartoon below middle; arrowed particles have the Fab arms in a central position, cartoon below right. Magnification bar 50 nm.

20

Figure 20. hIR ectodomain dimer complexed with MFab 83-7 and stained with uranyl formate showing the parallel bar structure in particles having the Fab arms displaced (circled). Magnification bar 50 nm.

25

Figure 21. (a) hIR ectodomain dimer complexed with MFab 83-14 stained with potassium phosphotungstate, showing Fab arms attached near the bottom of U-shaped particles (circled). The corresponding cartoon is shown below left. (b) hIR ectodomain dimer complexed with MFab 83-14 stained with uranyl acetate, showing both the view described above (circled) and the parallel-bar view with diagonally projecting Fab arms (arrowed), as in the cartoon below right. Magnification bars 50 nm.

30

35

Figure 22 . Double complex of hIR ectodomain dimer with MFabs 83-7 and 18-44 showing particles of complex shape (circled) with four Fab arms attached, consistent with the cartoon below. Magnification bar 50 nm.

- 5 **Figure 23.** Images of hIR ectodomain dimer co-complexed with MFabs 83-7, 83-14 and 18-44 showing examples of complex particles (circled) where it is possible to identify that there are more than four MFabs bound to the dimeric central region. Magnification bar 50 nm.
- 10 **Figure 24.** Schematic illustrating the proposed model of the hIR ectodomain dimer. The dimensions of the molecular envelope are as shown in the diagram, as is the position of the two-fold axis.

Detailed Description of the Invention

- 15 We describe herein the expression, purification, and crystallization of a recombinant IGF-1R fragment (residues 1-462) containing the L1-cysteine-rich-L2 region of the ectodomain. The selected truncation position is just downstream of the exon 6/exon 7 junction (Abbott, A. M., et al., 1992, J Biol Chem., 267:10759-10763) and occurs at a position where the sequences of the
- 20 IR and EGFR families diverge markedly (Ward, C. W., et al., 1995, Proteins: Struct., Funct., Genet. 22:141-153; Lax, I., et al., 1988, Molec. Cellul. Biol. 8:1970-1978) suggesting it represents a domain boundary. To limit the effects of glycosylation, the IGF-1R fragment was expressed in Lec8 cells, a glycosylation mutant of Chinese hamster ovary (CHO) cells, whose defined
- 25 glycosylation defect produces N-linked oligosaccharides truncated at N-acetyl glucosamine residues distal to mannose residues (Stanley, P. 1989, Molec. Cellul. Biol. 9:377-383). Such an approach has facilitated glycoprotein crystallization (Davis, S. J., et al., 1993, Protein Eng. 6:229-232; Liu, J., et al., 1996, J. Biol. Chem. 271:33639-33646).
- 30 The IGF-1R construct described herein included a c-myc peptide tag (Hoogenboom, H. R., et al., 1991, Nucleic Acids Res. 19:4133-4137) that is recognised by the Mab 9E10 (Evan, G. I., et al., 1985, Mol. Cell. Biol. 5:3610-3616) enabling the expressed product to be purified by peptide elution from an antibody affinity column followed by gel filtration over Superdex S200.
- 35 The purified proteins crystallized under a sparse matrix screen (Jancarik, J. & Kim, S.-H., 1991, J. Appl. Cryst. 24:409-411) but the crystals were of variable

quality, with the best diffracting to 3.0-3.5Å. Isocratic gradient elution by anion-exchange chromatography yielded protein that was less heterogenous and gave crystals of sufficient quality to determine the structure of the first three domains of the human IGF-1R.

5 The IGF-1R fragment consisted of residues 1-462 of IGF-1R linked via an enterokinase-cleavable pentapeptide sequence to an eleven residue c-myc peptide tag at the C-terminal end. The fragment was expressed in Lec8 cells by continuous media perfusion in a bioreactor using porous carrier disks. It was secreted into the culture medium and purified by peptide elution from
10 an anti-c-myc antibody column followed by Superdex S200 gel filtration. The receptor fragment bound two anti-IGF-1R monoclonal antibodies, 24-31 and 24-60, which recognize conformational epitopes, but could not be shown to bind IGF-1 or IGF-2. Crystals of variable quality were grown as rhombic
15 prisms in 1.7 M ammonium sulfate at pH 7.5 with the best diffracting to 3.0-3.5 Å. Further purification by isocratic elution on an anion-exchange column gave protein which produced better quality crystals, diffracting to 2.6 Å, that were suitable for X-ray structure determination.

 The structure of this fragment (IGF-1R residues 1-462; L1-cys rich-L2domains) has been determined to 2.6 Å resolution by X-ray diffraction. The
20 L domains each adopt a compact shape consisting of a single stranded right-handed β-helix. The cys-rich region is composed of eight disulphide-bonded modules, seven of which form a rod-shaped domain with modules associated in a novel manner. At the centre of this reasonably extended structure is a space, bounded by all three domains, and of sufficient size to accommodate a
25 ligand molecule. Functional studies on IGF-1R and other members of the insulin receptor family show that the regions primarily responsible for hormone-binding map to this central site. Thus this structure gives a first view of how members of the insulin receptor family might interact with their ligands.

30 Another group has reported the crystallization of a related receptor, the EGFR in a complex with its ligand EGF (Weber, W., et al., 1994, J Chromat. 679:181-189). However difficulties were encountered with these crystals which diffracted to only 6 Å, insufficient for the determination of an atomic resolution structure of this complex (Weber, W., et al., 1994, J
35 Chromat 679:181-189) or the generation of accurate models of structurally related receptor domains such as IGF-1R and IR by homology modelling.

The present inventors have applied the same process to the IR and generated a fragment (residues 1-485) that covers the first three domains of the IR. This fragment has been expressed in transformed Lec8 cells, purified, and crystallized by similar methodologies to yield crystals suitable for X-ray diffraction.

The present inventors have therefore developed 3D structural information about cytokine receptors to enable a more accurate understanding of how the binding of ligand leads to signal transduction. Such information provides a rational basis for the development of antagonists or agonists for specific therapeutic applications, something that heretofore could not have been predicted *de novo* from available sequence data.

The precise mechanisms underlying the binding of agonists and antagonists to the IGF-1 receptor site are not fully clarified. However, the binding of the agonists or antagonists to the receptor site, preferably with an affinity in the order of 10^{-8} M or higher, is understood to arise from enhanced stereochemical complementarity, relative to naturally occurring IGF-1 ligands.

Such stereochemical complementarity, pursuant to the present invention, is characteristic of a molecule that matches intra-site surface residues lining the groove of the receptor site as enumerated by the coordinates set out in Figure 1. The residues lining the groove are depicted in Figure 2. Substances which are complementary to the shape of the receptor site characterised by amino acids positioned at atomic coordinates set out in Figure 1 may be able to bind to the receptor site and, when the binding is sufficiently strong, substantially prohibit binding of the naturally occurring ligands to the site.

It will be appreciated that it is not necessary that the complementarity between agonists or antagonists and the receptor site extend over all residues lining the groove in order to inhibit binding of the natural ligand. Accordingly, agonists or antagonists which bind to a portion of the residues lining the groove are encompassed by the present invention.

In general, the design of a molecule possessing stereochemical complementarity can be accomplished by means of techniques that optimize, either chemically or geometrically, the "fit" between a molecule and a target receptor. Known techniques of this sort are reviewed by Sheridan and Venkataraghavan, *Acc. Chem. Res.* 1987 20 322; Goodford, *J. Med. Chem.*

1984 27 557; Beddell, Chem. Soc. Reviews 1985, 279; Hol, Angew. Chem. 1986 25 767 and Verlinde C.L.M.J & Hol, W.G.J. Structure 1994, 2, 577, the respective contents of which are hereby incorporated by reference. See also ~~Blundell et al., Nature 1987 326 347 (drug development based on information~~
5 regarding receptor structure).

Thus, there are two preferred approaches to designing a molecule, according to the present invention, that complements the shape of IGF-1R or a related receptor molecule. By the geometric approach, the number of internal degrees of freedom (and the corresponding local minima in the
10 molecular conformation space) is reduced by considering only the geometric (hard-sphere) interactions of two rigid bodies, where one body (the active site) contains "pockets" or "grooves" that form binding sites for the second body (the complementing molecule, as ligand). The second preferred approach entails an assessment of the interaction of respective chemical
15 groups ("probes") with the active site at sample positions within and around the site, resulting in an array of energy values from which three-dimensional contour surfaces at selected energy levels can be generated.

The geometric approach is illustrated by Kuntz et al., J. Mol. Biol. 1982 161 269, the contents of which are hereby incorporated by reference,
20 whose algorithm for ligand design is implemented in a commercial software package distributed by the Regents of the University of California and further described in a document, provided by the distributor, which is entitled "Overview of the DOCK Package, Version 1.0," the contents of which are hereby incorporated by reference. Pursuant to the Kuntz algorithm, the
25 shape of the cavity represented by the IGF-R1 site is defined as a series of overlapping spheres of different radii. One or more extant data bases of crystallographic data, such as the Cambridge Structural Database System maintained by Cambridge University (University Chemical Laboratory, Lensfield Road, Cambridge CB2 1EW, U.K.) and the Protein Data Bank
30 maintained by Brookhaven National Laboratory (Chemistry Dept. Upton, NY 11973, U.S.A.), is then searched for molecules which approximate the shape thus defined.

Molecules identified in this way, on the basis of geometric parameters, can then be modified to satisfy criteria associated with chemical
35 complementarity, such as hydrogen bonding, ionic interactions and Van der Waals interactions.

The chemical-probe approach to ligand design is described, for example, by Goodford, J. Med. Chem. 1985 28 849, the contents of which are hereby incorporated by reference, and is implemented in several commercial software packages, such as GRID (product of Molecular Discovery Ltd., West

- 5 Way House, Elms Parade, Oxford OX2 9LL, U.K.). pursuant to this approach, the chemical prerequisites for a site-complementing molecule are identified at the outset, by probing the active site (as represented via the atomic coordinates shown in Fig. 1) with different chemical probes, e.g., water, a methyl group, an amine nitrogen, a carboxyl oxygen, and a hydroxyl.
- 10 Favored sites for interaction between the active site and each probe are thus determined, and from the resulting three-dimensional pattern of such sites a putative complementary molecule can be generated.

- The chemical-probe approach is especially useful in defining variants of a molecule known to bind the target receptor. Accordingly,
- 15 crystallographic analysis of IGF-1 bound to the receptor site may provide useful information regarding the interaction between the archetype ligand and the active site of interest.

- In summary, the general principles of receptor-based drug design can be applied by persons skilled in the art, using the crystallographic results presented above, to produce agonists or antagonists of IGF-1R having
- 20 sufficient stereochemical complementarity to exhibit high affinity binding to the receptor site.

The present invention is further described below with reference to the following, non-limiting examples.

25

EXAMPLE 1

Expression, Purification and Crystalization of the IGF-1R Fragment.

- Several factors hamper macromolecular crystallization including sample selection, purity, stability, solubility (McPherson, A., et al., 1995, Structure 3:759-768); Gilliland, G. L., & Ladner, J. E., 1996, Curr. Opin. Struct. Biol. 6:595-603), and the nature and extent of glycosylation (Davis, S. J., et al., 1993, Protein Eng. 6:229-232). Initial attempts to obtain structural data from soluble IGF-1R ectodomain (residues 1-906) protein, expressed in Lec8 cells (Stanley, P. 1989, Molec. Cellul. Biol. 9:377-383) and purified by
- 30 affinity chromatography, produced large, well-formed crystals (1.0 mm x 0.2 mm x 0.2 mm) which gave no discernable X-ray diffraction pattern
- 35

(unpublished data). Similar difficulties have been encountered with crystals of the structurally related epidermal growth factor receptor (EGFR) ectodomain which diffracted to only 6 Å, insufficient for the determination of an atomic resolution structure (Weber, W. et al., 1994, *J Chromat* 679:181-

5 189). This prompted us to search for a fragment of IGF-1R that was more amenable to X-ray crystallographic studies.

The fragment expressed (residues 1-462) comprises the L1-cysteine-rich-L2 region of the ectodomain. The selected truncation position at Val462 is four residues downstream of the exon 6/exon 7 junction (Abbott, A. M., et al., 1992, *J Biol Chem.* 267:10759-10763) and occurs at a position where the sequences of the IR and the structurally related EGFR families diverge markedly (Lax, I., et al., 1988, *Molec Cell Biol.* 8:1970-1978; Ward, C. W., et al., 1995, *Proteins: Struct., Funct., Genet.* 22:141-153), suggesting it represents a domain boundary. The expression strategy included use of the pEE14 vector (Bebbington, C. R. & Hentschel, C. C. G., 1987, In: Glover, D. M., ed. *DNA Cloning*. Academic Press, San Diego. Vol 3, p163) in glycosidase-defective Lec8 cells (Stanley, P., 1989, *Molec. Cellul. Biol.* 9:377-383), which produce N-linked oligosaccharides lacking the terminal galactose and N-acetylneuraminic acid residues (Davis, S. J., et al., 1993, *Protein Eng.* 6:229-232; Liu, T., et al., 1996, *J Biol Chem* 271:33639-33646.). The construct contained a C-terminal c-myc affinity tag (Hoogenboom, H. R., et al., 1991, *Nucl Acids Res.* 19:4133-4137), which facilitated immunoaffinity purification by specific peptide elution and avoided aggressive purification conditions. These procedures yielded protein which readily crystallized after a gel filtration polish. This provided a general protocol to enhance crystallisation prospects for labile, multidomain glycoproteins.

The structure of this fragment is of considerable interest since it contains the major determinants governing insulin and IGF-1 binding specificity (Gustafson, T. A. & Rutter, W. J., 1990, *J. Biol. Chem.* 265:18663-18667; Andersen, A. S., et al., 1990, *Biochemistry*, 29:7363-7366; Schumacher, R., et al., 1991, *J. Biol. Chem.* 266:19288-19295; Schumacher, R., et al., 1993, *J. Biol. Chem.* 268:1087-1094; Schäffer, L., et al., 1993, *J. Biol. Chem.* 268:3044-3047; Williams, P. F., et al., 1995, *J. Biol. Chem.* 270:3012-3016) and is very similar to an IGF-1R fragment (residues 1-486) reported to act as a strong dominant negative for several growth functions and which

induces apoptosis of tumour cells in vivo (D'Ambrosio, C., et al., 1996, Cancer Res. 56:4013-4020).

The expression plasmid pEE14/IGF-1R/462 was constructed by inserting the oligonucleotide cassette:

5

AatII

5' GACGTC GACGATGACGATAAG GAACAAAACTCATC

D V D D D D K E Q K L I

(EK cleavage)

(c-myc tail)

10 S E E D L N (Stop)

TCAGAAGAGGATCTGAAT TAGAATTC GACGTC 3'

EcoRI AatII

encoding an enterokinase cleavage site, c-myc epitope tag (Hoogenboom, H. R., et al., 1991, Nucleic acids Res. 19:4133-4137) and stop codon into the *AatII* site (within codon 462) of IGF-1 receptor cDNA in the mammalian expression vector pECE (Ebina, Y., et al., 1985, Cell, 40:747-758; kindly supplied by W. J. Rutter, UCSF, USA), and introducing the DNA comprising the 5' 1521 bp of the cDNA (Ullrich, A., et al., 1986, EMBO J. 5:2503-2512) ligated to the oligonucleotide cassette into the *EcoRI* site of the mammalian plasmid expression vector pEE14 (Bebbington, C. R. & Hentschel, C. C. G., 1987, In: Glover, D. M., ed. DNA Cloning. Academic Press, San Diego. Vol 3, p163; Celltech Ltd., UK). Plasmid pEE14/IGF-1R/462 was transfected into Lec8 mutant CHO cells (Stanley, P. 1989, Molec. Cellul. Biol. 9:377-383) obtained from the American Tissue Culture Collection (CRL:1737) using Lipofectin (Gibco-BRL). Cell lines were maintained after transfection in glutamine-free medium (Glasgow modification of Eagle's medium (GMEM; ICN Biomedicals, Australia) and 10% dialysed FCS (Sigma, Australia) containing 25 µM methionine sulfoximine (MSX; Sigma, Australia) as described (Bebbington, C. R. & Hentschel, C. C. G., 1987, In: Glover, D. M., ed. DNA Cloning. Academic Press, San Diego. Vol 3, p163). Transfectants were screened for protein expression by Western blotting and sandwich enzyme-linked immunosorbant assay (ELISA) (Cosgrove, L., et al., 1995,) using monoclonal antibody (Mab) 9E10 (Evan et al., 1985) as the capture antibody and either biotinylated anti-IGF-1R Mab 24-60 or 24-31 for detection (Soos et al., 1992; gifts from Ken Siddle, University of Cambridge,

UK). Large-scale cultivation of selected clones expressing IGF-1R/462 was carried out in a Celligen Plus bioreactor (New Brunswick Scientific, USA) containing 70 g Fibra-Cel Disks (Sterilin, UK) as carriers in a 1.25 L working volume. Continuous perfusion culture using GMEM medium supplemented

5 with non-essential amino acids, nucleosides, 25 μ M MSX and 10% FCS was maintained for 1 to 2 weeks followed by the more enriched DMEM/F12 without glutamine, with the same supplementation for the next 4-5 weeks. The fermentation production run was carried out three times under similar conditions and resulted in an estimated overall yield of 50 mg of receptor
10 protein from 430 L of harvested medium. Cell growth was poor during the initial stages of the fermentation when GMEM medium was employed, but improved dramatically following the switch to the more enriched medium. Target protein productivity was essentially constant during the period from ~100 to 700 h of the 760 h fermentation, as measured by ELISA using Mab
15 9E10 as the capture antibody and biotinylated Mab 24-31 as the developing antibody.

Soluble IGF-1R/462 protein was recovered from harvested fermentation medium by affinity chromatography on columns prepared by coupling Mab 9E10 to divinyl sulphone-activated agarose beads (Mini Leak;
20 Kem En Tec, Denmark) as recommended by the manufacturer. Mini-Leak Low and Medium affinity columns with antibody loadings of 1.5-4.5 mg/ml of hydrated matrix were obtained, with the loading range of 2.5-3 mg/ml giving optimal performance (data not shown). Mab 9E10 was produced by growing hybridoma cells (American Tissue Culture Collection) in serum-free medium
25 in the Celligen Plus bioreactor and recovering the secreted antibody (4 g) using protein A glass beads (Prosep-A, Bioprocessing Limited, USA). Harvested culture medium containing IGF-1R/462 protein was adjusted to pH 8.0 with Tris-HCl (Sigma), made 0.02% (w/v) in sodium azide and passed at 3-5 ml/min over 50 ml Mab 9E10 antibody columns at 4° C. Bound protein
30 was recovered by recycling a solution of 2-10 mg of the undecamer c-myc peptide EQKLISEEDLN (Hoogenboom et al., 1991) in 20 ml of Tris-buffered saline containing 0.02% sodium azide (TBSA). Between 65% and 75% of the product was recovered from the medium as estimated by ELISA, with a further 15-25% being recovered by a second pass over the columns. Peptide
35 recirculation (~10 times) through the column eluted bound protein more efficiently than a single, slower elution. Residual bound protein was eluted

with sodium citrate buffer at pH 3.0 into 1 M Tris HCl pH 8.0 to neutralize the eluant, and columns were re-equilibrated with TBSA.

Gel filtration over Superdex S200 (Pharmacia, Sweden), of affinity-purified material showed a dominant protein peak at ~ 63 kDa, together with a smaller quantity of aggregated protein (Figure 3a). The peak protein migrated primarily as two closely spaced bands on reduced, sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE; Figure 3b), reacted positively in the ELISA with both Mab 24-60 and Mab 24-31, and gave a single sequence corresponding to the N-terminal 14 residues of IGF-1R. No binding of IGF-1 or IGF-2 could be detected in the solid plate binding assay (Cosgrove et al., 1995, Protein Express Purif. 6:789-798). The IGF-1R/462 fragment was further purified by ion-exchange chromatography on Resource Q (Pharmacia, Sweden). Using shallow salt gradients, protein enriched in the slowest migrating SDS-PAGE band was obtained (data not shown), which formed relatively large, well-formed crystals (see below). Isoelectric focusing showed the presence of one major and two minor isoforms. Protein purified on Resource Q with an isocratic elution step of 0.14 M NaCl in 20 mM TrisCl at pH 8.0 (fraction 2, Figure 4) showed less heterogeneity on isoelectric focusing (Figure 4 inset) and SDS-PAGE (data not shown) and produced crystals of sufficient quality for structure determination (see below).

Crystals were grown by the hanging drop vapour diffusion method using purified protein concentrated in Centricon 10 concentrators (Amicon Inc, USA) to 5-10 mg/ml in 10-20 mM Tris-HCl pH 8.0 and 0.02% (w/v) azide, or 100 mM ammonium sulfate and 0.02% (w/v) azide. A search for crystallization conditions was performed initially using the factorial screen (Jancarik, J. & Kim, S.-H., 1991, J Appl Cryst 24:409-411) and subsequently optimised. Crystals were examined on an M18XHF rotating anode generator (Siemens, Germany) equipped with Franks mirrors (MSC, USA) and RAXIS IIC and IV image plate detectors (Rigaku, Japan).

From the initial crystallization screen of this protein, crystals of about 0.1 mm in size grew in one week. Upon refining conditions, crystals of up to 0.6 x 0.4 x 0.4 mm could be grown from a solution of 1.7-2.0 M ammonium sulfate, 0.1 M HEPES pH 7.5. The crystals varied considerably in shape and diffraction quality, growing predominantly as rhombic prisms with a length to width ratio of up to 5:1, but sometimes as rhombic bipyramids, the latter form being favoured when using material which had been eluted

from the Mab 9E10 column at pH 3.0. Each crystal showed a minor imperfection in the form of very faint lines from the centre to the vertices. Protein from dissolved crystals did not appear to be different from the protein stock solution when run on an isoelectric focusing gel. Upon X-ray

5 examination, the crystals diffracted to 3.0-4.0 Å and were found to belong to the space group $P2_12_12_1$ with $a = 76.8$ Å, $b = 99.0$ Å, $c = 119.6$ Å. In the diffraction pattern, the crystal variability noted above was manifest as a large (1-2°) and anisotropic mosaic spread, with concomitant variation in resolution. To improve the quality of the crystals, they were grown in the
10 presence of various additives or were recrystallized. These methods failed to substantially improve the crystal quality although bigger crystals were obtained by recrystallization. The variability in crystal quality appeared to be due to protein heterogeneity, as demonstrated by the observation that more highly purified protein, eluted isocratically from the Resource Q column and
15 showing one major band on isoelectric focusing (Figure 4 inset), produced crystals of sufficient quality for structure determination. These crystals diffracted to 2.6 Å resolution with cell dimensions, $a = 77.0$ Å, $b = 99.5$ Å, $c = 120.1$ Å and mosaic spread of 0.5°. Heavy metal derivatives of the IGF-1R/462 crystals have been obtained and are leading to the determination of
20 an atomic resolution structure of this fragment, which contains the L1, cysteine-rich and L2 domains of human IGF-1R.

EXAMPLE 2

Expression, Purification and Crystalization of the IR Fragment

A similar strategy was adopted for the human insulin receptor. The
25 fragment expressed (residues 1-485) comprises the L1-cysteine-rich-L2 region of the IR ectodomain but extends 13 residues further before the attachment of the 17 residue EK cleavage site linker and c-myc tail. The selected truncation position corresponds to a unique and convenient Bgl II restriction site. The expression strategy was also based on the pEE14 expression vector in
30 glycosidase-defective Lec8 cells and use of a C-terminal c-myc affinity tag for immunoaffinity purification by specific peptide elution. These procedures yielded IR protein which readily crystallized after a gel filtration polish.

The expression plasmid pHIR485 was constructed by ligating the double-stranded oligonucleotide cassette:

Bgl II*Xba* I

5' AGATC TCCGACGATGACGATAAG GAACAAAACTCATCTCAGAAGAGGATCTGAAT TAG TCTAGA 3'

K I S D D D D K E Q K L I S E E D L N

EK cleavage

c-myc tail

Stop

5

- encoding an enterokinase cleavage site, c-myc epitope tag (Hoogenboom, H. R., et al., 1991, Nucleic acids Res. 19:4133-4137) and stop codon , to the larger 11.1 kilobasepair *Bgl* II / *Xba* I fragment isolated from digestion of the mammalian expression plasmid pEH3 (a derivative of the mammalian
- 10 plasmid expression vector pEE14 [Bebbington, C. R. & Hentschel, C. C. G., 1987, In: Glover, D. M., ed. DNA Cloning. Academic Press, San Diego. Vol 3, p163; Celltech Ltd., UK] which holds the entire coding sequence of human insulin receptor within a Hind III /*Xba* I fragment). Lec8 mutant CHO cells (Stanley, P. 1989, Molec. Cellul. Biol. 9:377-383) obtained from the American
- 15 Tissue Culture Collection (CRL:1737) were transfected with pHIR485 using Lipofectamine (Gibco-BRL). Cell lines were maintained after transfection in glutamine-free medium (Glasgow modification of Eagle's medium - GMEM; ICN Biomedicals, Australia) and 10% dialysed FCS (Sigma, Australia) containing 25 μ M methionine sulfoximine (MSX; Sigma, Australia) as
- 20 described (Bebbington, C. R. & Hentschel, C. C. G., 1987, In: Glover, D. M., ed. DNA Cloning. Academic Press, San Diego. Vol 3, p163). Transfectants were screened for protein expression by Western blotting and sandwich enzyme-linked immunosorbant assay (ELISA) (Cosgrove, L., et al., 1995,) using anti-hIR (Mab) 83.7 as the primary antibody and biotinylated
- 25 monoclonal antibody (Mab) 9E10 (Evan et al., 1985) for detection (Soos et al., 1986; gifts from Ken Siddle, University of Cambridge, UK). Large-scale cultivation of selected clones expressing IR/485 was carried out in a Celligen Plus bioreactor (New Brunswick Scientific, USA) containing 70 g Fibra-Cel Disks (Sterilin, UK) as carriers in a 1.25 L working volume.
- 30 Continuous perfusion culture was carried out using DMEM/F12 without glutamine medium (ICN), supplemented with non-essential amino acids, nucleosides, 25 μ M MSX and 5 - 10% FCS and resulted in an estimated overall yield of 115 mg of receptor protein from 165 L of harvested medium. Target protein productivity was essentially constant during the fermentation,
- 35 as measured by ELISA.

Soluble IR/485 protein was recovered from harvested fermentation medium by affinity chromatography on columns of Mab 9E10 essentially as described in Example 1. Between 92 -98% of the product was recovered from the medium by this affinity chromatography step, as estimated by ELISA.

5 Gel filtration over Superdex 200 (Pharmacia, Sweden), of the affinity-purified material at 1mg/ml produced a dominant protein peak at apparent mass ~140 kDa (Figure 5a - interpreted as dimer), whereas a peak at apparent mass ~85 kDa was obtained (Figure 5b - interpreted as monomer) at 0.02 mg/ml. The protein migrated as a single broad band of apparent molecular mass ~78 kDa (reduced- lane A) or ~68 kDa (non-reduced - lane B) on sodium dodecyl sulfate polyacrylamide gel electrophoresis (SDS-PAGE; Figure 6a) The IR/485 fragment reacted positively in the ELISA with Mab 83-7, gave a single sequence corresponding to the N-terminal 10 residues of IR, showing several isoforms on isoelectric focussing from pI 6.0 - 6.8 (Figure 6b). Crystallisation screening trials of the fragment produced crystals too small for X-ray diffraction studies. The fragment was further purified by ion-exchange chromatography on Uno Q (BioRad, USA), using stepwise isocratic elution with incremental changes in salt concentrations (Figure 7). Fractions A and D were each enriched in a component isoform from the ladder of isoforms present in the unfractionated mixture (Figure 6b). Both these fractions produced crystals, whereas no crystals were obtained from fractions B and C.

Crystals were grown by the hanging drop vapour diffusion method using purified protein concentrated in Centricon 10 concentrators (Amicon Inc, USA) to 5-10 mg/ml in 10mM Tris-HCl pH 8.0 and 0.02% (w/v) azide. A search for crystallization conditions was performed initially using the factorial screen (Jancarik, J. & Kim, S.-H.,1991, J Appl Cryst 24:409-411) and subsequently optimised. Crystals were examined on an M18XHF rotating anode generator (Siemens, Germany) equipped with Franks mirrors (MSC, USA) and an RAXIS IIC image plate detector (Rigaku, Japan).

From the initial crystallization screen of this protein fraction D fine needles grew in about one week. In further experiments, crystals of up to 0.04 x 0.04 x 0.2 mm could be grown from a solution of 1.9-2.0 M ammonium sulfate, 2% PEG 400, 0.1 M HEPES pH 7.5. Upon X-ray examination, the crystals diffracted to 4 Å and were found to belong to the space group $P2_12_12_1$ with $a = 103.2 \text{ Å}$, $b = 130.0 \text{ Å}$, $c = 161.6 \text{ Å}$. Despite their small size these

crystals diffracted sufficiently well to allow collection of a low resolution data set. Further purification of the protein and refinement of crystallisation conditions should yield larger crystals, providing data to determine the structure of this fragment at medium resolution or better.

5 **EXAMPLE 3**

Structure of the IGF-1R/1-462

Crystals were cryo-cooled to -170°C in a mother liquor containing 20% glycerol, 2.2 M ammonium sulfate and 100 mM Tris at pH 8.0. Native and derivative diffraction data were recorded on Rigaku RAXIS IIC or IV area
10 detectors using copper K α radiation from a Siemens rotating anode generator with Yale/MSU mirror optics. The space group was P2₁2₁2₁ with a = 77.39 Å, b = 99.72 Å, and c = 120.29 Å. Data were reduced using DENZO and SCALEPACK (Otwinowski, Z. & Minor, W., 1996, *Methods in Enzymology* 276:307-326). Diffraction was notably anisotropic for all crystals examined.

15 Phasing by multiple isomorphous replacement (MIR) was performed with PROTEIN (Steigeman, W. Dissertation (Technical Univ. Munich, 1974) using anomalous scattering for both UO₂ and PIP derivatives. Statistics for data collection and phasing are given in Table 1. In the initial MIR map regions of protein and solvent could clearly be seen but the path of the
20 polypeptide was by no means obvious. That map was subject to solvent flattening and histogram matching in DM (Cowtan, K., 1994, *Joint CCP4 and ESF-EACBM newsletter* Protein Crystallogr. 31:34-38). The structure was traced and rebuilt using O (Jones, T. A., et al., 1991, *Acta Crystallogr.* A47:110-119) and refined with X-PLOR 3.851 (Brunger, A. T., 1996, *X-PLOR Reference Manual* 3.851, Yale Univ., New Haven, CT). After 5 rounds of
25 rebuilding and energy minimisation the R-factor dropped to 0.279 and R_{free} = 0.359 for data 7-2.6 Å resolution. The current model contains 458 amino acids and 3 N-linked carbohydrates but no solvent molecules. For residues with B(Ca) > 70 Å² atomic positions are less reliable (37-42, 155-159, 305,
30 336-341, 404-406, 453-458). There is weak electron density for residues 459-461 but the c-myc tail appears completely disordered.

The 1-462 fragment consists of the N-terminal three domains of IGF-1R (L1, cys-rich, L2) and contains regions of the molecule which dictate
35 ligand specificity (17-23). The molecule adopts a reasonably extended structure (approximately 40 x 48 x 105 Å) with domain 2 (cys-rich region) making contact along the length of domain 1 (L1) but very little contact with

the third domain (L2) (see Figure 8). This leaves a space at the centre of the molecule of approximately $24 \text{ \AA} \times 24 \text{ \AA} \times 24 \text{ \AA}$ which is bounded on three sides by the three domains of the molecule. The space is of sufficient size to accommodate the ligand, IGF-1.

5 The L domains

Each of the L domains (residues 1-150 and 300-460) adopt a compact shape ($24 \times 32 \times 37 \text{ \AA}$) consisting of a single-stranded right handed β -helix and capped on the ends by short α -helices and disulfide bonds. The body of the domain looks like a loaf of bread with the base formed from a flat six-stranded β -sheet, 5 residues long and the sides being β -sheets three residues long (Figures 8 & 9). The top is irregular but in places is similar for the two domains. The two domains are superposable with an rms deviation in Ca positions of 1.6 \AA for 109 atoms (Figure 10). Although this fold is reminiscent of other β -helix proteins it is much simpler and smaller with very few elaborations and thus it represents a new superfamily of domains. One notable difference between the two domains is that the indole ring of Trp 176 from the cys-rich region (Figure 9b) is inserted into the hydrophobic core of L1 and the C-terminal helix is only vestigial (Figure 8). For the insulin receptor family the sequence motif of residues which form the Trp pocket in L1 does not occur in L2 (Figure 9a). However in the EGF receptor, which has an additional cys-rich region after the L2 domain (14, 15), the pocket motif can be found in both L domains and the Trp is conserved in both cys-rich regions (Figure 9b).

The repetitive nature of the β -helix is reflected in the sequence and the first five turns were correctly identified by Bajaj, M., et al. (1987, *Biochim.Biophys. Acta* 916:220-226), the conserved Gly residues being found in turns making one bottom edge of the domain. However, their conclusions about the fold were incorrect. The "helix-like" repeat is actually a pair of bends at the top edge of the domain. In their Motif V, the Gly is not in a bend but is followed by the insertion of a conserved loop of 7-8 residues (see Figure 9a). Glycine is structurally important in the Gly bends as mutation of these residues compromises folding of the receptor [van der Vorm, E.R., et al., 1992, *J. Biol. Chem.* 267, 66-71; Wertheimer, E. et al., 1994, *J. Biol. Chem.* 269, 7587-7592].

Upon comparing the L domains with other right-handed β -helix structures such as pectate lyase (Yoder, M. D., et al., 1993, *Structure*, 1:241-

251-1507) and the p22 tailspike protein (Steinbacher, S., et al., 1997, J.Mol. Biol. 267:865-880) there are some striking similarities as well as differences. In all cases the ends of the domain are capped by α -helices but the L domains also have a disulphide bond at each end to hold the termini. The other β -

5 helix domains are considerably longer and have significant twist to their sheets while the L domains have flat sheets. Although the sizes of the helix repeats are similar (here 24-25 residues vs 22-23 for pectate lyase) the cross-sections are quite different. The L domains have a rectangular cross-section while pectate lyase and p22 tailspike protein are V-shaped and have many,
10 and sometimes quite large, insertions (Yoder, M. D., et al., 1993, Structure, 1:241-251-1507; Steinbacher, S., et al., 1997, J.Mol. Biol. 267:865-880). In the hydrophobic core a common feature is the stacking of aliphatic residues from successive turns of the β -helix and near the C-terminus of each L domain there is also a short Asn ladder, reminiscent of the long Asn ladder
15 observed in pectate lyase (Yoder, M. D., et al., 1993, Structure 1:241-251-1507). On the opposite side of the L domains the Gly bend as well as the two bends and sheet preceding it have no counterpart in the other β -helix domains. Thus although the L domains are built on similar principles to the other β -helix domains they constitute a separate superfamily.

20 **The cys-rich domain**

The cys-rich domain is composed of eight disulfide-bonded modules (Figure 9b), the first of which sits at the end of L1 while the remainder make a curved rod running diagonally across L1 and reaching to L2 (Figure 8). The strands in modules 2-7 run roughly perpendicular to the axis of the rod in a
25 manner more akin to laminin (Stetefeld, J., et al., 1996, J.Mol.Biol. 257:644-657) than to TNF receptor (Banner, D. W., et al., 1993, Cell, 73:431-445) but the modular arrangement of the cys-rich domain is different to other cys-rich proteins for which structures are known. The first 3 modules of IGF-1R have a common core, containing a pair of disulfide bonds, but show considerable
30 variation in the loops (Figure 9b). The connectivity of these modules is the same as the first half of EGF (Cys 1-3 and 2-4) but their structures do not appear to be closely related to any member of the EGF family. Modules 4 to 7 have a different motif, β -finger, and best match residues 2152-2168 of fibrillin (Dowling, A. K., et al., 1996, Cell, 85:597-605). Each is composed of
35 three polypeptide strands, the first and third being disulfide bonded and the latter two forming a β -ribbon. The β -ribbon of each β -finger module lines up

antiparallel to form a tightly twisted 8-stranded β -sheet (Figures 8 and 11). Module 6 deviates from the common pattern with the first segment being replaced by an α -helix followed by a large loop that is likely to have a role in ligand binding (see below). As module 5 is most similar to module 7 it is

5 possible that the four modules arose from serial gene duplications. The final module is a disulfide linked bend of five residues.

The fact that the two major types of cys-rich modules occur separately implies that these are the minimal building blocks of cys-rich domains found in many proteins. Although it can be as short as 16 residues,
 10 the motif of modules 4-7 is clearly distinct and capable of forming a regular extended structure. Thus cys-rich domains such as these can be considered as made of repeat units each composed of a small number of modules.

Hormone binding

Attempts have been made to locate the IGF-1 (and insulin) binding
 15 site by examining natural (Taylor, S. I., 1992, Diabetes, 41:1473-1490) and site-directed mutants (Williams, P. F., et al., 1995, J. Biol. Chem. 270:3012-3016; Mynarcik, D. C et al., 1996, J. Biol. Chem. 271:2439-2442; Mynarcik, D. C., et al., 1997, J. Biol. Chem. 272:2077-2081), chimeric receptors (Andersen, A. S., et al., 1990, Biochemistry 29:7363-7366; Gustafson, T. A., & Rutter, W.
 20 J., 1990, J. Biol. Chem. 265:18663-18667; Schäffer, L., et al., 1993, J. Biol. Chem. 268:3044-3047; Schumacher, R., 1993, J. Biol. Chem. 268:1087-1094; Kjeldsen, T., et al., 1991, Proc. Natl Acad. Sci. USA, 88:4404-4408) and by crosslinking studies (Wedekind, F., et al., 1989, Biol. Chem Hoppe-Seyler, 370:251-258; Fabry, M., 1992, J. Biol. Chem. 267:8950-8956; Waugh, S. M., et
 25 al., 1989, Biochemistry, 28:3448-3458; Kurose, T., et al., 1994), J. Biol. Chem. 269:29190-29197-34). IGF-1R/IR chimeras not only show which regions of the receptors account for ligand specificity but also provide an efficient means of identifying some parts of the hormone binding site. Paradoxically regions controlling specificity are not the same for insulin and
 30 IGF-1. Replacing the first 68 residues of IGF-1R with those of IR confers insulin binding ability on the chimeric IGF-1R (Kjeldsen, T., et al., 1991, Proc. Natl Acad. Sci. USA, 88:4404-4408) and replacing residues 198-300 in the cys-rich region of IR with the corresponding residues 191-290 of IGF-1R allows the chimeric receptor to bind IGF-1 (Schäffer, L., et al., 1993, J. Biol.
 35 Chem. 268:3044-3047). Thus a receptor can be constructed which binds both

IGF-1 and insulin with near native affinity. From the structure it is clear that if the hormone bound in the central space it could contact both these regions.

From analysis a series of chimeras examined by Gustafson, T. A., & Rutter, W. J. (*J. Biol. Chem.* 265:18663-18667, 1990) the specificity

- 5 determinant in the cys-rich region can be limited further to residues 223-274. This region corresponds to modules 4-6 and includes a large and somewhat mobile loop (residues 255-263, mean B[Ca atoms] = 57 Å²) which extends into the central space (see Figure 8). In IR this loop is four residues bigger and is stabilised by an additional disulfide bond (Schäffer, L. & Hansen, 10 P.H., 1996, *Exp. Clin. Endocrinol. Diabetes*, 104: Suppl. 2, 89). The larger loop of IR may serve to exclude IGF-1 from the hormone binding site but allow the smaller insulin molecule to bind. It is interesting to note that mosquito IR homologue, which has a loop two residues larger than the mammalian IRs, also appears to bind insulin but not IGF-1 (Graf, R., et al., 15 1997, *Insect Molec. Biol.* 6:151-163). Analysis of the structure indicates that the insulin/IGF-1 specificity is controlled by residues in this loop (amino acids 253-272 in IGF-1R; amino acids 260-283 in IR)

- As chimeras only address residues which differ between the two receptors a more precise analysis of the site can be obtained from single site 20 mutants. In particular, from an alanine-replacement study, four regions of L1 important for insulin binding were identified (Williams, P. F., et al., 1995, *J. Biol. Chem.* 270:3012-3016). The first three are at similar positions on successive turns of the b-helix and the fourth lies on the conserved bulge on the large b-sheet (Figure 12). Thus there is a footprint for insulin binding to 25 the L1 domain which lies on the first half of large b-sheet facing into the central space. Residues further along the sheet which are conserved in IGF-1R and could also be important. The conservative substitution of leucine for methionine at residue 119 of IR (113 of IGF-1R) causes a mild form of leprechaunism [Hone, J. et al., 1994, *J. Med. Genet.* 31, 715-716]. This 30 residue is buried and the mutation could perturb neighbouring residues to affect insulin binding.

- The axis of the L2 domain is perpendicular to that of the L1 domain and N-terminal end of its β -helix is presented to the hormone-binding site. On this face of the L2 domain the only mutation studied so far is the 35 naturally occurring IR mutant, S323L, which gives rise to Rabson-Mendenhall syndrome and severe insulin resistance (Roach, P., 1994, *Diabetes* 43:1096-

1102). As this mutant only affects insulin binding and not cell-surface expression, residue 323 of IR (residue 313 of IGF-1R) is probably at or near the binding site. Structurally this residue lies in the middle of a region

(residues 309-318 of IGF-1R) which is conserved in both IR and IGF-1R and

5 the surrounding region, 332-345 (of IGF-1R), is also quite well conserved in the these receptors (Figure 9a). Therefore this region is quite likely to form part of the hormone-binding site but would not have been detected by chimeras. It is interesting to note that in this region IRR is not as well conserved as the other two receptors (Shier, P. & Watt, V.M., 1989, 10 J.Biol.Chem. 264:4605-14608).

The distance from this putative hormone-binding region on L2 to that found on L1 is about 30 Å (Figure 8). Thus L1 and L2 appear too far apart to bind IGF-1 or insulin. However, in the crystal structure there is a deep cleft between part of the cys-rich domain (residue 262) and L2 (residue 305) and 15 this cleft is occupied by a loop from a neighbouring molecule. Thus it seems probable that the position of the L2 domain in the receptor structure or the hormone-receptor complex adopts a different position with respect to the cys-rich domain than that found in the crystal. The movement required to bring L2 sufficiently close to L1 is small, namely a rotation of approximately 20 25° about residue 298.

A number of IR mutants have been identified which constitutively activate the receptor and the majority of these are found in the α chain. Curiously all α chain mutants involve changes to or from proline or the deletion of an amino acid, implying that they cause local structural 25 rearrangements. The mutation R86N is similar to wild type but R86P reduces cell-surface expression and insulin binding while constitutively activating autophosphorylation [Grønskov, K. et al., 1993, Biochem. Biophys. Res. Commun. 192, 905-911]. The proline mutation probably disturbs residues preceding 87 which lie in the interface between the L1 and cys-rich domains 30 but it could also affect insulin binding. In the cys-rich domain residues 233, 281, 244 and 247 of IR are not conserved in IGF-1R (Figure 9b) yet L233P [Klinkhamer, M.P. et al., 1989, EMBO J. 8, 2503-2507], deletion of N281 [Debois-Mouthon, C. et al., 1996, J. Clin. Endocrinol. Metab. 81, 719-727] or the triple mutant P243R, P244R and H247D [Rafaeloff, R. et al., 1989, J. Biol. 35 Chem. 264, 15900-15904] cause constitutive kinase activation. Due to their locations each of these three mutants appears likely to compromise the

folding of a β -finger domain and, in turn, the structural integrity of the rod-like cys-rich domain. The structural ramifications of these mutations could be significant for the whole receptor ectodomain as disturbing the L1/cys-rich interface or distorting the rod-like domain could affect the relative position of

5 L1 and the cys-rich domain in this context.

L1 has been further implicated as deletion of K121 on the opposite side of L1 from the cys-rich domain was also found to cause autophosphorylation [Jospe, N. et al., 1994, *J. Clin. Endocrinol. Metab.* 79, 1294-1302]. By contrast this mutation does not affect insulin binding. Thus a possible mechanism emerges for insulin binding and signal transduction. When insulin binds between L1 and L2 it modifies the relative position of L1 and the cys-rich domain in the receptor, perhaps by hinge motion between L2 and the cys-rich domain like that suggested above, and the structural rearrangement is transmitted across the plasma membrane. In the absence of insulin the same signal can be initiated by mutations in the cys-rich region or at the L1/cys-rich interface but at the expense on insulin binding. The signal can also be initiated more directly by mutations on the opposite side of L1 which affect the interaction of L1 with other parts of the ectodomain, possibly the other half of the receptor dimer.

20 **Ligand Studies**

Although there is no structural information about an IGF-1/IGF-1R complex a number of studies have probed the nature of this interaction. Results from cross-linking experiments with IGF-1 and insulin and their cognate receptors are consistent with the hormone binding site proposed above. For example B29 of insulin can be cross-linked to the cys-rich region (residues 205-316) (Yip, C. C., et al., 1988, *Biochim. Biophys. Res. Commun.* 157:321-329) or the L1 domain (Wedekind, F., et al., 1989, *Biol. Chem Hoppe-Seyler*, 370:251-258). However these two regions are reasonably well separated and those studies may indicate that B29 is mobile. Other studies unfortunately do not map the site any more precisely.

Analogues and site-directed mutants of IGF-1 and -2 have been more fruitful. Relative to insulin IGF-1 and -2 contain two extra regions, the C region between B and A and a D peptide at the C-terminus. For IGF-1 replacement of the C region by a four Gly linker reduced affinity for IGF-1R by a factor of 40 but increased affinity for IR 5-fold (Bayne, M.L., et al., 1988, *J. Biol.Chem.* 264:11004-11008). Changes in affinity are consistent with the

deletion in IGF-1 complementing differences in the cys-rich regions of IGF-1R and IR noted above. Mutation of residues either side of the C region (residue 24 for IGF-1 [Cascieri, M.A., et al., 1988, *Biochemistry* 27:3229-3233], residues 27,43 for IGF-2, [Sakano, K., et al., 1991, *J. Biol. Chem.*

5 266:20626-20635]) also have deleterious effects on the affinity of the hormone for IGF-1R as has truncation of the nearby D peptide in IGF-2 (Roth, B.V., et al., 1991, *Biochem. Biophys. Res. Commun.* 181:907-914). Insulin has been extensively mutated. Binding studies [summarised in Kristensen, C. et al., 1997, *J. Biol. Chem.* 272, 12978-12983] indicate that insulin may
 10 bind its receptor via a hydrophobic patch (residues A2, A3, A19, B8, B11, B12, B15 and possibly B23 & B24). However this patch is normally buried and requires the removal of the B chain's C-terminus from the observed position. Assuming IGF-1, -2 and insulin bind their receptors in the same orientation, these data suggest an approximate orientation for the hormone
 15 when bound to the receptor.

One notable feature of IGF-1 and -2 is the large number of charged residues and their uneven distribution over the surface. Basic residues are predominantly found in the C region and, in solution, this region is not well ordered in either IGF-1 or -2 (Sato, A., et al., 1993, *Int J Peptide Protein Res.*
 20 41:433-440; Torres, A. M., et al., 1995, *J. Mol. Biol.* 248:385-401). In contrast the binding site of the receptor has a sizable patch of acidic residues in the corner where the cys-rich domain departs from L1. Other acidic residues which are specific to this receptor are found along the inside face of the cys-rich domain and the loop (residues 255-263) extending from module 6. Thus
 25 it is possible that electrostatics play an important part in IGF-1 binding with the C region binding to the acidic patch of the cys-rich region near L1 and the acidic patch on the other side of the hormone directed towards a small patch of basic residues (residues 307-310) on the N-terminal end of L2.

Although the structure of this fragment gives significant information
 30 about the nature of the hormone binding site, residues outside this region have also been shown to affect binding of ligand. A number of studies have implicated residues 704-715 of IR (Mynarcik, D. C et al., 1996, *J. Biol. Chem.* 271, 2439-2442; Kurose, T., et al., 1994, *J. Biol. Chem.* 269:29190-29197). These residues could contact insulin on one of the sides left open in the
 35 current structure. Using insulin labelled at the B1 residue, Fabry, M., et al., (1992, *J. Biol. Chem.* 267:8950-8956) cross linked insulin to the fragment

390-488, part of which is not near the site as described. The explanation for this could be either 488 reaches back to the hormone binding site, or this region could contact another hormone bound to the other half of the receptor.

Further structural information is needed to establish how these other regions

- 5 contact the hormone and to elucidate how binding of the hormone is communicated to the kinase inside the cell.

- The structure of the L1-cys-rich-L2 domains of IGF-1R presented here represents the first structural information for the extracellular portion of a member of the insulin receptor family. The L domains display a novel fold
- 10 which is common to the EGF receptor family and the modular architecture of the cys-rich domain implies that smaller building blocks should be used to describe the composition of cysteine-rich domains. This fragment contains the major specificity determinants of receptors of this class for their ligands. It has an elongated structure with a space in the middle which could
- 15 accommodate the ligand. The three sides of this site correspond to regions which have been implicated in hormone binding. Although other sites are present in the receptor ectodomain which interact with the ligand this structure gives us an initial view of how the insulin, IGF-1 and -2 might interact with their cell surface receptors to control their metabolic and
- 20 mitogenic effects

- Such information will provide valuable insight into the structure of the corresponding domains of the IR and insulin receptor-related receptor as well as members of the related EGFR family (Bajaj, M., et al., 1987, Biochim Biophys Acta 916:220-226; Ward, C. W. et al., 1995, Proteins: Struct Funct
- 25 Genet 22:141-153).

EXAMPLE 4

Prediction of 3D Structure of the Corresponding Domains of IRR and IR Based on Structure of IGF-1R Frgament.

- The sequence identities between the different members of the insulin
- 30 receptor family are sufficient to allow accurate sequence alignments to facilitate 3D structure predictions by homology modelling. The alignments of the ectodomains of human IGF-1R, IR, and IRR are shown in Figure 13.

EXAMPLE 5**Prediction of 3D Structure of EGFR and its Family Members ERB2, ERB3 and ERB4.**

The sequence identities between the different members of the EGFR
 5 receptor family and the insulin receptor family are sufficient to allow
 accurate sequence alignments to facilitate 3D structure predictions by
 homology modelling. The alignments of the ectodomains of human EGFR,
 ERB2, ERB3 and ERB4 are shown in Figure 14. The ectodomains of the EGFR
 family members are composed of four domains : L1 domain, cys-rich domain,
 10 L2 domain and a second cys-rich domain all of which can be modelled from
 the structure of the IGF-1R fragment residues 1-462.

The sequence alignment analysis and characterization of the repeat
 modules in the cys-rich region of IGF-1R and the homologous regions of the
 IR, IRR and the first and second cys-rich regions of EGFR, ErbB2, ErbB3 and
 15 ErbB4 are shown in Figure 15. A representative of each subtype of cys repeat
 is found in the IGF-1R fragment 1-462 and is used to model each of these
 modules in the other receptors. Note the nature and order of modules in the
 second cys-rich repeat of the EGFR family is different to that seen in the first
 cys-rich region.

20 EXAMPLE 6**Single-Molecule Imaging of Human Insulin Receptor Ectodomain and its Fab Complexes****Cloning and expression of hIR -11 ectodomain protein**

A full length clone of the human IR exon -11 form (hIR -11) was
 25 prepared by exchanging an Aat II fragment, nucleotides 1195 to 2987 , of the
 exon +11 clone (plasmid pET; Ellis et al., 1986; gift from Dr W. J. Rutter,
 UCSF) of hIR (Ebina et al., 1985, *Cell* 40, 747-758) with the equivalent Aat II
 fragment from a plasmid (pHIR/P12-1, ATCC 57493) encoding part of the
 extracellular domain and the entire cytoplasmic domain of hIR -11 (Ullrich
 30 et al., 1985, *Nature* 313 , 756-761). The ectodomain fragment of hIR -11
 (2901 bp, coding for the 27 residue signal sequence and residues His1-
 Asn914) was produced by SalI and SspI digestion and inserted into the
 mammalian expression vector pEE6.HCMV-GS (Celltech Limited, Slough,
 Berkshire, UK) into which a stop codon linker had been inserted, as
 35 described previously (Cosgrove et al., 1995, *Protein Expression and
 Purification* 6, 789-798) for the hIR exon +11 ectodomain.

The resulting recombinant plasmid pHIR II (2 µg) was transfected into glycosylation deficient Chinese hamster ovary (Lec 8) cells (Stanley, 1989, *Molec. Cellul. Biol.* 9, 377-383) with Lipofectin (Gibco-BRL). After transfection, the cells were maintained in glutamine-free medium GMEM

5 (ICN Biomedicals, Australia) as described previously (Bebbington & Hentschel, 1987, In *DNA Cloning* (Glover, D., ectodomain.), Vol III, Academic Press, san Diego; Cosgrove et al., 1995, *Protein Expression and Purification* 6, 789-798). Expressing cell lines were selected for growth in GMEM with 25 µM methionine sulfoximine (MSX, Sigma). Transfectants were screened for
10 protein expression using sandwich ELISA with anti-IR monoclonal antibodies 83-7 and 83-14. Metabolic labelling of cells, immunoprecipitations, insulin binding assays and Scatchard analyses were performed as described previously for the exon +11 form of hIR ectodomain (Cosgrove et al., 1995, , *Protein Expression and Purification* 6, 789-798).

15 **hIR -11 ectodomain production and purification**

The selected clone (inoculum of 1.28×10^8 cells) was grown in a spinner flask packed with 10 g of Fibra-cel disc carriers (Sterilin, U.K.) in 500 ml of GMEM medium containing 10% fetal calf serum (FCS) and 25 µM MSX. Selection pressure was maintained for the duration of the culture.

20 Ectodomain was recovered from harvested media by affinity chromatography on immobilized insulin and further purified by gel filtration chromatography on Superdex S200 (Pharmacia; 1 x 40 cm) in Tris-buffered saline containing 0.02% sodium azide (TBSA) as described previously (Cosgrove et al., 1995, *Protein Expression and Purification* 6, 789-798).

25 Solutions of purified hIR -11 ectodomain were stored at 4° C prior to use.

Production of Fab fragments and their complexes with ectodomain

Purification of Mabs 83-7, 83-14 and 18-44 from ascites fluid by affinity chromatography using Protein A-Sepharose, and the production of Fabs, were based on the methodologies described in Coligan et al., 1993,
30 Current Protocols in Immunology, Vol 1, pp 2.7.1-2.8.9, Greene Publishing Associates & Wiley - Interscience, John Wiley and Sons. Fab was produced from monoclonal antibody by mercuripapain digestion for 1-4 h, followed by gel filtration on Superdex S200. Products were monitored by reducing and non-reducing SDS-PAGE. For 83-7 Mab, an IgG Type 1 monoclonal antibody,
35 the bivalent (Fab)₂' isolated by this method was reduced to monovalent Fab 83-7 by mild reduction with mM L-cysteine.HCl in 100 mM Tris pH 8.0

(Coligan et al., 1993, Current Protocols in Immunology, Vol 1, pp 2.7.1-2.8.9, Greene Publishing Associates & Wiley - Interscience, John Wiley and Sons).

Complexes of Fab with hIR -11 ectodomain were produced by mixing

~ 2.5 to 3.5 molar excess of Fab with hIR -11 ectodomain at ambient

- 5 temperature in TBSA at pH 8.0. After 1-3 h, the complex was separated from unbound Fab by gel filtration over a Superdex S200 column in the same buffer.

Electron microscopy

- 10 Uncomplexed hIR -11 ectodomain and the Fab complexes described above were diluted in phosphate-buffered saline (PBS) to concentrations of the order of 0.01-0.03 mg/ml. Prior to dilution, 10% glutaraldehyde (Fluka) was added to the PBS to achieve a final concentration of 1% glutaraldehyde. Droplets of ~ 3ml of this solution were applied to thin carbon film on 700-
- 15 mesh gold grids after glow-discharging in nitrogen for 30 s. After 1 min. the excess protein solution was drawn off and followed by application and withdrawal of 4-5 droplets of negative stain [2% uranyl acetate (Agar), 2% uranyl formate (K and K), 2% potassium phosphotungstate (Probing and Structure) adjusted to pH 6.0 with KOH, or 2% methylamine tungstate (Agar) adjusted to pH 6.8 with NH₄OH]. In the case of both uranyl acetate and
- 20 uranyl formate staining, an intermediate wash with 2 or 3 droplets of PBS was included prior to application of the stain. The grids were air-dried and then examined at 60kV accelerating voltage in a JEOL 100B transmission electron microscope at a magnification of 100,000x. It was found that there was a typical thickness of negative stain in which Fabs were most easily
- 25 seen, hence areas for photography had to be chosen from particular zones of the grid. Electron micrographs were recorded on Kodak SO-163 film and developed in undiluted Kodak D19 developer. The electron-optical magnification was calibrated under identical imaging conditions by recording single-molecule images of the antigen-antibody complex of influenza virus neuraminidase heads and NC10 MFab (Tulloch et al., 1986, *J.Mol. Biol.* **190**, 215-225; Malby et al., 1994, *Structure*, **2**, 733-746).

Image processing

- Electron micrographs showing particles in a limited number of identifiable projections were chosen for digitisation. Micrographs were
- 35 digitised on a Perkin-Elmer model 1010 GMS PDS flatbed scanning microdensitometer with a scanning aperture (square) size of 20 mm and

stepping increment of 20 nm corresponding to a distance of 0.2 nm on the specimen. Particles were selected from the digitised micrograph using the interactive windowing facility of the SPIDER image processing system (Frank et al., 1996, *J. Struct. Biol.* **116**, 190-199). Particles were scaled to an optical

5 density range of 0.0 - 2.0 and aligned by the PSPC reference-free alignment algorithm (Marco et al., 1996, *Ultramicroscopy*, **66**, 5-10). Averages were then calculated over a subset of correctly aligned particles chosen interactively as being representative of a single view of the particle. The final average image presented here is derived from a library of 94 images.

10 **Biochemical characterization of expressed hIR -11 ectodomain**

The recombinant protein examined corresponded to the the first 914 residues of the 917 residue ectodomain of the exon -11 form of the human insulin receptor (Ullrich et al., 1986, *Nature* **313**, 756-761). Expressed protein was shown, by SDS-PAGE and autoradiography of immunoprecipitated
15 product from metabolically labelled cells, to exist as a homodimeric complex of ~270 - 320 kDa apparent mass, which dissociated under reducing conditions into monomeric α and β' subunits of respective apparent mass ~120 kDa and ~35 kDa (data not shown).

Purified hIR -11 ectodomain, expressed in Lec8 cells and purified by
20 affinity chromatography on an insulin affinity column, ran as a symmetrical peak on a Superdex S200 gel filtration column (Figure 16). The protein eluted with an apparent mass of ~400 kDa, calculated from a standard curve generated by the elution positions of standard proteins (not shown). As expected for protein expressed in Lec 8 cells, whose glycosylation defect
25 produces truncated oligosaccharides (Stanley, 1989, *Molec. Cellul. Biol.* **9**, 377-383), this value is less than the apparent mass (450 - 500 kDa) reported for hIR +11 ectodomain expressed in wild-type CHO-K1 cells (Johnson et al., 1988, *Proc. Natl Acad. Sci USA* **85**, 7516-7520; Cosgrove et al., 1995, *Protein Expression and Purification* **6**, 789-798).

30 Radioassay of insulin binding to purified ectodomain gave linear Scatchard plots and Kd values of $1.5 - 1.8 \times 10^{-9}$ M, similar to the values of $2.4 - 5.0 \times 10^{-9}$ M reported for the hIR -11 ectodomain (Andersen et al., 1990, *Biochemistry* **29**, 7363-7366; Markussen et al., 1991, *J. Biol. Chem.* **266**, 18814-18818; Schaffer, 1994, *Eur. J. Biochem.* **221**, 1127-1132) and the values
35 of $\sim 1.0 - 5.0 \times 10^{-9}$ M reported for the hIR +11 ectodomain (Schaefer et al., 1992, *J. Biol. Chem.* **267**, 23393-23402; Whittaker et al., 1994, *Molec.*

Endocrinol. **8**, 1521-1527; Cosgrove et al., 1995, *Protein Expression and Purification* **6**, 789-798).

Expression of hIGF-1R ectodomain

Cloning, expression and purification of this protein used elements

- 5 common to those described for hIR -11 ectodomain (Cosgrove et al., 1995, *Protein Expression and Purification* **6**, 789-798) and resulted in purified product that was recognised by receptor-specific Mabs 17-69, 24-31 and 24-60 (Soos et al., 1992, *J. Biol. Chem.* **267**, 12955-63) and was composed of α and β' subunits of mass similar to those of hIR ectodomain (unpublished data).

10 **Preparation of hIR -11 ectodomain/MFab complexes**

- A complex of hIR -11 ectodomain and Fab from antibody 83-14 eluted as a symmetrical peak of 460 -500 kDa (Figure 16), as did complexes generated from a mixture of hIR -11 ectodomain with Fab from antibody 18-44 and a mixture of hIR -11 ectodomain with Fab 83-7 (not shown). A co-
 15 complex of ectodomain with Fabs from antibodies 18-44 and 83-14 eluted at \sim 620 kDa (Figure 12), as did a co-complex with MFabs 83-14/83-7 and another with MFabs 83-7/18-44 (not shown). A complex of hIR -11 ectodomain with all three MFab derivatives, 18-44, 83-7 and 83-14, eluted at an apparent mass of \sim 710 kDa (Figure 16).

20 **Electron microscopy**

Imaging of hIR -11 and hIGF-1R ectodomains

- Single-molecule imaging of undecorated dimeric hIR -11 ectodomain was carried out under a variety of negative staining conditions, which emphasised different aspects of the structure of the molecular envelope. The
 25 least aggressive or penetrative stain was potassium phosphotungstate (KPT) , which revealed consistent globular particles with very little internal structure other than a suggestion of a division into two parallel bars. Staining with methylamine tungstate also revealed the parallel bar images, as shown in Figure 17a.

- 30 Further investigation using progressively more penetrative, but also potentially more disruptive, stains confirmed the observations above. Staining with uranyl acetate and uranyl formate showed the separation of the parallel bars most clearly (Figure 17b), but uranyl acetate showed evidence of disrupting the structure of the particles, i.e. a decrease in the consistency
 35 of the particle shape and a tendency for particles to look unravelled or denatured despite having been subjected to chemical cross-linking prior to

staining. In areas of thicker stain, parallel bars predominated (Figure 17b), whereas in more thinly stained regions, U-shaped particles could be identified, sometimes outnumbering the parallel-bar structures (Figure 18a).

An averaged image of the parallel bars seen by staining hIR -11 ectodomain

5 with uranyl formate is shown as an insert in Figure 17b.

In Figures 17c and 18b, images of hIGF-1R ectodomain are shown for comparison with Figure 17b and 18a, respectively, under similar staining conditions.

Imaging of hIR -11 ectodomain complexed with 83-7 MFab

10 This complex was particularly noteworthy for the consistency of the form of the particles, especially under the gentler staining conditions afforded by stains such as KPT and methylamine tungstate. The particles were interpreted as having been restricted in the views they presented, after
15 air-drying on the carbon support film, by the almost diametrically opposite binding of the two Fab arms to the antigen to form a highly elongated complex structure. Under these conditions three distinct views could be recognised as shown in Figure 19. Two views (interpreted as top-down/bottom-up) show the Fab arms displaced clockwise or anti-clockwise as
20 extensions of the parallel plates with two-fold symmetry. The third view shows an image with the two Fab arms in line roughly through the centre of the receptor on its opposite sides, interpreted as a side projection of binding half-way up the plates (Figure 19).

Figure 20 shows a field of particles of hIR -11 ectodomain complexed with 83-7 MFab, stained with uranyl formate. The use of the more aggressive
25 uranyl stains operating at lower pHs revealed internal structure of the molecular envelope at the expense of consistency of the particle morphology. For example, staining with uranyl acetate or uranyl formate showed that parallel bars can be seen in particles in which the Fab arms are displaced either clockwise or anticlockwise but not where the intermediate central or
30 axial position of the two Fab arms is presented in projection. These observations show 83-7 MFab binding roughly half-way up the side-edge of each hIR -11 ectodomain plate. The epitope recognised by Mab 83-7 has been mapped to the cys-rich region, residues 191-297, by analysis of chimeric receptors (Zhang and Roth, 1991, *Proc. Natl. Acad. Sci. USA* **88**, 9858-9862).

Imaging of hIR -11 ectodomain complexed with either 83-14 MFab or 18-44 MFab

Figure 21a shows the complexes formed with Fabs from the most insulin-mimetic antibody Mab 83-14. Projections showing the Fab arms
 5 bound to and extending out from near the base of the U-shaped particles can be identified. A second field of particles (Figure 21b) shows objects composed of two parallel bars as observed for the undecorated ectodomain, with Fab arms projecting obliquely from diametrically opposite extremities. Similar but less definitive images were also seen when MFab 18-44 was
 10 bound to hIR -11 ectodomain (not shown). The epitope for Mab 83-14 is between residues 469-592 (Prigent et al., 1990) in the connecting domain. This domain contains one of the disulphide bonds (Cys524-Cys524) between the two monomers in the IR dimer (Schaffer and Ljungqvist, 1992, *Biochem. Biophys. Res. Commun.* **189**, 650-653). The epitope for Mab 18-44 is a linear
 15 epitope, residues 765-770 (Prigent et al., 1990, *J. Biol. Chem.* **265**, 9970-9977) in the β -chain, near the end of the insert domain (O'Bryan et al., 1991, *Mol. Cell. Biol.* **11**, 5016-5031). The insert domain contains the second disulphide bond connecting the two monomers in the IR dimer (Sparrow et al., 1997, *J. Biol. Chem.*, **272**, 29460-29467).

20 Imaging of hIR -11 ectodomain co-complexed with two different MFabs per monomer

The double complex of hIR -11 ectodomain with MFabs 83-7 and 18-44 was stained with 2% KPT at pH 6.0, and revealed the molecular envelopes shown in Figure 22. The particle appears complex in shape and
 25 can assume a number of different orientations on the carbon support film, giving rise to a number of different projections in the micrograph. The predominant view is of an asymmetric X-shape (some examples circled). It shows the 83-7 MFab arms bound at opposite ends of the parallel bars with the two 18-44 MFabs appearing as shorter projections extending out from
 30 either side of each ectodomain.

Images of the double complex of hIR -11 ectodomain with 83-7 and 83-14 MFabs gave X-shaped images similar to those seen with the 83-7/18-44 double complex (not shown). In contrast the double complex of hIR -11
 35 ectodomain with 18-44 and 83-14 MFabs did not present the characteristic asymmetric X-shapes described above (images not shown). Instead, the molecular envelope appeared to be elongated in many views, with only an

occasional X-shaped projection. While a detailed interpretation of these images would be premature, it is clear that MFabs 18-44 and 83-14, two of the more potent insulin mimetic antibodies (Prigent et al., 1990, *J. Biol.*

Chem. 265, 9970-9977), can bind simultaneously to the receptor.

5 **Imaging of hIR -11 ectodomain co-complexed with three different MFabs per monomer**

Figure 23 shows a field of particles from a micrograph of hIR -11 ectodomain complexed simultaneously with MFabs 83-7, 83-14 and 18-44. In the thicker stain regions the molecular envelope is X-shaped, and looks very
10 similar to that of the double complexes of hIR -11 ectodomain with either 83-7 and 18-44 or 83-7 and 83-14. However, in the more thinly stained regions, particles of greater complexity are visible and it is possible occasionally to identify that there are in fact more than four MFabs bound to the ectodomain dimer.

15 The single-molecule imaging of hIR -11 ectodomain presented here suggests a molecular envelope for this dimeric species significantly different from that of any previously published study. However, an unequivocal determination of the molecular envelope even from the present study is not entirely straightforward. A major complicating factor here has been the
20 relative fragility of the expressed ectodomain when exposed to the rigors of electron microscope preparation by negative staining. For example, staining with potassium phosphotungstate (KPT, pH 6.0-7.0) frequently suggested a denaturation of the dimeric molecules, but when appropriate conditions were satisfied, good seemingly interpretable molecular envelope images were
25 achieved; staining with methylamine tungstate (pH ~7.0) supported the best KPT molecular envelope images, but had the suggestion of a swelling of the molecular structure at neutral pH; and the acid-pH stains of uranyl acetate (pH ~4.2) and uranyl formate (pH~3.0), with their ability to penetrate the ectodomain structure, appeared to illuminate not so much the molecular
30 envelope as the zones of high projected protein density within the dimer.

An amalgam of impressions from these various staining regimens has led to the following interpretation of single-molecule images of these undecorated, or naked, dimers: the predominant dimeric molecular image encountered here has been that of 'parallel bars' of projected protein density.
35 This view is so predominant, indeed, that it suggests there is either a single preferred orientation of the molecules on the glow-discharged carbon support

film, or that this impression of parallel bars of density may represent a mixture of superficially similar structure projections, with the subtleties of these different projections being masked by the relatively coarse resolution of this single-molecule direct imaging. The impression of parallel bars of

5 projected protein density is particularly predominant in regions of thicker negative stain. A second view of the molecular envelope, appreciably less well represented in regions of thicker stain but predominant in regions of thin staining, is that of 'open' U's, or V's. These two views of hIR -11 ectodomain were supported by the single-molecule imaging of hIGF-1R
10 ectodomain under comparable conditions of negative staining.

If the assumption is made that these two recognisable projected views, that of parallel bars and of open U's/V's, are different views of the same dimeric molecule, an assumption strongly supported by the MFab complex imaging, a coarse model of the molecular envelope can be
15 rationalized as in the schematic Figure 24. The model structure is roughly that of a cube, composed of two almost-parallel plates of high protein density, separated by a deep cleft of low protein main-chain and side-chain density able to be penetrated by stain, and connected by intermediate stain-excluding density near what is assumed here to be their base (that is,
20 nearest the membrane-anchoring region). The width of the low-density cleft appears to be of the order of 30-35Å, sufficient to accommodate the binding of the insulin molecule of diameter ca. 30Å, although we have no electron microscopical evidence to support insulin-binding in this cleft at this stage.

It has been established through imaging of bound 83-7 MFab that
25 there is a dimeric two-fold axis normal to the membrane surface between these plates of density. Occasionally, dimer images display a relative displacement of the bars of density, interpreted here as a limited capacity for a shearing of the interconnecting zone between the two plates along their horizontal axis parallel to the membrane; other images show bars skewed
30 from parallel, implying a limited capacity for the plates to rotate independently around the two-fold axis, again via this interconnecting zone. These two observations each suggest a relatively flexible connectivity between the dimer plates in the membrane-proximal region of intermediate protein density, which could possibly contribute to the transmembrane
35 signalling process.

The approximate overall measured dimensions of the ectodomain dimer depicted in Figure 24 are 110 x 90 x 120 Å, calibrated against the dimensions of imaged influenza neuraminidase heads, known from the solved X-ray structure (Varghese et al., 1983, *Nature* **303**, 35-40). It can be

5 noted that there is a compatibility here between the molecular weights and molecular dimensions of these two molecular species: the compact tetrameric influenza neuraminidase heads of Mr ~200 kDa occupy a volume almost 100 x 100 x 60 Å; the more open dimeric insulin receptor ectodomains of similar Mr ~240 kDa imaged here occupy a volume approximately 110 x
10 90 x 120 Å, roughly twice that of the neuraminidase heads, accommodating the slightly higher molecular weight and substantial central low-density cleft.

The low-resolution roughly cubic compact structure proposed here differs substantially from the T-shaped model proposed by Christiansen et al. (1991, *Proc. Natl. Acad. Sci. U. S. A.* **88**, 249-252) and Tranum-Jensen et al.,
15 (1994, *J. Membrane Biol.* **140**, 215-223) for the whole receptor and the elongated model proposed by Schaefer et al. (1992, *J. Biol. Chem.* **267**, 23393-23402) for soluble ectodomain. Significantly, those previous studies did not provide any convincing independent electron microscopical evidence that their imaged objects were in fact insulin receptor.

20 In the present study, the identity of the imaged molecules as hIR -11 ectodomain has been confirmed by imaging complexes of the dimer with Fabs of the three well-established conformational Mabs against native hIR, 83-7, 83-14 and 18-44 (Soos et al., 1986, *Biochem. J.* **235**, 199-208; 1989, *Proc. Natl Acad. Sci. USA* **86**, 5217-5221), bound singly and in combination. In all
25 these instances, virtually every particle in the field of view exhibited MFab decoration through binding to conformational epitopes, establishing not only the identity of the imaged particles but also the conformational integrity of the expressed ectodomains. Furthermore, the cleanliness and uniformity of these hIR -11 ectodomain preparations, both naked and decorated, visualised
30 here by electron microscopy demonstrate their high suitability for X-ray crystallization trials.

The known flexibility of the Fab arms exacerbates image-to-image variability beyond the limited extent already described for the undecorated dimeric ectodomains, complicating any precise interpretation of these
35 antigen-antibody complexes. Such molecular flexibility also renders largely impractical any single-molecule computer image averaging to facilitate image

interpretation, progressively more so with the higher order antigen-antibody complexes studied here.

The most readily interpretable of these images, showing least image-to-image variability, are those of 83-7 MFab bound to dimers where,

5 fortuitously, the antigen-antibody complex is constrained in its degrees of rotational freedom on the carbon support film. Many projected images show the two Fab arms in line roughly through the centre of the antigen on its opposite sides (Figure 19, arrowed examples), interpreted as a side projection of binding half-way up the plates from their membrane-proximal
10 base. Other sub-sets of images (Figure 19, circled examples) show the two Fab arms still parallel but displaced clockwise or anticlockwise with 2-fold symmetry, each Fab approximating an extension of one of the parallel bars of antigen density, interpreted here as representing top or bottom projections along the 2-fold axis. The third projection, along the axis of the Fab arms,
15 could not be sampled here because of the constraining geometry of this molecular complex. These observations suggest binding of 83-7 MFab roughly half-way up the side-edge of the hIR -11 ectodomain plate. This then allows an initial attempt at spatially mapping the 83-7 MFab epitope, which has been sequence-mapped to residues 191-297 in the cys-rich region of the
20 insulin receptor (Zhang and Roth, 1991, *Proc. Natl. Acad. Sci. USA* **88**, 9858-9862). The spatial separation and relative orientations of the two binding epitopes of Mab 83-7 on the hIR -11 ectodomain dimer as indicated here appear inconsistent with the proposal that Mab 83-7 could bind intramolecularly to hIR (O'Brien et al., 1987, *Biochem J.* **6**, 4003-4010).

25 Decoration of the ectodomain dimer with 83-7 MFab established that the two plates of high protein-density are arranged with 2-fold symmetry. Decoration with either 83-14 or 18-44 MFab , on the other hand, allowed sampling of the third projection of the ectodomain dimer precluded by 83-7 MFab binding. Significantly, this third view established unequivocally the U-
30 shaped projection of the hIR -11 ectodomain dimer, something which was only able to be assumed with the undecorated ectodomain images. Further, this projection has allowed a rough spatial mapping close to the base of the U-shaped dimer for the epitopes recognised by 83-14 MFab (residues 469-592, connecting domain) and 18-44 MFab (residues 765-770, b-chain insert
35 domain; exon 11 plus numbering, Prigent et al., 1990, *J. Biol. Chem.* **265**, 9970-9977).

Inherent in the model structure presented in Figure 20 is the implication that, with the two-fold axis aligned normal to the membrane surface, the mouth of the low-density cleft where insulin binding may occur would lie most distant from the transmembrane anchor, whilst the zone of

5 intermediate density connecting the two high-density plates would be in close proximity to the membrane. It follows, in this model, that the L1/cys-rich/L2 domains (Bajaj et al., 1997, *Biochim. Biophys. Acta* **916**, 220-226; Ward et al., 1995, *Proteins: Struct., Funct., Genet.* **22**, 141-153), which comprise much of the insulin-binding region (see Mynarcik et al., 1997, *J. Biol. Chem.* **272**, 2077-2081), most probably lie in the membrane-distal upper halves of
10 the two plates, whilst the membrane-proximal lower halves contain the connecting domains, the fibronectin-type domains, the insert domains and the interchain disulphide bonds (Schaffer and Ljungqvist, 1992, *Biochem. Biophys. Res. Commun.* **189**, 650-653; Sparrow et al., 1997, *J. Biol. Chem.*, **272**,
15 29460-29467). Such a disposition of domains is supported by the images seen with the single MFab decoration, the 83-7 MFab epitope in the cys-rich region being spatially mapped roughly half-way up the side-edge of the ectodomain plates, and the 83-14 and 18-44 MFab epitopes (connecting domain and β -chain insert domain, respectively) being mapped near the base
20 of the plates. Our preference is for a single α - β monomer to occupy a single plate, although the possibility of a single monomer straddling the two plates of protein density cannot be discounted.

The more complex images involving co-binding of two, and even more so of all three, MFabs to each monomer of the ectodomain dimer
25 (Figures 22 and 23) are not easily interpretable with respect to relative domain arrangements within the monomer at present, not least of all because of the difficulty of finding conditions of negative staining that will simultaneously maintain the integrity of the Fab binding while highlighting recognisable and reproducible details of the internal structure of the dimeric
30 IR ectodomain.

The data presented here demonstrate the ability of single-molecule imaging to give an initial insight into the topology of multidomain structures such as the ectodomain of hIR, and the value of combining this technique with that of either single or multiple monoclonal Fab attachment per
35 monomer as a potential means of epitope (and domain) mapping of the structure. By imaging Fab complexes of other members of the family (such as

hIGF-1R ectodomain) and combining available sequence-mapped epitope information with that presented here, a more comprehensive understanding of domain arrangements within the IR family ectodomains should be forthcoming.

- 5 It will be appreciated by persons skilled in the art that numerous variations and/or modifications may be made to the invention as shown in the specific embodiments without departing from the spirit or scope of the invention as broadly described. The present embodiments are, therefore, to be considered in all respects as illustrative and not restrictive

Dated this twenty-seventh day of November 1997

COMMONWEALTH SCIENTIFIC
AND INDUSTRIAL RESEARCH
ORGANISATION
Patent Attorneys for the Applicant:

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ATOM	1	CB	GLU	1	55.633	12.234	66.610	1.00	42.02	AAAA	ATOM	167	CB	ARG	18	53.308	8.825	54.760	1.00	39.69	AAAA
ATOM	2	CC	GLU	1	55.854	13.637	66.613	1.00	51.06	AAAA	ATOM	168	CC	ARG	18	53.873	8.785	53.342	1.00	40.93	AAAA
ATOM	4	C	GLU	1	53.329	12.897	66.183	1.00	34.99	AAAA	ATOM	169	CD	ARG	18	52.939	9.448	52.327	1.00	37.73	AAAA
ATOM	5	O	GLU	1	52.656	12.446	65.263	1.00	41.70	AAAA	ATOM	170	NE	ARG	18	52.625	10.841	52.337	1.00	37.73	AAAA
ATOM	8	N	GLU	1	53.788	10.591	66.588	1.00	40.49	AAAA	ATOM	172	CZ	ARG	18	51.625	11.518	52.093	1.00	37.73	AAAA
ATOM	10	CA	GLU	1	54.191	11.970	66.984	1.00	38.02	AAAA	ATOM	173	NH1	ARG	18	50.849	10.930	51.194	1.00	38.94	AAAA
ATOM	11	N	ILE	1	53.341	11.184	66.491	1.00	26.59	AAAA	ATOM	176	NH2	ARG	18	51.403	12.789	52.413	1.00	36.66	AAAA
ATOM	13	CA	ILE	2	52.502	15.073	65.714	1.00	25.52	AAAA	ATOM	179	C	ARG	18	53.295	8.061	57.096	1.00	39.61	AAAA
ATOM	14	CB	ILE	2	52.756	16.507	66.101	1.00	22.36	AAAA	ATOM	180	O	ARG	18	53.696	9.077	57.718	1.00	41.77	AAAA
ATOM	15	CG2	ILE	2	52.323	17.382	64.982	1.00	24.98	AAAA	ATOM	181	N	LEU	19	52.441	7.138	58.788	1.00	41.35	AAAA
ATOM	16	CG1	ILE	2	51.978	16.887	67.357	1.00	23.05	AAAA	ATOM	183	CA	LEU	19	51.685	6.631	58.570	1.00	39.31	AAAA
ATOM	17	CD1	ILE	2	52.855	17.006	68.594	1.00	21.14	AAAA	ATOM	184	CB	LEU	19	50.273	6.631	58.570	1.00	36.05	AAAA
ATOM	18	O	ILE	2	52.865	14.890	64.282	1.00	28.44	AAAA	ATOM	186	CD1	LEU	19	49.207	7.351	57.734	1.00	32.89	AAAA
ATOM	19	C	ILE	2	54.020	14.655	63.994	1.00	32.38	AAAA	ATOM	187	CD2	LEU	19	49.149	8.824	58.164	1.00	35.67	AAAA
ATOM	20	N	CYS	3	51.891	14.963	63.365	1.00	31.85	AAAA	ATOM	188	C	LEU	19	52.272	6.449	59.935	1.00	38.29	AAAA
ATOM	22	CA	CYS	3	52.186	14.845	61.930	1.00	29.41	AAAA	ATOM	190	N	GLU	20	53.297	5.667	59.666	1.00	38.74	AAAA
ATOM	23	C	CYS	3	51.601	15.854	61.031	1.00	30.00	AAAA	ATOM	192	CA	GLU	20	53.843	4.820	60.690	1.00	41.24	AAAA
ATOM	24	O	CYS	3	50.443	16.292	61.169	1.00	29.00	AAAA	ATOM	193	CB	GLU	20	54.850	3.854	60.085	1.00	46.02	AAAA
ATOM	25	CB	CYS	3	51.746	13.513	61.375	1.00	30.73	AAAA	ATOM	194	CG	GLU	20	56.120	3.567	58.576	1.00	53.96	AAAA
ATOM	26	SG	CYS	3	52.926	12.196	61.747	1.00	31.80	AAAA	ATOM	195	CD	GLU	20	56.898	1.567	58.676	1.00	61.99	AAAA
ATOM	27	N	GLY	4	52.249	16.485	60.230	1.00	34.42	AAAA	ATOM	196	OE1	GLU	20	58.033	3.925	58.297	1.00	61.99	AAAA
ATOM	29	CA	GLY	4	50.887	17.951	58.799	1.00	41.10	AAAA	ATOM	197	OE2	GLU	20	56.372	2.483	58.334	1.00	60.18	AAAA
ATOM	30	C	GLY	4	49.960	17.153	58.680	1.00	44.32	AAAA	ATOM	198	C	GLU	20	54.409	5.413	61.951	1.00	41.44	AAAA
ATOM	31	N	PRO	5	50.797	19.206	58.384	1.00	42.33	AAAA	ATOM	199	O	GLU	20	54.693	4.669	62.865	1.00	44.25	AAAA
ATOM	32	CD	PRO	5	52.063	19.226	58.233	1.00	42.52	AAAA	ATOM	200	N	ASN	21	54.584	6.719	62.048	1.00	42.05	AAAA
ATOM	34	CA	PRO	5	49.691	20.010	57.865	1.00	42.27	AAAA	ATOM	202	CA	ASN	21	55.111	7.241	63.310	1.00	45.95	AAAA
ATOM	35	CB	PRO	5	50.355	20.904	56.816	1.00	44.20	AAAA	ATOM	203	C	ASN	21	56.477	7.913	63.078	1.00	51.49	AAAA
ATOM	36	CG	PRO	5	51.858	20.609	56.921	1.00	44.20	AAAA	ATOM	204	CG	ASN	21	57.581	6.905	62.739	1.00	57.62	AAAA
ATOM	37	C	PRO	5	48.449	19.291	57.346	1.00	42.51	AAAA	ATOM	205	OD1	ASN	21	57.501	5.714	63.094	1.00	60.62	AAAA
ATOM	38	O	PRO	5	47.329	19.842	57.390	1.00	48.00	AAAA	ATOM	206	ND2	ASN	21	58.612	7.399	62.048	1.00	60.62	AAAA
ATOM	39	N	GLY	6	48.606	18.068	56.880	1.00	35.01	AAAA	ATOM	208	C	ASN	21	54.151	8.199	64.052	1.00	45.71	AAAA
ATOM	41	CA	GLY	6	47.438	17.402	56.368	1.00	33.10	AAAA	ATOM	209	O	ASN	21	54.252	8.384	65.276	1.00	46.54	AAAA
ATOM	42	C	GLY	6	48.000	16.378	55.455	1.00	34.10	AAAA	ATOM	210	N	CYS	22	53.219	8.788	63.298	1.00	44.56	AAAA
ATOM	43	CG1	GLY	6	48.899	16.674	54.658	1.00	35.52	AAAA	ATOM	212	CA	CYS	22	52.203	9.731	63.797	1.00	38.05	AAAA
ATOM	44	N	ILE	7	47.472	15.171	55.593	1.00	34.55	AAAA	ATOM	213	C	CYS	22	51.389	9.260	64.983	1.00	34.60	AAAA
ATOM	46	CA	ILE	7	47.914	14.021	54.833	1.00	35.07	AAAA	ATOM	214	C	CYS	22	50.890	8.131	64.995	1.00	37.23	AAAA
ATOM	47	CB	ILE	7	48.148	12.849	55.824	1.00	31.09	AAAA	ATOM	215	CB	CYS	22	51.202	10.031	62.686	1.00	34.22	AAAA
ATOM	48	CG2	ILE	7	48.447	11.546	55.127	1.00	29.86	AAAA	ATOM	216	SG	CYS	22	52.005	10.512	61.147	1.00	32.23	AAAA
ATOM	49	CG1	ILE	7	49.361	13.211	56.679	1.00	31.85	AAAA	ATOM	217	N	THR	23	51.253	10.100	65.987	1.00	27.55	AAAA
ATOM	50	CD1	ILE	7	49.274	12.809	58.114	1.00	37.54	AAAA	ATOM	219	CA	THR	23	50.379	9.717	67.068	1.00	29.84	AAAA
ATOM	51	C	ILE	7	46.920	13.706	53.716	1.00	38.26	AAAA	ATOM	220	CB	THR	23	51.050	9.691	68.424	1.00	29.77	AAAA
ATOM	52	O	ASP	8	45.709	13.795	53.901	1.00	43.01	AAAA	ATOM	221	CG1	THR	23	51.858	10.863	68.567	1.00	35.63	AAAA
ATOM	53	N	ASP	8	47.463	13.399	52.540	1.00	39.79	AAAA	ATOM	222	CG2	THR	23	51.861	8.436	68.591	1.00	26.22	AAAA
ATOM	55	CA	ASP	8	46.693	13.083	51.348	1.00	38.94	AAAA	ATOM	224	C	THR	23	49.342	10.821	67.103	1.00	30.23	AAAA
ATOM	56	CB	ASP	8	46.794	14.238	50.347	1.00	39.19	AAAA	ATOM	225	TS	THR	23	48.389	10.746	67.859	1.00	34.55	AAAA
ATOM	57	CG	ASP	8	45.545	14.398	49.521	1.00	40.50	AAAA	ATOM	226	N	VAL	24	49.531	11.850	66.379	1.00	26.83	AAAA
ATOM	58	OD1	ASP	8	45.010	13.367	49.047	1.00	40.87	AAAA	ATOM	228	CA	VAL	24	48.621	12.969	66.219	1.00	23.15	AAAA
ATOM	59	OD2	ASP	8	45.100	15.559	49.357	1.00	42.02	AAAA	ATOM	229	CB	VAL	24	48.915	14.010	67.308	1.00	17.88	AAAA
ATOM	60	C	ASP	8	47.244	11.826	50.697	1.00	38.78	AAAA	ATOM	230	CG1	VAL	24	47.993	15.249	67.168	1.00	10.19	AAAA
ATOM	61	O	ASP	8	47.883	11.918	49.657	1.00	44.34	AAAA	ATOM	231	CG2	VAL	24	48.749	13.378	68.627	1.00	18.89	AAAA
ATOM	62	N	ILE	9	47.010	10.667	51.315	1.00	37.37	AAAA	ATOM	232	C	VAL	24	48.656	13.711	64.909	1.00	27.08	AAAA
ATOM	64	CA	ILE	9	47.465	9.373	50.793	1.00	34.17	AAAA	ATOM	233	O	VAL	24	49.156	14.812	64.886	1.00	34.93	AAAA
ATOM	65	CB	ILE	9	47.176	8.244	51.820	1.00	28.02	AAAA	ATOM	234	N	ILE	25	48.143	13.156	63.814	1.00	26.66	AAAA
ATOM	66	CG2	ILE	9	47.749	6.929	51.328	1.00	25.06	AAAA	ATOM	236	CA	ILE	25	48.341	13.920	62.556	1.00	26.06	AAAA
ATOM	67	CG1	ILE	9	47.568	8.647	53.227	1.00	23.84	AAAA	ATOM	237	CB	ILE	25	47.351	13.277	61.487	1.00	22.27	AAAA
ATOM	68	CD1	ILE	9	49.168	8.876	53.413	1.00	14.18	AAAA	ATOM	238	CG2	ILE	25	47.398	14.074	60.248	1.00	21.10	AAAA
ATOM	69	C	ILE	9	46.652	9.110	49.510	1.00	37.46	AAAA	ATOM	239	CG1	ILE	25	47.713	11.849	61.237	1.00	26.19	AAAA
ATOM	70	O	ILE	9	45.563	9.785	49.298	1.00	40.86	AAAA	ATOM	240	CD1	ILE	25	46.718	11.005	60.485	1.00	28.07	AAAA
ATOM	71	N	ARG	10	47.074	8.178	48.650	1.00	38.88	AAAA	ATOM	241	C	ILE	25	47.551	15.328	62.821	1.00	25.86	AAAA
ATOM	72	CA	ARG	10	46.332	7.839	47.418	1.00	38.18	AAAA	ATOM	242	O	ILE	25	46.357	15.457	62.933	1.00	31.12	AAAA
ATOM	74	CB	ARG	10	45.567	9.035	46.864	1.00	35.74	AAAA	ATOM	243	N	GLU	26	48.392	16.356	62.950	1.00	23.69	AAAA
ATOM	75	CG	ARG	10	46.317	9.888	45.893	1.00	35.74	AAAA	ATOM	245	CA	GLU	26	47.869	17.691	63.195	1.00	19.28	AAAA
ATOM	76	CD	ARG	10	46.020	11.358	46.397	1.00	41.62	AAAA	ATOM	246	CB	GLU	26	48.319	18.587	64.456	1.00	17.36	AAAA
ATOM	77	NE	ARG	10	45.274	12.029	45.121	1.00	44.13	AAAA	ATOM	247	CG	GLU	26	48.365	19.807	64.456	1.00	20.66	AAAA
ATOM	79	CZ	ARG	10	44.416	13.042	45.295	1.00	42.45	AAAA	ATOM	248	CD	GLU	26	49.459	20.683	64.959	1.00	28.67	AAAA
ATOM	80	NH1	ARG	10	44.164	13.527	46.504	1.00	38.95	AAAA	ATOM	249	OE1	GLU	26	50.340	20.970	64.241	1.00	36.49	AAAA
ATOM	83	NH2	ARG	10	43.824	13.597	44.247	1.00	43.32	AAAA	ATOM	250	OE2	GLU	26	49.456	21.084	66.143	1.00	32.	

Figure 1 (cont'd)

ATOM	325	CG2	ILE	34	44.578	2.278	48.573	1.00	29.54	AAAA	ATOM	488	CA	ILE	51	43.330	13.615	64.097	1.00	18.06	AAAA
ATOM	326	CG1	ILE	34	45.030	4.674	48.137	1.00	32.29	AAAA	ATOM	489	CB	ILE	51	42.410	12.877	63.181	1.00	17.87	AAAA
ATOM	327	CD1	ILE	34	45.632	4.831	49.505	1.00	36.92	AAAA	ATOM	490	CG2	ILE	51	42.347	13.595	61.824	1.00	20.56	AAAA
ATOM	328	C	ILE	34	42.997	1.911	46.541	1.00	41.31	AAAA	ATOM	491	CD1	ILE	51	42.914	11.441	62.995	1.00	19.01	AAAA
ATOM	329	O	ILE	34	41.816	1.677	46.767	1.00	45.84	AAAA	ATOM	492	CD1	ILE	51	42.192	10.630	61.930	1.00	12.40	AAAA
ATOM	330	N	SER	35	43.843	0.938	46.228	1.00	46.80	AAAA	ATOM	493	C	ILE	51	42.704	14.924	64.461	1.00	18.06	AAAA
ATOM	332	CA	SER	35	43.344	-0.421	46.209	1.00	49.73	AAAA	ATOM	494	O	ILE	51	41.547	14.938	64.815	1.00	18.83	AAAA
ATOM	333	CB	SER	35	43.673	-1.112	44.905	1.00	50.47	AAAA	ATOM	495	N	THR	52	43.450	16.022	64.394	1.00	18.18	AAAA
ATOM	334	OC	SER	35	42.462	-1.424	44.229	1.00	52.41	AAAA	ATOM	497	CA	THR	52	42.895	17.284	64.825	1.00	19.65	AAAA
ATOM	336	C	SER	35	43.913	-1.202	47.367	1.00	53.88	AAAA	ATOM	498	CB	THR	52	43.984	18.250	65.229	1.00	16.95	AAAA
ATOM	337	O	SER	35	44.271	-0.631	48.398	1.00	57.19	AAAA	ATOM	499	OG1	THR	52	44.551	18.826	64.064	1.00	20.53	AAAA
ATOM	338	N	LYS	36	43.966	-2.514	47.210	1.00	56.59	AAAA	ATOM	501	CG2	THR	52	45.049	17.554	65.977	1.00	17.67	AAAA
ATOM	340	CA	LYS	36	44.494	-3.384	48.248	1.00	60.80	AAAA	ATOM	502	C	THR	52	41.961	18.002	63.874	1.00	25.70	AAAA
ATOM	341	CB	LYS	36	43.687	-3.225	49.301	1.00	59.09	AAAA	ATOM	503	O	THR	52	41.647	19.180	64.091	1.00	32.66	AAAA
ATOM	342	C	LYS	36	44.411	-4.813	47.708	1.00	65.16	AAAA	ATOM	504	N	GLU	53	41.505	17.310	62.828	1.00	28.12	AAAA
ATOM	343	O	LYS	36	43.311	-5.940	47.400	1.00	65.62	AAAA	ATOM	506	CA	GLU	53	40.566	17.078	61.052	1.00	22.37	AAAA
ATOM	344	N	ALA	37	45.591	-5.402	47.475	1.00	68.90	AAAA	ATOM	507	CB	GLU	53	41.318	18.409	60.658	1.00	14.18	AAAA
ATOM	346	CA	ALA	37	45.736	-6.754	46.919	1.00	71.53	AAAA	ATOM	508	CG	GLU	53	41.622	19.828	60.894	1.00	18.95	AAAA
ATOM	347	CB	ALA	37	47.153	-6.950	46.387	1.00	71.74	AAAA	ATOM	509	CD1	GLU	53	42.306	20.429	59.755	1.00	23.84	AAAA
ATOM	348	C	ALA	37	45.435	-7.848	47.910	1.00	73.82	AAAA	ATOM	510	OE1	GLU	53	42.405	19.724	58.744	1.00	38.01	AAAA
ATOM	349	O	ALA	37	46.052	-7.902	48.972	1.00	75.57	AAAA	ATOM	511	OE2	GLU	53	42.755	21.581	59.848	1.00	25.54	AAAA
ATOM	350	N	GLU	38	44.494	-9.712	47.555	1.00	75.52	AAAA	ATOM	512	GLU	GLU	53	42.418	18.212	55.917	1.00	22.42	AAAA
ATOM	352	CA	GLU	38	45.172	-8.825	48.464	1.00	77.32	AAAA	ATOM	513	O	GLU	53	38.688	16.407	62.242	1.00	26.32	AAAA
ATOM	353	CB	GLU	38	43.312	-10.831	47.746	1.00	78.46	AAAA	ATOM	514	N	TYR	54	39.550	16.455	60.146	1.00	23.93	AAAA
ATOM	354	CG	GLU	38	43.676	-12.133	48.144	1.00	82.68	AAAA	ATOM	516	CA	TYR	54	38.570	15.473	59.725	1.00	23.46	AAAA
ATOM	356	C	GLU	38	45.540	-10.396	48.768	1.00	78.90	AAAA	ATOM	517	CB	TYR	54	37.659	16.094	58.690	1.00	22.70	AAAA
ATOM	357	O	GLU	38	45.867	-10.748	49.904	1.00	78.29	AAAA	ATOM	518	CG	TYR	54	38.346	16.500	57.403	1.00	22.22	AAAA
ATOM	358	N	ASP	39	46.348	-10.454	47.718	1.00	81.01	AAAA	ATOM	519	CD1	TYR	54	38.551	15.588	56.389	1.00	23.21	AAAA
ATOM	360	CA	ASP	39	47.710	-10.934	47.820	1.00	83.83	AAAA	ATOM	520	CE1	TYR	54	39.052	15.979	55.135	1.00	22.51	AAAA
ATOM	361	CB	ASP	39	48.237	-11.303	46.452	1.00	86.03	AAAA	ATOM	522	CD2	TYR	54	38.666	17.819	57.153	1.00	22.12	AAAA
ATOM	362	CG	ASP	39	47.138	-11.457	45.455	1.00	88.51	AAAA	ATOM	521	CE2	TYR	54	39.161	17.212	55.917	1.00	22.42	AAAA
ATOM	363	OD1	ASP	39	46.526	-10.429	45.092	1.00	88.03	AAAA	ATOM	523	C	TYR	54	39.349	17.285	54.912	1.00	22.77	AAAA
ATOM	364	OD2	ASP	39	46.883	-12.612	45.052	1.00	91.17	AAAA	ATOM	524	OH	TYR	54	39.845	17.686	53.690	1.00	27.17	AAAA
ATOM	365	C	ASP	39	48.464	-9.739	48.337	1.00	84.97	AAAA	ATOM	526	C	TYR	54	39.233	14.225	59.169	1.00	21.86	AAAA
ATOM	366	O	ASP	39	48.888	-8.877	47.553	1.00	84.08	AAAA	ATOM	527	O	TYR	54	40.421	14.245	58.960	1.00	21.84	AAAA
ATOM	367	N	TYR	40	48.606	-9.712	49.661	1.00	85.80	AAAA	ATOM	528	N	LEU	55	38.476	13.144	58.958	1.00	16.41	AAAA
ATOM	369	CA	TYR	40	49.268	-8.649	50.412	1.00	87.22	AAAA	ATOM	530	CA	LEU	55	39.027	11.934	58.384	1.00	14.49	AAAA
ATOM	370	CB	TYR	40	49.271	-7.326	49.643	1.00	89.17	AAAA	ATOM	531	CB	LEU	55	38.977	10.801	59.382	1.00	8.63	AAAA
ATOM	371	CG	TYR	40	50.111	-6.208	50.243	1.00	90.06	AAAA	ATOM	532	CG	LEU	55	39.185	9.435	58.731	1.00	12.26	AAAA
ATOM	372	CD1	TYR	40	50.898	-5.405	49.416	1.00	90.06	AAAA	ATOM	533	CD1	LEU	55	39.418	9.508	57.915	1.00	11.44	AAAA
ATOM	373	CE1	TYR	40	51.635	-4.344	49.929	1.00	89.79	AAAA	ATOM	534	CD2	LEU	55	39.342	8.332	59.767	1.00	10.03	AAAA
ATOM	375	CD2	TYR	40	50.087	-5.919	51.613	1.00	89.86	AAAA	ATOM	535	C	LEU	55	38.158	11.604	57.163	1.00	20.33	AAAA
ATOM	377	CE2	TYR	40	50.828	-4.850	52.129	1.00	90.33	AAAA	ATOM	536	O	LEU	55	36.965	11.428	57.305	1.00	25.00	AAAA
ATOM	376	CZ	TYR	40	51.598	-4.069	51.263	1.00	89.97	AAAA	ATOM	537	N	LEU	56	38.747	11.519	55.967	1.00	24.00	AAAA
ATOM	377	OH	TYR	40	52.338	-3.005	51.706	1.00	89.12	AAAA	ATOM	539	CA	LEU	56	37.995	11.243	54.728	1.00	23.70	AAAA
ATOM	379	C	TYR	40	48.347	-8.502	51.592	1.00	87.00	AAAA	ATOM	540	CB	LEU	56	37.999	12.495	53.855	1.00	22.94	AAAA
ATOM	380	O	TYR	40	47.168	-8.186	51.421	1.00	86.23	AAAA	ATOM	541	CG	LEU	56	37.799	12.541	52.336	1.00	20.25	AAAA
ATOM	381	N	ARG	41	48.863	-8.735	52.790	1.00	88.41	AAAA	ATOM	542	CD1	LEU	56	36.533	11.858	51.917	1.00	24.05	AAAA
ATOM	383	CA	ARG	41	48.007	-8.610	53.961	1.00	88.61	AAAA	ATOM	543	CD2	LEU	56	37.704	13.993	51.944	1.00	18.90	AAAA
ATOM	384	CB	ARG	41	48.591	-9.396	55.162	1.00	90.32	AAAA	ATOM	544	C	LEU	56	38.565	10.065	53.941	1.00	27.77	AAAA
ATOM	385	CG	ARG	41	47.741	-10.487	55.527	1.00	89.97	AAAA	ATOM	545	O	LEU	56	39.692	10.125	53.450	1.00	31.41	AAAA
ATOM	387	C	ARG	41	47.695	-7.143	54.331	1.00	85.71	AAAA	ATOM	546	N	LEU	57	37.770	9.001	53.807	1.00	28.80	AAAA
ATOM	388	O	ARG	41	47.256	-6.354	53.475	1.00	87.54	AAAA	ATOM	548	CA	LEU	57	38.282	7.783	53.115	1.00	28.24	AAAA
ATOM	389	N	SER	42	47.939	-6.759	55.577	1.00	79.21	AAAA	ATOM	549	CB	LEU	57	38.080	6.514	54.093	1.00	26.29	AAAA
ATOM	391	CA	SER	42	47.575	-5.425	55.974	1.00	72.84	AAAA	ATOM	550	CG	LEU	57	39.186	6.639	55.150	1.00	23.36	AAAA
ATOM	392	CB	SER	42	46.337	-5.509	56.858	1.00	73.48	AAAA	ATOM	551	CD1	LEU	57	38.605	6.932	56.543	1.00	16.08	AAAA
ATOM	393	OG	SER	42	45.360	-4.573	56.452	1.00	75.54	AAAA	ATOM	552	CD2	LEU	57	39.515	5.307	55.084	1.00	16.55	AAAA
ATOM	395	C	SER	42	48.621	-4.595	56.679	1.00	69.63	AAAA	ATOM	553	CD2	LEU	57	37.377	7.488	51.845	1.00	29.41	AAAA
ATOM	396	O	SER	42	49.730	-5.062	56.962	1.00	70.85	AAAA	ATOM	554	O	LEU	57	36.448	6.515	51.767	1.00	31.40	AAAA
ATOM	397	N	SER	43	48.226	-3.343	56.929	1.00	62.90	AAAA	ATOM	555	N	PHE	58	37.592	8.315	50.837	1.00	30.58	AAAA
ATOM	399	CA	TYR	43	49.008	-2.336	57.641	1.00	53.34	AAAA	ATOM	557	CA	PHE	58	36.910	8.288	49.539	1.00	31.21	AAAA
ATOM	400	CB	TYR	43	49.014	-1.010	56.901	1.00	49.41	AAAA	ATOM	558	CB	PHE	58	37.034	9.718	49.012	1.00	28.90	AAAA
ATOM	401	CG	TYR	43	49.844	-0.967	55.645	1.00	47.49	AAAA	ATOM	559	CG	PHE	58	36.496	9.944	47.668	1.00	27.29	AAAA
ATOM	402	CD1	TYR	43	50.529	-2.083	55.191	1.00	47.33	AAAA	ATOM	560	CD1	PHE	58	37.041	9.339	46.574	1.00	30.10	AAAA
ATOM	403	CE1	TYR	43	51.325	-2.016	54.051	1.00	49.39	AAAA	ATOM	561	CD2	PHE	58	35.524	10.893	47.481	1.00	30.47	AAAA
ATOM	404	CD2	TYR	43	49.969	0.219	54.924	1.00	48.45	AAAA	ATOM	562	CE1	PHE	58	36.631	9.689	45.310	1.00	30.47	

Figure 1 (cont'd)

ATOM	444	N	ASP	58	41.050	-2.279	61.101	1.00	40.43	AAAA	803	CG	TYR	83	31.479	19.989	52.109	1.00	41.19	AAAA	
ATOM	445	CA	ASP	58	42.148	-3.116	60.675	1.00	39.06	AAAA	804	CD1	TYR	83	10.812	19.227	51.145	1.00	43.93	AAAA	
ATOM	447	CB	ASP	58	41.803	-3.788	59.348	1.00	38.97	AAAA	805	CD2	TYR	83	32.440	20.928	51.725	1.00	39.25	AAAA	
ATOM	448	CG	ASP	58	41.035	-5.095	59.552	1.00	43.61	AAAA	806	C	TYR	83	11.532	20.690	55.954	1.00	30.98	AAAA	
ATOM	449	OD1	ASP	58	40.891	-5.601	60.666	1.00	47.80	AAAA	807	O	TYR	83	31.999	21.532	56.727	1.00	37.23	AAAA	
ATOM	450	OD2	ASP	58	40.575	-5.641	58.513	1.00	47.13	AAAA	808	N	ASN	84	30.655	19.811	56.398	1.00	26.48	AAAA	
ATOM	451	C	ASP	58	43.340	-2.184	60.506	1.00	39.04	AAAA	809	CA	ASN	84	30.278	19.928	57.801	1.00	24.16	AAAA	
ATOM	452	O	ASP	58	44.474	-2.611	60.583	1.00	43.53	AAAA	810	CB	ASN	84	28.789	20.218	57.975	1.00	27.22	AAAA	
ATOM	453	N	LEU	69	43.086	-0.900	60.293	1.00	36.34	AAAA	811	CD1	ASN	84	28.218	21.108	57.058	1.00	35.15	AAAA	
ATOM	454	CA	LEU	69	44.171	0.035	60.114	1.00	33.82	AAAA	812	CD2	ASN	84	28.049	22.325	57.058	1.00	33.70	AAAA	
ATOM	455	CB	LEU	69	43.931	0.907	58.887	1.00	31.58	AAAA	813	OD1	ASN	84	27.905	20.503	55.733	1.00	25.06	AAAA	
ATOM	456	CG	LEU	69	44.271	0.359	57.492	1.00	32.42	AAAA	814	ND2	ASN	84	30.578	18.615	58.429	1.00	25.06	AAAA	
ATOM	457	CD1	LEU	69	43.850	1.410	56.473	1.00	31.21	AAAA	815	C	ASN	84	30.111	18.309	59.511	1.00	25.08	AAAA	
ATOM	458	CD2	LEU	69	45.757	0.058	57.335	1.00	35.44	AAAA	816	N	TYR	85	31.371	17.820	57.740	1.00	25.57	AAAA	
ATOM	459	C	LEU	69	44.403	0.909	61.333	1.00	35.44	AAAA	817	CA	TYR	85	31.667	16.499	58.234	1.00	24.08	AAAA	
ATOM	460	CA	LEU	69	45.501	0.905	61.865	1.00	38.91	AAAA	818	CB	TYR	85	31.579	15.531	57.052	1.00	26.41	AAAA	
ATOM	461	N	PHE	76	43.522	1.837	61.777	1.00	34.60	AAAA	819	CD1	TYR	85	30.211	15.603	56.429	1.00	32.86	AAAA	
ATOM	462	CA	PHE	76	43.525	2.532	62.957	1.00	34.07	AAAA	820	CD2	TYR	85	30.017	16.201	55.184	1.00	35.46	AAAA	
ATOM	463	CB	PHE	76	42.919	3.911	62.661	1.00	32.17	AAAA	821	CE1	TYR	85	28.708	16.397	54.663	1.00	39.67	AAAA	
ATOM	464	CG	PHE	76	43.349	4.500	61.356	1.00	34.96	AAAA	822	CE2	TYR	85	29.079	15.185	57.139	1.00	36.81	AAAA	
ATOM	465	CD1	PHE	76	42.403	4.969	60.447	1.00	37.93	AAAA	823	CE2	TYR	85	27.772	15.372	56.635	1.00	37.07	AAAA	
ATOM	466	CD2	PHE	76	44.592	5.579	61.016	1.00	37.23	AAAA	824	CZ	TYR	85	27.585	15.985	55.403	1.00	38.10	AAAA	
ATOM	467	CE1	PHE	76	42.795	5.114	59.192	1.00	38.80	AAAA	825	OH	TYR	85	26.294	16.253	54.952	1.00	38.56	AAAA	
ATOM	468	CE2	PHE	76	45.097	5.118	59.774	1.00	38.80	AAAA	826	O	TYR	85	32.994	16.408	58.961	1.00	22.97	AAAA	
ATOM	469	C	PHE	76	44.136	5.585	58.860	1.00	36.98	AAAA	827	N	ALA	86	33.864	17.243	58.806	1.00	23.68	AAAA	
ATOM	470	C	PHE	76	42.888	1.984	64.268	1.00	34.58	AAAA	828	CA	ALA	86	33.119	15.387	59.784	1.00	19.80	AAAA	
ATOM	471	O	PHE	76	42.028	2.632	64.870	1.00	35.55	AAAA	829	CB	ALA	86	34.308	15.169	60.544	1.00	18.47	AAAA	
ATOM	472	N	PRO	71	43.346	0.817	64.761	1.00	33.10	AAAA	830	CB	ALA	86	34.003	15.329	62.038	1.00	16.82	AAAA	
ATOM	473	CD	PRO	71	44.436	-0.034	64.264	1.00	32.31	AAAA	831	C	ALA	86	34.769	13.752	60.243	1.00	19.67	AAAA	
ATOM	474	CA	PRO	71	42.764	0.267	65.981	1.00	29.95	AAAA	832	O	ALA	86	35.875	13.385	60.598	1.00	26.38	AAAA	
ATOM	475	CB	PRO	71	44.671	-0.902	65.319	1.00	30.32	AAAA	833	N	LEU	87	33.924	12.960	59.589	1.00	18.21	AAAA	
ATOM	476	CG	PRO	71	44.287	-1.774	65.070	1.00	30.97	AAAA	834	CA	LEU	87	34.234	11.569	59.252	1.00	17.46	AAAA	
ATOM	477	C	PRO	71	42.630	1.192	67.164	1.00	30.13	AAAA	835	CB	LEU	87	33.733	10.617	60.359	1.00	15.18	AAAA	
ATOM	478	O	PRO	71	41.814	0.935	68.032	1.00	32.19	AAAA	836	CD1	LEU	87	34.392	9.248	60.579	1.00	17.81	AAAA	
ATOM	479	CA	ASN	72	43.403	2.269	67.227	1.00	32.30	AAAA	837	CD2	LEU	87	33.454	8.293	61.318	1.00	15.41	AAAA	
ATOM	480	CB	ASN	72	43.308	3.129	68.406	1.00	30.05	AAAA	838	OD1	LEU	87	34.808	8.664	59.242	1.00	15.94	AAAA	
ATOM	481	CB	ASN	72	44.627	3.106	69.148	1.00	31.90	AAAA	839	CD1	LEU	87	33.468	11.268	57.990	1.00	21.57	AAAA	
ATOM	482	CG	ASN	72	44.722	1.953	70.108	1.00	37.74	AAAA	840	C	LEU	87	32.266	11.080	58.050	1.00	25.36	AAAA	
ATOM	483	OD1	ASN	72	44.925	2.138	71.321	1.00	43.32	AAAA	841	N	VAL	88	34.134	11.236	56.842	1.00	25.20	AAAA	
ATOM	484	OD2	ASN	72	44.572	0.742	69.580	1.00	38.86	AAAA	842	CA	VAL	88	33.450	10.935	55.579	1.00	23.38	AAAA	
ATOM	485	C	ASN	72	42.848	4.568	69.270	1.00	27.94	AAAA	843	CB	VAL	88	33.034	12.006	54.537	1.00	20.53	AAAA	
ATOM	486	O	ASN	72	42.935	5.334	69.231	1.00	27.94	AAAA	844	CG1	VAL	88	33.282	13.357	55.035	1.00	16.49	AAAA	
ATOM	487	N	LEU	73	42.343	4.945	67.100	1.00	29.19	AAAA	845	CG2	VAL	88	33.976	9.607	55.054	1.00	26.49	AAAA	
ATOM	488	CA	LEU	73	41.863	6.317	66.924	1.00	29.71	AAAA	846	C	VAL	88	35.175	9.468	54.876	1.00	21.57	AAAA	
ATOM	489	CB	LEU	73	41.221	6.507	65.526	1.00	26.82	AAAA	847	N	ILE	89	33.086	8.638	54.829	1.00	28.28	AAAA	
ATOM	490	CG	LEU	73	40.853	7.926	65.067	1.00	23.15	AAAA	848	CA	ILE	89	33.436	7.293	54.333	1.00	27.07	AAAA	
ATOM	491	CD1	LEU	73	42.074	8.769	65.023	1.00	23.71	AAAA	849	CB	ILE	89	33.091	6.197	55.374	1.00	24.54	AAAA	
ATOM	492	CD2	LEU	73	40.199	7.904	63.724	1.00	21.56	AAAA	850	CG2	ILE	89	33.293	4.816	54.777	1.00	27.52	AAAA	
ATOM	493	C	LEU	73	40.843	6.847	68.040	1.00	29.01	AAAA	851	CG1	ILE	89	33.973	6.341	56.606	1.00	20.83	AAAA	
ATOM	494	O	LEU	73	40.010	5.634	68.263	1.00	27.72	AAAA	852	CD1	ILE	89	33.296	5.897	57.855	1.00	18.09	AAAA	
ATOM	495	CA	THR	74	40.889	7.613	68.733	1.00	27.72	AAAA	853	C	ILE	89	32.565	7.053	53.112	1.00	29.32	AAAA	
ATOM	496	CB	THR	74	39.981	7.805	69.835	1.00	28.07	AAAA	854	CG1	ILE	89	31.536	6.406	53.272	1.00	32.09	AAAA	
ATOM	497	CG	THR	74	40.805	7.705	71.094	1.00	33.31	AAAA	855	CG2	ILE	89	33.008	7.533	51.923	1.00	29.27	AAAA	
ATOM	498	OD1	THR	74	41.556	6.556	70.947	1.00	36.36	AAAA	856	N	PHE	90	32.233	7.478	50.709	1.00	26.81	AAAA	
ATOM	499	OD2	THR	74	39.949	7.573	72.347	1.00	32.71	AAAA	857	CB	PHE	90	32.329	8.876	50.101	1.00	26.19	AAAA	
ATOM	500	C	THR	74	39.247	9.122	69.766	1.00	26.87	AAAA	858	CG	PHE	90	31.564	9.053	48.845	1.00	28.17	AAAA	
ATOM	501	CA	THR	74	38.158	9.257	70.315	1.00	28.19	AAAA	859	CD1	PHE	90	30.331	9.678	48.859	1.00	29.79	AAAA	
ATOM	502	CB	THR	74	39.831	10.086	69.071	1.00	23.32	AAAA	860	CD2	PHE	90	32.083	8.627	47.642	1.00	30.39	AAAA	
ATOM	503	C	THR	74	39.220	11.404	68.931	1.00	23.31	AAAA	861	CE1	PHE	90	32.623	9.881	47.693	1.00	31.81	AAAA	
ATOM	504	CB	THR	74	39.769	12.347	69.966	1.00	28.88	AAAA	862	CE2	PHE	90	31.391	8.820	46.465	1.00	32.27	AAAA	
ATOM	505	CG	THR	74	39.203	13.730	69.780	1.00	18.13	AAAA	863	C	PHE	90	34.150	9.454	48.479	1.00	34.74	AAAA	
ATOM	506	CD1	THR	74	39.492	11.782	71.332	1.00	21.05	AAAA	864	CZ	PHE	90	32.582	6.420	49.655	1.00	27.79	AAAA	
ATOM	507	CD2	THR	74	39.559	11.981	67.582	1.00	28.70	AAAA	865	C	PHE	90	33.715	6.079	49.484	1.00	31.26	AAAA	
ATOM	508	C	THR	74	40.664	11.765	67.104	1.00	34.70	AAAA	866	N	GLU	91	31.601	5.924	48.920	1.00	30.86	AAAA	
ATOM	509	O	THR	74	38.598	12.681	66.962	1.00	29.20	AAAA	867	CA	GLU	91	31.853	4.936	47.879	1.00	31.49	AAAA	
ATOM	510	N	ILE	76	38.740	13.353	65.654	1.00	22.21	AAAA											
ATOM	721	CB	ILE	76	37.882	12.671	64.548	1.00	15.15	AAAA	ATOM	880	CB	GLU	91	31.993	5.635	46.532	1.00	35.10	AAAA
ATOM	722	CG	ILE	76	37.825	11.532	63.303	1.00	12.00	AAAA	ATOM	881	CG	GLU	91	31.326	4.876	45.410	1.00	43.61	AAAA
ATOM	723	CD1	ILE	76	37.473	10.463	64.195	1.00	13.43												

Figure 1 (cont'd)

ATOM	960	O	GLY	99	37.444	-0.712	64.884	1.00	30.61	AAAA	ATOM	1129	C	LYS	115	28.391	1.478	49.588	1.00	32.76	AAAA
ATOM	961	N	LEU	100	36.304	1.118	64.206	1.00	27.75	AAAA	ATOM	1130	C	LYS	115	29.478	1.462	49.059	1.00	37.00	AAAA
ATOM	963	CA	LEU	100	37.041	2.075	65.016	1.00	20.90	AAAA	ATOM	1131	N	ASN	116	28.025	2.576	50.500	1.00	34.74	AAAA
ATOM	964	CB	LEU	100	36.784	3.507	64.495	1.00	17.96	AAAA	ATOM	1132	CA	ASN	116	28.905	1.492	50.940	1.00	33.69	AAAA
ATOM	965	CG	LEU	100	37.104	3.790	63.009	1.00	15.59	AAAA	ATOM	1133	CA	ASN	116	29.448	1.816	52.310	1.00	34.99	AAAA
ATOM	966	CD	LEU	100	36.820	5.238	62.704	1.00	11.00	AAAA	ATOM	1134	CB	ASN	116	30.334	1.002	52.472	1.00	38.07	AAAA
ATOM	967	CD2	LEU	100	38.570	3.495	62.695	1.00	11.68	AAAA	ATOM	1135	CG	ASN	116	31.332	2.983	51.600	1.00	37.45	AAAA
ATOM	968	C	LEU	100	36.580	1.936	66.440	1.00	19.15	AAAA	ATOM	1136	OD1	ASN	116	29.944	4.061	52.963	1.00	35.45	AAAA
ATOM	969	O	LEU	100	36.117	2.893	67.028	1.00	18.03	AAAA	ATOM	1137	ND2	ASN	116	28.153	0.179	50.986	1.00	36.41	AAAA
ATOM	970	N	TYR	101	36.730	0.741	66.994	1.00	23.04	AAAA	ATOM	1140	C	ASN	116	27.932	-0.396	52.062	1.00	37.95	AAAA
ATOM	972	CA	TYR	101	36.270	0.456	68.358	1.00	29.36	AAAA	ATOM	1141	O	ASN	116	27.796	-0.303	49.801	1.00	36.22	AAAA
ATOM	973	CB	TYR	101	36.488	-1.023	68.722	1.00	29.53	AAAA	ATOM	1142	N	ALA	117	27.095	-1.512	49.636	1.00	36.19	AAAA
ATOM	974	CG	TYR	101	37.890	-1.499	68.515	1.00	30.31	AAAA	ATOM	1144	CA	ALA	117	27.260	-2.093	48.290	1.00	38.96	AAAA
ATOM	975	CD1	TYR	101	38.844	-1.386	69.525	1.00	29.96	AAAA	ATOM	1145	CB	ALA	117	27.078	-2.595	50.684	1.00	37.86	AAAA
ATOM	976	CD1	TYR	101	40.133	-1.828	69.342	1.00	29.17	AAAA	ATOM	1146	C	ALA	117	26.036	-3.065	51.118	1.00	41.77	AAAA
ATOM	977	CD2	TYR	101	38.268	-2.065	67.306	1.00	30.96	AAAA	ATOM	1147	O	ALA	117	28.272	-3.003	51.106	1.00	38.19	AAAA
ATOM	978	CD2	TYR	101	38.643	-3.540	67.108	1.00	33.23	AAAA	ATOM	1148	N	ASP	118	28.370	-4.080	52.108	1.00	38.19	AAAA
ATOM	979	CZ	TYR	101	40.480	-2.397	68.131	1.00	36.10	AAAA	ATOM	1150	CA	ASP	118	29.378	-5.158	51.719	1.00	44.53	AAAA
ATOM	980	OH	TYR	101	41.755	-2.908	67.934	1.00	44.43	AAAA	ATOM	1151	CB	ASP	118	29.527	-5.314	50.222	1.00	52.11	AAAA
ATOM	982	C	TYR	101	36.834	1.319	69.470	1.00	32.61	AAAA	ATOM	1152	CG	ASP	118	28.722	-6.072	49.628	1.00	56.23	AAAA
ATOM	983	O	TYR	101	36.387	1.213	70.622	1.00	35.65	AAAA	ATOM	1153	OD1	ASP	118	30.452	-4.683	49.645	1.00	55.85	AAAA
ATOM	984	N	ASN	102	37.800	2.733	69.146	1.00	34.59	AAAA	ATOM	1154	OD2	ASP	118	28.759	-3.647	53.471	1.00	33.85	AAAA
ATOM	986	CA	ASN	102	38.368	3.060	70.156	1.00	32.17	AAAA	ATOM	1155	C	ASP	118	28.977	-4.487	54.258	1.00	33.86	AAAA
ATOM	987	CB	ASN	102	39.883	2.919	70.153	1.00	34.62	AAAA	ATOM	1156	CG	ASP	118	28.682	-2.353	53.755	1.00	32.24	AAAA
ATOM	988	CG	ASN	102	40.355	1.842	71.107	1.00	38.21	AAAA	ATOM	1157	N	ASN	119	29.050	-1.880	55.080	1.00	37.83	AAAA
ATOM	989	OD1	ASN	102	40.312	2.018	72.328	1.00	39.16	AAAA	ATOM	1159	CA	LEU	119	29.252	-0.351	55.121	1.00	32.44	AAAA
ATOM	990	ND2	ASN	102	40.783	0.709	70.560	1.00	37.20	AAAA	ATOM	1160	CB	LEU	119	29.561	0.333	56.478	1.00	24.81	AAAA
ATOM	993	C	ASN	102	37.928	4.506	69.909	1.00	33.35	AAAA	ATOM	1161	CG	LEU	119	30.010	1.749	56.268	1.00	19.88	AAAA
ATOM	994	O	ASN	102	38.322	5.443	70.621	1.00	34.93	AAAA	ATOM	1162	CD1	LEU	119	27.975	-2.281	56.057	1.00	32.07	AAAA
ATOM	995	LEU	103	37.089	4.681	68.889	1.00	33.71	AAAA	ATOM	1163	CD2	LEU	119	26.809	-1.990	55.858	1.00	34.96	AAAA	
ATOM	997	CA	LEU	103	36.571	6.006	68.539	1.00	31.56	AAAA	ATOM	1164	C	LEU	119	28.376	-2.980	57.107	1.00	33.19	AAAA
ATOM	998	CB	LEU	103	35.879	5.969	67.157	1.00	27.71	AAAA	ATOM	1165	O	LEU	119	27.448	-3.401	58.132	1.00	33.58	AAAA
ATOM	999	CG	LEU	103	35.499	7.287	66.474	1.00	24.01	AAAA	ATOM	1166	N	CYS	120	27.930	-2.883	59.481	1.00	32.10	AAAA
ATOM	1000	CD1	LEU	103	36.520	8.393	66.654	1.00	14.91	AAAA	ATOM	1169	C	CYS	120	28.803	-2.019	59.546	1.00	31.93	AAAA
ATOM	1001	CD2	LEU	103	35.342	6.955	65.028	1.00	26.24	AAAA	ATOM	1170	O	CYS	120	27.352	-4.917	58.155	1.00	37.34	AAAA
ATOM	1002	C	LEU	103	35.598	6.465	69.623	1.00	30.84	AAAA	ATOM	1171	CB	CYS	120	26.669	-5.595	56.623	1.00	50.24	AAAA
ATOM	1003	O	LEU	103	34.479	5.966	69.711	1.00	30.84	AAAA	ATOM	1172	SG	CYS	120	27.385	-3.430	60.558	1.00	28.35	AAAA
ATOM	1004	N	ARG	104	36.027	7.422	70.437	1.00	28.81	AAAA	ATOM	1173	N	TYR	121	27.776	-2.984	61.864	1.00	27.48	AAAA
ATOM	1006	CA	ARG	104	35.135	7.877	71.502	1.00	26.99	AAAA	ATOM	1175	CA	TYR	121	29.331	-3.567	62.266	1.00	26.64	AAAA
ATOM	1007	CB	ARG	104	35.854	7.732	72.815	1.00	21.56	AAAA	ATOM	1176	CB	TYR	121	28.977	-4.954	62.829	1.00	28.75	AAAA
ATOM	1008	CG	ARG	104	36.358	6.291	73.131	1.00	33.27	AAAA	ATOM	1177	CD1	TYR	121	29.360	-0.651	62.981	1.00	32.77	AAAA
ATOM	1009	CD	ARG	104	35.132	5.480	73.542	1.00	35.96	AAAA	ATOM	1178	CD1	TYR	121	29.080	-7.364	62.543	1.00	38.16	AAAA
ATOM	1010	NE	ARG	104	35.425	4.508	74.671	1.00	40.69	AAAA	ATOM	1179	CD1	TYR	121	28.318	-5.160	64.047	1.00	33.31	AAAA
ATOM	1011	CZ	ARG	104	34.889	5.012	75.977	1.00	46.43	AAAA	ATOM	1180	CD2	TYR	121	28.030	-6.440	64.512	1.00	36.57	AAAA
ATOM	1015	C	ARG	104	34.577	9.269	71.408	1.00	26.71	AAAA	ATOM	1181	CD2	TYR	121	28.409	-7.543	63.750	1.00	39.97	AAAA
ATOM	1016	O	ARG	104	33.618	9.562	72.120	1.00	28.45	AAAA	ATOM	1182	CZ	TYR	121	28.087	-8.825	54.171	1.00	45.83	AAAA
ATOM	1017	N	ASN	105	35.067	10.131	70.531	1.00	24.00	AAAA	ATOM	1183	OH	TYR	121	27.832	-1.496	61.770	1.00	27.62	AAAA
ATOM	1019	CA	ASN	105	34.510	11.764	70.528	1.00	20.78	AAAA	ATOM	1185	C	TYR	121	28.835	-0.886	62.121	1.00	31.85	AAAA
ATOM	1020	CB	ASN	105	35.116	12.263	71.694	1.00	18.92	AAAA	ATOM	1186	O	TYR	121	26.741	-0.924	61.246	1.00	31.58	AAAA
ATOM	1021	CG	ASN	105	34.633	13.727	71.779	1.00	27.94	AAAA	ATOM	1188	CA	LEU	122	26.544	0.022	61.095	1.00	34.25	AAAA
ATOM	1022	OD1	ASN	105	35.169	14.600	71.096	1.00	28.87	AAAA	ATOM	1189	CB	LEU	122	26.220	0.889	59.629	1.00	34.73	AAAA
ATOM	1023	ND2	ASN	105	33.540	13.937	72.645	1.00	29.96	AAAA	ATOM	1190	CG	LEU	122	26.446	2.307	59.080	1.00	32.00	AAAA
ATOM	1025	C	ASN	105	34.835	12.187	69.278	1.00	21.18	AAAA	ATOM	1191	CG	LEU	122	27.332	3.142	60.012	1.00	35.62	AAAA
ATOM	1026	O	ASN	105	35.585	12.254	68.930	1.00	24.92	AAAA	ATOM	1192	CD1	LEU	122	27.075	2.197	57.705	1.00	34.68	AAAA
ATOM	1027	N	ILE	106	33.831	12.707	68.584	1.00	21.36	AAAA	ATOM	1193	CD2	LEU	122	25.366	0.937	61.989	1.00	34.51	AAAA
ATOM	1029	CA	ILE	106	34.085	13.506	67.421	1.00	16.76	AAAA	ATOM	1194	C	LEU	122	25.439	1.927	62.727	1.00	36.07	AAAA
ATOM	1030	CB	ILE	106	33.202	13.383	66.231	1.00	15.96	AAAA	ATOM	1195	N	SER	123	24.285	0.162	61.928	1.00	34.51	AAAA
ATOM	1031	CG1	ILE	106	33.129	12.064	65.230	1.00	17.07	AAAA	ATOM	1196	N	SER	123	24.094	0.456	62.108	1.00	32.96	AAAA
ATOM	1032	CG1	ILE	106	33.645	11.702	65.747	1.00	15.14	AAAA	ATOM	1197	CB	SER	123	21.887	-0.229	62.107	1.00	30.94	AAAA
ATOM	1033	CD1	ILE	106	33.124	11.361	64.414	1.00	13.19	AAAA	ATOM	1200	CG	SER	123	21.890	-1.604	62.443	1.00	35.07	AAAA
ATOM	1034	C	ILE	106	33.748	14.909	67.876	1.00	19.75	AAAA	ATOM	1202	C	SER	123	23.223	0.032	64.199	1.00	31.93	AAAA
ATOM	1035	O	ILE	106	32.665	15.401	67.655	1.00	26.42	AAAA	ATOM	1203	O	SER	123	22.302	0.265	65.017	1.00	28.30	AAAA
ATOM	1036	N	THR	107	34.699	15.538	68.563	1.00	21.76	AAAA	ATOM	1204	N	THR	124	24.370	-0.578	64.525	1.00	27.90	AAAA
ATOM	1038	CA	THR	107	34.555	16.878	69.137	1.00	19.73	AAAA	ATOM	1206	CA	THR	124	24.662	-1.054	65.851	1.00	24.08	AAAA
ATOM	1039	CB	THR	107	35.964	17.477	69.516	1.00	20.13	AAAA	ATOM	1207	CB	THR	124	25.409	-2.336	65.826	1.00	24.86	AAAA
ATOM	1040	CG1																			

Figure 1 (cont'd)

ATOM	1285	CG	ASP	132	27.203	17.299	71.693	1.00	40.20	AAAA	1445	C	ASP	150	24.550	-8.922	63.833	1.00	41.30	AAAA
ATOM	1286	OD1	ASP	132	26.867	16.379	72.461	1.00	46.09	AAAA	1446	C	ASP	150	23.455	-8.771	64.376	1.00	44.58	AAAA
ATOM	1287	OD2	ASP	132	27.531	18.454	72.055	1.00	44.38	AAAA	1447	N	LEU	151	25.536	-9.597	64.376	1.00	47.30	AAAA
ATOM	1288	C	ASP	132	25.570	15.814	68.876	1.00	29.06	AAAA	1448	CA	LEU	151	25.539	-10.181	65.699	1.00	35.09	AAAA
ATOM	1289	O	ASP	132	24.460	15.362	69.093	1.00	34.86	AAAA	1449	CA	LEU	151	25.407	-11.702	65.571	1.00	38.32	AAAA
ATOM	1290	N	ALA	133	25.913	16.511	67.792	1.00	26.57	AAAA	1450	CG	LEU	151	24.168	-12.207	65.097	1.00	36.38	AAAA
ATOM	1292	CA	ALA	133	25.060	16.891	66.665	1.00	22.34	AAAA	1451	C	LEU	151	26.452	-9.655	66.624	1.00	34.61	AAAA
ATOM	1293	CB	ALA	133	25.767	17.973	65.893	1.00	22.29	AAAA	1454	O	LEU	151	27.640	-9.885	66.409	1.00	33.41	AAAA
ATOM	1294	C	ALA	133	24.683	15.764	65.703	1.00	23.13	AAAA	1455	N	CYS	152	26.045	-8.937	67.659	1.00	35.32	AAAA
ATOM	1295	O	ALA	133	24.814	15.896	64.478	1.00	20.05	AAAA	1457	CA	CYS	152	27.005	-8.165	68.593	1.00	40.29	AAAA
ATOM	1296	N	VAL	134	24.194	14.667	66.252	1.00	22.27	AAAA	1458	CG	CYS	152	27.570	-9.463	69.488	1.00	43.08	AAAA
ATOM	1298	CA	VAL	134	23.813	13.524	65.448	1.00	24.74	AAAA	1459	O	CYS	152	26.977	-10.542	69.568	1.00	45.04	AAAA
ATOM	1299	CB	VAL	134	23.360	12.371	66.353	1.00	23.00	AAAA	1460	CB	CYS	152	26.342	-7.288	69.473	1.00	37.72	AAAA
ATOM	1300	CG1	VAL	134	22.194	12.804	67.154	1.00	27.35	AAAA	1461	SG	CYS	152	26.098	-5.631	68.736	1.00	37.05	AAAA
ATOM	1301	CG2	VAL	134	22.996	11.152	65.547	1.00	29.73	AAAA	1462	N	PRO	153	28.739	-9.216	70.121	1.00	45.27	AAAA
ATOM	1302	C	VAL	134	22.699	13.884	64.478	1.00	29.00	AAAA	1463	CD	PRO	153	29.565	-8.005	69.275	1.00	41.44	AAAA
ATOM	1303	O	VAL	134	22.585	13.243	64.447	1.00	28.69	AAAA	1464	CA	PRO	153	29.381	-10.190	71.012	1.00	47.26	AAAA
ATOM	1304	N	SER	135	21.880	14.869	64.824	1.00	29.17	AAAA	1465	CB	PRO	153	30.591	-9.439	71.562	1.00	46.10	AAAA
ATOM	1306	CA	SER	135	20.796	15.257	63.950	1.00	29.17	AAAA	1466	CG	PRO	153	30.894	-8.438	70.530	1.00	44.99	AAAA
ATOM	1307	CB	SER	135	20.111	16.477	64.504	1.00	27.10	AAAA	1467	C	PRO	153	28.442	-10.694	72.113	1.00	50.81	AAAA
ATOM	1308	OG	SER	135	21.062	17.327	65.070	1.00	23.39	AAAA	1468	O	PRO	153	27.825	-9.923	72.865	1.00	47.27	AAAA
ATOM	1310	C	SER	135	21.232	15.552	62.511	1.00	34.63	AAAA	1469	N	GLY	154	28.339	-12.015	72.167	1.00	57.66	AAAA
ATOM	1311	O	SER	135	20.404	15.509	61.586	1.00	36.08	AAAA	1471	CA	GLY	154	27.477	-12.680	73.125	1.00	67.39	AAAA
ATOM	1312	N	ASN	136	22.506	15.866	62.272	1.00	38.35	AAAA	1472	C	GLY	154	26.279	-11.335	72.442	1.00	72.88	AAAA
ATOM	1314	CA	ASN	136	22.847	16.132	60.882	1.00	39.46	AAAA	1473	O	GLY	154	25.998	-14.517	72.652	1.00	75.86	AAAA
ATOM	1315	CB	ASN	136	23.071	17.638	60.650	1.00	43.49	AAAA	1474	N	THR	155	25.592	-12.553	71.612	1.00	75.09	AAAA
ATOM	1316	CG	ASN	136	24.278	18.155	61.314	1.00	48.28	AAAA	1476	CA	THR	155	24.400	-12.555	70.877	1.00	76.56	AAAA
ATOM	1317	OD1	ASN	136	24.266	18.420	62.514	1.00	51.34	AAAA	1477	OD2	THR	155	24.209	-12.007	69.709	1.00	74.93	AAAA
ATOM	1318	ND2	ASN	136	25.349	18.334	60.533	1.00	58.07	AAAA	1478	OG1	THR	155	24.169	-10.665	70.210	1.00	73.48	AAAA
ATOM	1321	C	ASN	136	23.897	15.261	60.189	1.00	35.57	AAAA	1480	CG2	THR	155	22.932	-12.329	68.965	1.00	74.24	AAAA
ATOM	1322	O	ASN	136	24.619	15.688	59.273	1.00	35.22	AAAA	1481	C	THR	155	24.253	-14.386	70.351	1.00	80.38	AAAA
ATOM	1323	N	ASN	137	23.894	14.002	60.613	1.00	30.60	AAAA	1482	O	THR	155	23.234	-15.039	70.602	1.00	80.09	AAAA
ATOM	1325	CA	ASN	137	24.734	12.974	60.051	1.00	27.82	AAAA	1483	N	MET	156	25.248	-14.868	69.609	1.00	85.70	AAAA
ATOM	1326	CB	ASN	137	24.732	11.747	60.947	1.00	20.95	AAAA	1485	CA	MET	156	25.191	-16.219	69.038	1.00	89.85	AAAA
ATOM	1327	CG	ASN	137	25.518	11.972	62.173	1.00	17.44	AAAA	1486	CB	MET	156	26.072	-16.305	67.777	1.00	93.38	AAAA
ATOM	1328	OD1	ASN	137	26.004	13.483	63.075	1.00	18.82	AAAA	1487	CG	MET	156	25.481	-15.618	66.526	1.00	96.33	AAAA
ATOM	1329	ND2	ASN	137	25.654	10.958	62.993	1.00	17.51	AAAA	1488	SD	MET	156	24.697	-16.732	65.287	1.00	100.00	AAAA
ATOM	1332	C	ASN	137	24.005	12.675	58.769	1.00	30.30	AAAA	1489	CE	MET	156	22.937	-16.735	65.851	1.00	99.65	AAAA
ATOM	1333	O	ASN	137	22.792	12.889	58.720	1.00	32.71	AAAA	1490	C	MET	156	25.611	-17.282	70.053	1.00	90.29	AAAA
ATOM	1334	N	TYR	138	24.732	12.178	57.754	1.00	31.84	AAAA	1491	O	MET	156	25.660	-18.479	69.752	1.00	90.33	AAAA
ATOM	1336	CA	TYR	138	24.188	11.869	56.427	1.00	27.90	AAAA	1492	N	GLU	157	25.918	-16.820	71.258	1.00	90.50	AAAA
ATOM	1337	CB	TYR	138	24.795	12.791	55.375	1.00	30.75	AAAA	1494	CA	GLU	157	26.315	-17.690	72.360	1.00	89.67	AAAA
ATOM	1338	CG	TYR	138	24.084	12.786	54.027	1.00	39.45	AAAA	1495	CB	GLU	157	27.735	-17.337	72.819	1.00	91.16	AAAA
ATOM	1339	OD1	TYR	138	23.004	13.483	53.075	1.00	39.30	AAAA	1496	C	GLU	157	25.926	-17.500	72.512	1.00	87.84	AAAA
ATOM	1340	CE1	TYR	138	22.744	13.928	52.305	1.00	41.16	AAAA	1497	O	GLU	157	25.653	-17.718	74.678	1.00	87.91	AAAA
ATOM	1341	OD2	TYR	138	24.274	11.732	53.122	1.00	41.34	AAAA	1498	N	GLU	158	24.103	-17.090	73.164	1.00	85.28	AAAA
ATOM	1342	CE2	TYR	138	23.721	11.774	51.838	1.00	41.53	AAAA	1500	CA	GLU	158	23.047	-16.845	74.135	1.00	83.35	AAAA
ATOM	1343	CH	TYR	138	22.966	12.866	51.439	1.00	43.01	AAAA	1501	CB	GLU	158	22.200	-18.100	74.328	1.00	88.22	AAAA
ATOM	1344	OZ	TYR	138	22.480	12.879	50.148	1.00	45.40	AAAA	1502	CG	GLU	158	20.684	-17.845	74.202	1.00	94.30	AAAA
ATOM	1346	C	TYR	138	25.480	10.462	56.060	1.00	24.71	AAAA	1503	CD	GLU	158	20.072	-18.391	72.903	1.00	96.87	AAAA
ATOM	1347	O	TYR	138	25.569	10.147	55.635	1.00	26.78	AAAA	1504	OE1	GLU	158	20.829	-18.638	71.924	1.00	99.26	AAAA
ATOM	1348	N	ILE	139	23.474	9.621	56.185	1.00	25.08	AAAA	1505	N	GLU	158	18.931	-18.590	72.865	1.00	97.31	AAAA
ATOM	1350	CA	ILE	139	23.646	8.221	55.906	1.00	25.42	AAAA	1506	C	GLU	158	23.616	-16.384	75.460	1.00	79.85	AAAA
ATOM	1351	CB	ILE	139	23.385	7.409	57.190	1.00	24.65	AAAA	1507	O	GLU	158	23.141	-16.755	76.529	1.00	78.54	AAAA
ATOM	1352	CG2	ILE	139	23.951	5.998	57.062	1.00	21.93	AAAA	1508	N	LYS	159	24.660	-15.574	75.356	1.00	77.41	AAAA
ATOM	1353	CG1	ILE	139	24.040	8.105	58.385	1.00	26.03	AAAA	1510	CA	LYS	159	25.354	-14.997	76.497	1.00	76.20	AAAA
ATOM	1354	CD1	ILE	139	24.283	7.160	59.572	1.00	30.19	AAAA	1511	CB	LYS	159	26.512	-15.875	76.893	1.00	77.33	AAAA
ATOM	1355	C	ILE	139	22.777	7.663	54.788	1.00	26.95	AAAA	1512	C	LYS	159	25.861	-13.645	75.997	1.00	74.53	AAAA
ATOM	1356	O	ILE	139	21.562	7.686	54.868	1.00	32.63	AAAA	1513	O	LYS	159	27.042	-13.497	75.673	1.00	73.55	AAAA
ATOM	1357	V	VAL	140	21.377	7.386	53.740	1.00	26.25	AAAA	1514	N	GLU	160	24.960	-12.912	74.932	1.00	72.49	AAAA
ATOM	1359	CA	VAL	140	22.578	6.560	52.694	1.00	26.60	AAAA	1515	CD	PRO	160	23.560	-12.800	76.372	1.00	72.38	AAAA
ATOM	1360	CB	VAL	140	21.881	7.660	51.875	1.00	27.22	AAAA	1516	CA	PRO	160	25.213	-11.269	75.476	1.00	69.47	AAAA
ATOM	1361	CG1	VAL	140	22.850	8.255	50.882	1.00	23.87	AAAA	1517	CB	PRO	160	23.843	-10.597	75.562	1.00	69.23	AAAA
ATOM	1362	CG2	VAL	140	20.616	7.087	51.169	1.00	26.11	AAAA	1518	CG	PRO	160	23.108	-11.381	76.566	1.00	70.96	AAAA
ATOM	1363	C	VAL	140	23.392	5.694	51.754	1.00	30.61	AAAA	1519	C	PRO	160	26.272	-10.481	76.222	1.00	66.84	AAAA
ATOM	1364	O	VAL	140	24.579	5.945	51.567	1.00	34.21	AAAA	1520	O	PRO	160	26.306	-10.439	77.458	1.00	65.35	AAAA
ATOM	1365	N	GLY	141	22.751	4.661	51.201	1.00	30.44	AAAA	1521	N	MET	161	27.111	-9				

Figure 1 (cont'd)

ATOM	1603	OD1	ASN	169	10.912	5.715	70.234	1.00	34.96	AAAA	1775	O	LVS	183	32.015	3.424	75.624	1.00	44.70	AAAA
ATOM	1604	ND2	ASN	169	8.145	6.266	68.070	1.00	37.96	AAAA	1776	N	MET	184	30.602	1.945	76.503	1.00	40.72	AAAA
ATOM	1607	C	ASN	169	12.891	2.134	68.758	1.00	38.51	AAAA	1777	C	MET	184	29.890	2.991	77.223	1.00	41.46	AAAA
ATOM	1608	O	ASN	169	11.977	1.320	68.658	1.00	38.10	AAAA	1778	C	MET	184	28.422	2.563	77.447	1.00	40.50	AAAA
ATOM	1609	N	GLU	170	14.170	1.803	68.845	1.00	38.73	AAAA	1779	CG	MET	184	27.393	1.666	76.484	1.00	39.92	AAAA
ATOM	1611	CA	GLU	170	14.636	0.439	68.874	1.00	39.83	AAAA	1780	CG	MET	184	26.858	2.058	75.119	1.00	40.97	AAAA
ATOM	1612	CB	GLU	170	15.283	0.061	67.536	1.00	49.61	AAAA	1781	SD	MET	184	26.902	0.624	75.907	1.00	38.40	AAAA
ATOM	1613	CG	GLU	170	15.641	-1.439	67.359	1.00	61.29	AAAA	1782	CE	MET	184	30.691	3.044	78.534	1.00	42.46	AAAA
ATOM	1614	CD	GLU	170	14.986	-2.100	66.120	1.00	68.52	AAAA	1783	C	MET	184	31.461	2.136	78.790	1.00	42.71	AAAA
ATOM	1615	OE1	GLU	170	14.678	-3.324	66.196	1.00	74.04	AAAA	1784	O	MET	184	30.540	4.086	79.352	1.00	47.34	AAAA
ATOM	1616	OE2	GLU	170	14.779	-1.406	65.083	1.00	69.19	AAAA	1785	N	CYS	185	31.275	4.163	80.629	1.00	51.78	AAAA
ATOM	1617	C	GLU	170	15.660	0.444	69.987	1.00	36.76	AAAA	1786	C	CYS	185	30.392	4.206	81.906	1.00	56.36	AAAA
ATOM	1618	O	GLU	170	16.590	1.243	70.006	1.00	34.63	AAAA	1787	CA	CYS	185	29.204	4.519	81.832	1.00	57.21	AAAA
ATOM	1619	N	TYR	171	15.441	-0.436	70.943	1.00	35.62	AAAA	1788	O	CYS	185	33.220	5.365	80.614	1.00	51.40	AAAA
ATOM	1621	CA	TYR	171	16.312	-0.577	72.086	1.00	34.58	AAAA	1791	SG	CYS	185	33.964	4.877	80.835	1.00	51.40	AAAA
ATOM	1622	CB	TYR	171	15.447	-0.785	71.125	1.00	12.84	AAAA	1792	N	PRO	186	30.874	3.814	82.104	1.00	60.68	AAAA
ATOM	1623	CG	TYR	171	16.170	-1.153	74.595	1.00	32.37	AAAA	1793	CD	PRO	186	32.378	3.611	83.424	1.00	60.70	AAAA
ATOM	1624	CD1	TYR	171	17.036	-0.249	75.207	1.00	28.71	AAAA	1794	CA	PRO	186	30.176	3.932	84.329	1.00	57.39	AAAA
ATOM	1625	CE1	TYR	171	17.645	-0.536	76.382	1.00	25.22	AAAA	1795	CB	PRO	186	31.203	3.773	85.448	1.00	57.78	AAAA
ATOM	1626	CD2	TYR	171	15.938	-2.381	75.217	1.00	30.01	AAAA	1796	CG	PRO	186	32.488	4.105	84.839	1.00	58.33	AAAA
ATOM	1627	CE2	TYR	171	16.547	-2.679	76.401	1.00	32.15	AAAA	1797	C	PRO	186	29.387	5.199	84.460	1.00	57.19	AAAA
ATOM	1628	CH	TYR	171	17.403	-1.744	76.978	1.00	33.12	AAAA	1798	CG	PRO	186	29.923	6.303	84.386	1.00	56.02	AAAA
ATOM	1629	OH	TYR	171	18.020	-2.018	78.167	1.00	35.12	AAAA	1799	N	SER	187	29.092	4.992	84.632	1.00	58.28	AAAA
ATOM	1631	C	TYR	171	17.170	-1.804	71.798	1.00	35.29	AAAA	1801	CA	SER	187	27.102	6.045	84.782	1.00	61.94	AAAA
ATOM	1632	O	TYR	171	16.658	-2.907	71.652	1.00	33.63	AAAA	1802	CB	SER	187	25.817	5.410	85.330	1.00	67.34	AAAA
ATOM	1633	N	ASN	172	18.478	-1.618	71.679	1.00	36.68	AAAA	1803	OG	SER	187	25.839	3.986	85.161	1.00	70.00	AAAA
ATOM	1635	CA	ASN	172	19.316	-2.760	71.416	1.00	34.29	AAAA	1805	C	SER	187	27.603	7.143	85.710	1.00	61.19	AAAA
ATOM	1636	CB	ASN	172	19.231	-3.099	69.947	1.00	37.52	AAAA	1806	O	SER	187	27.481	8.334	85.436	1.00	60.53	AAAA
ATOM	1637	CG	ASN	172	18.789	-4.515	69.721	1.00	43.69	AAAA	1807	N	THR	188	28.161	6.704	86.824	1.00	60.90	AAAA
ATOM	1638	ND1	ASN	172	19.263	-5.438	70.384	1.00	48.62	AAAA	1809	CA	THR	188	28.737	7.572	87.832	1.00	58.92	AAAA
ATOM	1639	ND2	ASN	172	17.872	-7.705	68.795	1.00	33.45	AAAA	1810	CG	THR	188	29.629	6.737	88.727	1.00	60.61	AAAA
ATOM	1642	C	ASN	172	20.765	-2.644	71.842	1.00	33.14	AAAA	1811	OG1	THR	188	28.938	5.523	89.068	1.00	63.91	AAAA
ATOM	1643	O	ASN	172	21.208	-1.619	72.381	1.00	29.53	AAAA	1813	CG2	THR	188	30.048	7.525	89.960	1.00	62.56	AAAA
ATOM	1644	N	TYR	173	21.483	-3.738	71.619	1.00	33.30	AAAA	1814	C	THR	188	29.604	8.652	87.201	1.00	56.82	AAAA
ATOM	1646	CA	TYR	173	22.899	-3.835	71.930	1.00	34.05	AAAA	1815	O	THR	188	29.470	9.837	87.501	1.00	57.66	AAAA
ATOM	1647	CB	TYR	173	23.383	-5.249	71.721	1.00	31.77	AAAA	1816	N	CYS	189	30.508	8.196	86.337	1.00	53.62	AAAA
ATOM	1648	CG	TYR	173	22.759	-6.251	72.613	1.00	31.54	AAAA	1818	CA	CYS	189	31.467	9.028	85.639	1.00	49.24	AAAA
ATOM	1649	CD1	TYR	173	21.904	-7.206	72.107	1.00	31.55	AAAA	1819	C	CYS	189	30.896	10.316	85.098	1.00	48.89	AAAA
ATOM	1650	CE1	TYR	173	21.376	-8.179	72.912	1.00	35.13	AAAA	1820	O	CYS	189	31.542	11.356	85.151	1.00	51.71	AAAA
ATOM	1652	CE2	TYR	173	22.056	-9.283	73.981	1.00	33.45	AAAA	1821	CG	CYS	189	32.116	12.589	84.534	1.00	48.12	AAAA
ATOM	1652	CE2	TYR	173	22.056	-9.283	73.981	1.00	33.45	AAAA	1822	SG	CYS	189	31.375	7.087	85.322	1.00	46.39	AAAA
ATOM	1653	CZ	TYR	173	21.684	-8.205	74.253	1.00	37.83	AAAA	1823	N	GLY	190	29.681	10.259	84.582	1.00	46.39	AAAA
ATOM	1654	OH	TYR	173	21.140	-9.183	75.072	1.00	42.03	AAAA	1825	CA	GLY	190	29.068	11.468	84.070	1.00	42.95	AAAA
ATOM	1656	C	TYR	173	23.664	-2.964	70.958	1.00	36.13	AAAA	1826	C	GLY	190	29.612	11.844	82.724	1.00	40.11	AAAA
ATOM	1657	O	TYR	173	23.283	-2.895	69.781	1.00	38.28	AAAA	1827	O	GLY	190	29.877	11.002	81.884	1.00	41.46	AAAA
ATOM	1658	N	ARG	174	24.751	-2.338	71.426	1.00	35.38	AAAA	1828	N	LYS	191	29.778	13.122	82.502	1.00	39.74	AAAA
ATOM	1660	CA	ARG	174	25.569	-1.478	70.571	1.00	32.21	AAAA	1830	CA	LYS	191	30.300	13.539	81.228	1.00	42.54	AAAA
ATOM	1661	CB	ARG	174	25.498	-0.037	71.086	1.00	32.61	AAAA	1831	CB	LYS	191	29.919	15.002	80.951	1.00	43.99	AAAA
ATOM	1662	CG	ARG	174	24.061	1.507	71.300	1.00	33.85	AAAA	1832	CG	LYS	191	30.473	16.023	81.444	1.00	48.52	AAAA
ATOM	1663	CD	ARG	174	23.277	0.687	69.980	1.00	36.54	AAAA	1833	CD	LYS	191	29.851	15.877	83.357	1.00	53.24	AAAA
ATOM	1664	NE	ARG	174	21.819	0.818	70.138	1.00	37.12	AAAA	1834	CE	LYS	191	30.904	15.820	84.480	1.00	53.69	AAAA
ATOM	1666	CZ	ARG	174	20.913	0.542	69.287	1.00	37.83	AAAA	1835	NZ	LYS	191	30.465	14.897	85.544	1.00	51.30	AAAA
ATOM	1667	NH1	ARG	174	21.270	0.113	67.984	1.00	35.61	AAAA	1839	C	LYS	191	31.811	13.380	81.262	1.00	44.17	AAAA
ATOM	1670	NH2	ARG	174	19.623	0.682	69.443	1.00	42.36	AAAA	1840	O	LYS	191	32.529	14.285	80.837	1.00	48.34	AAAA
ATOM	1673	C	ARG	174	27.020	-1.986	70.509	1.00	31.04	AAAA	1841	N	ARG	192	32.303	12.242	81.757	1.00	42.01	AAAA
ATOM	1674	O	ARG	174	27.668	-2.167	71.521	1.00	31.23	AAAA	1843	CA	ARG	192	31.750	12.034	81.848	1.00	39.82	AAAA
ATOM	1675	N	CYS	175	27.508	-2.223	69.302	1.00	30.36	AAAA	1844	CB	ARG	192	34.209	11.987	83.294	1.00	43.32	AAAA
ATOM	1677	C	CYS	175	28.845	-2.716	68.967	1.00	34.48	AAAA	1845	CG	ARG	192	32.717	13.589	82.037	1.00	45.16	AAAA
ATOM	1678	C	CYS	175	29.398	-2.294	67.668	1.00	34.21	AAAA	1846	CD	ARG	192	34.785	13.211	85.334	1.00	55.04	AAAA
ATOM	1679	O	CYS	175	28.639	-1.800	66.806	1.00	30.35	AAAA	1847	NE	ARG	192	34.450	14.488	85.958	1.00	66.46	AAAA
ATOM	1680	CB	CYS	175	28.790	-4.254	69.197	1.00	31.73	AAAA	1849	CZ	ARG	192	34.973	14.911	87.109	1.00	70.26	AAAA
ATOM	1681	SG	CYS	175	27.867	-5.115	67.886	1.00	36.86	AAAA	1850	NH1	ARG	192	35.860	14.147	87.756	1.00	74.15	AAAA
ATOM	1682	N	TRP	176	30.713	-2.459	67.440	1.00	35.30	AAAA	1853	NH2	ARG	192	34.600	16.084	87.620	1.00	68.30	AAAA
ATOM	1684	CA	TRP	176	31.338	-2.029	66.166	1.00	32.69	AAAA	1856	C	ARG	192	34.176	10.756	81.211	1.00	37.62	AAAA
ATOM	1685	CB	TRP	176	32.700	-1.342	66.416	1.00	25.62	AAAA	1857	O	ARG	192	31.357	9.859	81.001	1.00	35.02	AAAA
ATOM	1686	CG	TRP	176	32.634	-0.056	67.947	1.00	19.80	AAAA	1858	N	ALA	193	35.471	10.674	80.917	1.00	35.49	AAAA
ATOM	1687	CD2	TRP	176	32.534	1.320	66.364	1.00	14.22	AAAA	1860	CA	ALA	193	36.022	9.481	80.293	1.00	34.93	AAAA
ATOM	1688	CE2																		

Figure 1 (cont'd)

ATOM	1940	CG	HIS	202	42.033	15.187	85.071	1.00	33.88	AAAA	ATOM	2097	CD	ARG	222	36.483	18.089	78.943	1.00	55.19	AAAA
ATOM	1941	CD2	HIS	202	42.478	15.717	86.230	1.00	34.57	AAAA	ATOM	2098	NE	ARG	222	36.491	16.622	79.070	1.00	59.31	AAAA
ATOM	1942	ND1	HIS	202	41.801	16.250	84.229	1.00	39.51	AAAA	ATOM	2100	CG	ARG	222	35.631	15.787	78.472	1.00	61.07	AAAA
ATOM	1944	CD1	HIS	202	42.084	17.379	84.856	1.00	38.52	AAAA	ATOM	2101	NH1	ARG	222	34.665	16.251	77.482	1.00	62.50	AAAA
ATOM	1945	NE2	HIS	202	42.496	17.082	86.073	1.00	34.70	AAAA	ATOM	2104	NH2	ARG	222	35.727	14.476	78.673	1.00	60.03	AAAA
ATOM	1947	C	HIS	202	39.332	14.312	84.100	1.00	35.68	AAAA	ATOM	2107	C	ARG	222	39.591	19.394	75.795	1.00	43.68	AAAA
ATOM	1948	O	HIS	202	39.123	14.067	82.918	1.00	40.37	AAAA	ATOM	2108	O	ARG	222	38.794	19.842	74.961	1.00	41.43	AAAA
ATOM	1949	N	PRO	203	38.750	15.340	84.710	1.00	34.78	AAAA	ATOM	2109	N	HIS	223	40.679	20.036	76.226	1.00	43.22	AAAA
ATOM	1950	CD	PRO	203	38.881	15.735	86.115	1.00	36.37	AAAA	ATOM	2111	CA	HIS	223	41.062	21.377	75.802	1.00	40.49	AAAA
ATOM	1951	CA	PRO	203	37.850	16.233	83.982	1.00	33.97	AAAA	ATOM	2112	CB	HIS	223	41.067	22.304	76.996	1.00	40.04	AAAA
ATOM	1952	CB	PRO	203	37.614	17.389	84.954	1.00	32.34	AAAA	ATOM	2113	CG	HIS	223	39.884	22.489	77.715	1.00	42.81	AAAA
ATOM	1953	CG	PRO	203	38.582	17.199	86.058	1.00	36.27	AAAA	ATOM	2115	CD2	HIS	223	38.923	21.602	78.054	1.00	43.38	AAAA
ATOM	1954	C	PRO	203	38.381	16.724	82.642	1.00	34.97	AAAA	ATOM	2117	CE1	HIS	223	39.412	23.726	78.656	1.00	44.77	AAAA
ATOM	1955	O	PRO	203	37.614	17.144	81.765	1.00	36.16	AAAA	ATOM	2118	NE2	HIS	223	38.242	23.594	78.656	1.00	45.50	AAAA
ATOM	1956	N	GLU	204	39.693	16.676	82.477	1.00	33.97	AAAA	ATOM	2120	C	HIS	223	42.188	21.418	75.113	1.00	43.06	AAAA
ATOM	1958	CA	GLU	204	40.290	17.148	81.242	1.00	31.62	AAAA	ATOM	2121	O	HIS	223	42.529	20.994	73.969	1.00	41.67	AAAA
ATOM	1959	CB	GLU	204	41.545	17.955	81.552	1.00	29.26	AAAA	ATOM	2122	N	TYR	224	43.386	21.939	75.843	1.00	40.87	AAAA
ATOM	1960	CG	GLU	204	41.235	19.253	82.260	1.00	30.14	AAAA	ATOM	2124	CA	TYR	224	44.740	22.109	75.351	1.00	39.54	AAAA
ATOM	1961	CD	GLU	204	40.739	20.368	81.331	1.00	34.01	AAAA	ATOM	2125	CB	TYR	224	45.119	23.562	75.465	1.00	38.87	AAAA
ATOM	1962	OE1	GLU	204	40.657	20.192	80.085	1.00	35.20	AAAA	ATOM	2126	CG	TYR	224	44.499	24.297	74.345	1.00	43.98	AAAA
ATOM	1963	OE2	GLU	204	40.434	21.445	81.868	1.00	35.07	AAAA	ATOM	2127	CD1	TYR	224	45.121	25.121	74.561	1.00	42.61	AAAA
ATOM	1964	C	GLU	204	40.603	16.003	80.299	1.00	30.09	AAAA	ATOM	2128	CE1	TYR	224	42.771	25.714	73.517	1.00	45.59	AAAA
ATOM	1965	O	GLU	204	40.585	16.151	79.082	1.00	30.45	AAAA	ATOM	2129	CD2	TYR	224	44.938	24.086	73.035	1.00	45.01	AAAA
ATOM	1966	N	CYS	205	40.865	14.835	80.840	1.00	29.97	AAAA	ATOM	2130	CE2	TYR	224	44.310	24.671	71.988	1.00	47.82	AAAA
ATOM	1968	CA	CYS	205	41.180	13.767	79.931	1.00	32.60	AAAA	ATOM	2131	CE2	TYR	224	43.218	25.491	72.231	1.00	48.72	AAAA
ATOM	1969	C	CYS	205	40.046	13.710	78.966	1.00	31.35	AAAA	ATOM	2132	OH	TYR	224	42.570	26.111	71.174	1.00	59.60	AAAA
ATOM	1970	O	CYS	205	38.908	13.801	79.362	1.00	35.49	AAAA	ATOM	2134	C	TYR	224	45.804	21.284	76.000	1.00	39.81	AAAA
ATOM	1971	CB	CYS	205	41.327	12.444	80.662	1.00	33.33	AAAA	ATOM	2135	O	TYR	224	46.017	21.370	77.202	1.00	45.49	AAAA
ATOM	1972	SC	CYS	205	42.981	11.223	81.401	1.00	38.25	AAAA	ATOM	2136	CH	TYR	225	46.488	20.492	75.186	1.00	39.39	AAAA
ATOM	1973	N	LEU	206	40.365	13.637	77.691	1.00	30.01	AAAA	ATOM	2138	CA	TYR	225	47.561	19.649	75.794	1.00	36.83	AAAA
ATOM	1975	CA	LEU	206	39.345	13.523	76.677	1.00	30.91	AAAA	ATOM	2139	CB	TYR	225	47.875	18.578	74.667	1.00	34.69	AAAA
ATOM	1976	CB	LEU	206	39.466	14.655	75.637	1.00	28.62	AAAA	ATOM	2140	CG	TYR	225	49.135	17.882	75.031	1.00	36.21	AAAA
ATOM	1977	CG	LEU	206	38.601	14.521	74.370	1.00	24.69	AAAA	ATOM	2141	CD1	TYR	225	49.118	16.785	75.878	1.00	35.50	AAAA
ATOM	1978	CD1	LEU	206	37.580	14.146	74.723	1.00	23.66	AAAA	ATOM	2142	CE1	TYR	225	50.249	16.097	76.166	1.00	37.02	AAAA
ATOM	1979	CD2	LEU	206	38.198	15.791	73.620	1.00	21.51	AAAA	ATOM	2143	CD2	TYR	225	50.335	18.282	74.487	1.00	38.38	AAAA
ATOM	1980	C	LEU	206	39.579	12.154	76.040	1.00	32.93	AAAA	ATOM	2144	CE2	TYR	225	51.480	17.606	74.762	1.00	42.33	AAAA
ATOM	1981	O	LEU	206	40.706	11.677	75.931	1.00	35.10	AAAA	ATOM	2145	CH	TYR	225	51.437	16.505	75.600	1.00	42.96	AAAA
ATOM	1982	GLY	207	38.510	11.488	75.650	1.00	36.31	AAAA	ATOM	2146	CH	TYR	225	52.600	15.796	75.933	1.00	51.09	AAAA	
ATOM	1984	CA	GLY	207	38.688	10.195	75.033	1.00	38.16	AAAA	ATOM	2148	C	TYR	225	49.781	20.484	75.794	1.00	37.42	AAAA
ATOM	1985	C	GLY	207	38.752	9.019	75.979	1.00	39.99	AAAA	ATOM	2149	O	TYR	225	49.024	21.345	74.979	1.00	39.91	AAAA
ATOM	1986	O	GLY	207	37.793	8.239	76.076	1.00	40.86	AAAA	ATOM	2150	N	TYR	226	49.567	20.215	76.821	1.00	40.71	AAAA
ATOM	1987	N	SER	208	39.870	8.880	76.684	1.00	38.15	AAAA	ATOM	2152	CA	TYR	226	50.757	20.995	77.055	1.00	41.82	AAAA
ATOM	1989	CA	SER	208	40.026	7.745	77.576	1.00	36.49	AAAA	ATOM	2153	CB	TYR	226	50.372	22.358	77.612	1.00	47.49	AAAA
ATOM	1990	CB	SER	208	40.194	6.471	76.745	1.00	36.46	AAAA	ATOM	2154	CG	TYR	226	51.564	23.143	78.074	1.00	56.26	AAAA
ATOM	1991	OG	SER	208	40.426	5.334	77.554	1.00	42.23	AAAA	ATOM	2155	CD1	TYR	226	52.096	22.947	79.344	1.00	61.49	AAAA
ATOM	1993	C	SER	208	41.245	7.990	78.419	1.00	36.06	AAAA	ATOM	2156	CE1	TYR	226	53.251	23.613	79.750	1.00	65.13	AAAA
ATOM	1994	N	SER	208	41.639	6.134	78.564	1.00	35.56	AAAA	ATOM	2157	CD2	TYR	226	52.210	20.210	77.215	1.00	61.56	AAAA
ATOM	1995	N	CYS	209	41.843	6.927	78.964	1.00	38.43	AAAA	ATOM	2158	CE2	TYR	226	53.365	24.706	77.604	1.00	64.51	AAAA
ATOM	1997	CA	CYS	209	43.029	7.065	79.809	1.00	43.02	AAAA	ATOM	2159	CG	TYR	226	53.882	24.489	78.876	1.00	65.38	AAAA
ATOM	1998	C	CYS	209	43.688	5.783	80.251	1.00	44.54	AAAA	ATOM	2160	OH	TYR	226	55.030	25.136	79.280	1.00	68.79	AAAA
ATOM	1999	O	CYS	209	43.236	4.582	79.939	1.00	45.60	AAAA	ATOM	2162	C	TYR	226	51.725	20.315	78.003	1.00	41.20	AAAA
ATOM	2000	CB	CYS	209	42.699	7.837	81.063	1.00	43.03	AAAA	ATOM	2163	O	TYR	226	51.392	20.007	79.141	1.00	42.16	AAAA
ATOM	2001	SG	CYS	209	40.927	7.852	81.449	1.00	52.23	AAAA	ATOM	2164	N	ALA	227	52.931	20.069	77.521	1.00	39.70	AAAA
ATOM	2002	N	SER	210	44.760	5.961	81.020	1.00	46.33	AAAA	ATOM	2166	CA	ALA	227	53.933	19.456	78.350	1.00	36.97	AAAA
ATOM	2004	CA	SER	210	45.553	4.858	81.565	1.00	46.93	AAAA	ATOM	2168	CB	ALA	227	54.354	20.423	79.387	1.00	38.99	AAAA
ATOM	2005	CB	SER	210	47.039	5.091	82.654	1.00	46.74	AAAA	ATOM	2168	CB	ALA	227	54.354	20.423	79.387	1.00	38.99	AAAA
ATOM	2006	OG	SER	210	47.443	6.393	81.659	1.00	47.68	AAAA	ATOM	2169	O	ALA	227	53.181	18.206	80.214	1.00	40.28	AAAA
ATOM	2008	C	SER	210	45.354	4.737	83.079	1.00	46.56	AAAA	ATOM	2170	N	GLY	228	53.219	17.160	78.265	1.00	38.53	AAAA
ATOM	2009	O	SER	210	45.485	3.656	83.660	1.00	46.51	AAAA	ATOM	2172	CA	GLY	228	52.708	15.952	78.873	1.00	41.07	AAAA
ATOM	2010	N	ALA	211	45.043	5.864	83.705	1.00	46.12	AAAA	ATOM	2173	C	GLY	228	51.434	16.146	79.690	1.00	43.45	AAAA
ATOM	2012	CA	ALA	211	44.831	5.915	85.139	1.00	46.73	AAAA	ATOM	2174	O	GLY	228	50.996	15.223	80.388	1.00	46.10	AAAA
ATOM	2013	CB	ALA	211	46.141	6.241	85.848	1.00	48.68	AAAA	ATOM	2175	N	VAL	229	50.800	17.310	79.602	1.00	41.21	AAAA
ATOM	2014	C	ALA	211	43.783	6.964	85.472	1.00	47.65	AAAA	ATOM	2177	CA	VAL	229	49.619	17.497	80.414	1.00	38.16	AAAA
ATOM	2015	O	ALA	211	43.684	8.016	84.824	1.00	49.87	AAAA	ATOM	2178	CB	VAL	229	49.981	18.276	81.648	1.00	38.46	AAAA
ATOM	2016	N	PRO	212	42.383	6.701	86.500	1.00	46.58												

Figure 1 (cont'd)

ATOM	2256	CG	TYR	239	51.005	31.971	72.517	1.00	27.41	AAAA	ATOM	2420	CA	ASN	254	60.669	29.024	63.833	1.00	41.51	AAAA
ATOM	2257	CD1	TYR	239	52.175	31.957	72.680	1.00	26.72	AAAA	ATOM	2421	CB	ASN	254	61.949	28.613	64.508	1.00	42.41	AAAA
ATOM	2258	CE1	TYR	239	53.015	31.016	73.260	1.00	26.83	AAAA	ATOM	2422	CG	ASN	254	62.825	29.791	64.764	1.00	45.51	AAAA
ATOM	2259	CD2	TYR	239	50.304	33.070	72.938	1.00	26.63	AAAA	ATOM	2423	OD1	ASN	254	63.493	30.120	63.846	1.00	48.66	AAAA
ATOM	2260	CE2	TYR	239	50.927	34.119	73.515	1.00	27.10	AAAA	ATOM	2424	ND2	ASN	254	62.817	30.249	66.008	1.00	45.81	AAAA
ATOM	2261	C	TYR	239	52.272	34.094	73.679	1.00	26.41	AAAA	ATOM	2427	C	ASN	254	59.966	27.820	63.261	1.00	44.77	AAAA
ATOM	2262	CH	TYR	239	52.855	35.161	74.301	1.00	31.09	AAAA	ATOM	2428	O	ASN	254	60.551	26.739	63.150	1.00	45.62	AAAA
ATOM	2264	C	TYR	239	50.166	32.416	72.616	1.00	26.09	AAAA	ATOM	2429	N	ILE	255	58.718	28.053	62.854	1.00	47.11	AAAA
ATOM	2265	O	TYR	239	49.793	27.662	72.091	1.00	34.50	AAAA	ATOM	2430	CB	ILE	255	57.827	27.950	62.285	1.00	44.95	AAAA
ATOM	2266	N	ARG	240	47.852	28.768	72.201	1.00	30.65	AAAA	ATOM	2432	CB	ILE	255	56.375	27.514	62.515	1.00	39.68	AAAA
ATOM	2268	CA	ARG	240	47.110	27.655	71.583	1.00	27.19	AAAA	ATOM	2433	CG2	ILE	255	55.585	27.423	61.248	1.00	43.91	AAAA
ATOM	2269	CB	ARG	240	45.626	27.940	71.580	1.00	25.69	AAAA	ATOM	2434	CG1	ILE	255	55.740	26.688	63.625	1.00	36.11	AAAA
ATOM	2270	CG	ARG	240	45.255	29.132	72.421	1.00	26.86	AAAA	ATOM	2435	CD1	ILE	255	54.400	27.238	64.112	1.00	32.00	AAAA
ATOM	2271	CD	ARG	240	43.771	29.143	72.648	1.00	29.42	AAAA	ATOM	2436	C	ILE	255	58.126	26.778	60.787	1.00	46.54	AAAA
ATOM	2272	NE	ARG	240	43.117	30.082	71.753	1.00	30.88	AAAA	ATOM	2437	O	ILE	255	58.351	25.631	60.393	1.00	48.08	AAAA
ATOM	2274	CZ	ARG	240	43.256	31.352	71.869	1.00	34.12	AAAA	ATOM	2438	N	LEU	256	58.143	27.819	59.960	1.00	48.83	AAAA
ATOM	2275	NH1	ARG	240	44.021	31.888	72.842	1.00	20.93	AAAA	ATOM	2440	CB	LEU	256	58.464	28.469	59.333	1.00	52.02	AAAA
ATOM	2276	NH2	ARG	240	42.985	32.197	71.011	1.00	33.99	AAAA	ATOM	2442	CG	LEU	256	57.827	28.797	57.721	1.00	48.70	AAAA
ATOM	2281	C	ARG	240	47.602	27.535	70.168	1.00	33.37	AAAA	ATOM	2443	CD1	LEU	256	56.004	30.443	57.261	1.00	46.78	AAAA
ATOM	2282	O	ARG	240	48.038	28.509	69.605	1.00	26.68	AAAA	ATOM	2444	CD2	LEU	256	55.612	27.990	56.935	1.00	49.57	AAAA
ATOM	2283	N	PHE	241	47.538	26.357	69.583	1.00	20.36	AAAA	ATOM	2445	C	LEU	256	59.981	27.997	58.339	1.00	54.86	AAAA
ATOM	2285	CA	PHE	241	48.034	26.193	68.232	1.00	19.13	AAAA	ATOM	2446	O	LEU	256	60.483	27.600	57.235	1.00	55.37	AAAA
ATOM	2286	CB	PHE	241	49.574	26.040	68.294	1.00	20.49	AAAA	ATOM	2447	N	SER	257	60.706	27.505	59.431	1.00	57.82	AAAA
ATOM	2287	CG	PHE	241	50.214	25.654	66.985	1.00	15.08	AAAA	ATOM	2449	CA	SER	257	62.155	27.536	59.421	1.00	61.01	AAAA
ATOM	2288	CD1	PHE	241	50.346	26.569	65.967	1.00	17.58	AAAA	ATOM	2450	CB	SER	257	62.705	27.201	60.815	1.00	61.83	AAAA
ATOM	2289	CD2	PHE	241	50.620	24.367	66.759	1.00	16.72	AAAA	ATOM	2451	CG	SER	257	62.038	26.206	55.577	1.00	70.65	AAAA
ATOM	2290	CE1	PHE	241	50.868	26.193	64.732	1.00	20.68	AAAA	ATOM	2453	C	SER	257	62.814	26.636	58.398	1.00	63.24	AAAA
ATOM	2291	CE2	PHE	241	51.144	23.984	65.529	1.00	20.21	AAAA	ATOM	2454	O	SER	257	63.828	27.006	57.818	1.00	66.36	AAAA
ATOM	2292	CZ	PHE	241	51.266	24.899	64.516	1.00	15.78	AAAA	ATOM	2455	N	ALA	258	62.276	25.452	58.165	1.00	65.73	AAAA
ATOM	2293	C	PHE	241	47.369	24.962	67.601	1.00	22.49	AAAA	ATOM	2457	CA	ALA	258	62.928	24.588	57.191	1.00	68.76	AAAA
ATOM	2294	O	PHE	241	47.498	23.845	68.137	1.00	22.07	AAAA	ATOM	2458	CB	ALA	258	62.517	23.131	57.414	1.00	69.08	AAAA
ATOM	2295	N	GLU	242	46.563	25.164	66.479	1.00	21.89	AAAA	ATOM	2459	C	ALA	258	62.633	25.021	55.749	1.00	70.99	AAAA
ATOM	2297	CA	GLU	242	45.952	24.075	65.766	1.00	21.22	AAAA	ATOM	2460	CB	GLU	258	62.947	24.288	54.802	1.00	74.18	AAAA
ATOM	2298	CB	GLU	242	46.918	22.913	65.439	1.00	25.33	AAAA	ATOM	2461	N	GLU	259	62.038	26.206	55.577	1.00	70.65	AAAA
ATOM	2299	CG	GLU	242	47.783	23.045	64.149	1.00	25.74	AAAA	ATOM	2463	CA	GLU	259	61.727	26.708	54.237	1.00	68.39	AAAA
ATOM	2300	CD	GLU	242	47.099	23.768	63.007	1.00	28.55	AAAA	ATOM	2464	CB	GLU	259	60.268	26.445	53.876	1.00	69.57	AAAA
ATOM	2301	OE1	GLU	242	46.182	23.192	62.388	1.00	36.80	AAAA	ATOM	2465	CG	GLU	259	59.667	25.233	54.529	1.00	70.62	AAAA
ATOM	2302	OE2	GLU	242	47.471	24.916	62.710	1.00	28.23	AAAA	ATOM	2466	CD	GLU	259	58.270	25.512	55.022	1.00	74.23	AAAA
ATOM	2303	C	GLU	242	44.801	23.573	66.639	1.00	20.26	AAAA	ATOM	2467	OE1	GLU	259	57.651	26.472	54.495	1.00	73.42	AAAA
ATOM	2304	O	GLU	242	44.369	22.426	66.573	1.00	14.94	AAAA	ATOM	2468	OE2	GLU	259	57.797	24.779	55.930	1.00	76.56	AAAA
ATOM	2305	N	GLY	243	44.303	24.688	67.455	1.00	25.09	AAAA	ATOM	2469	C	GLU	259	62.025	28.177	53.990	1.00	66.31	AAAA
ATOM	2307	CA	GLY	243	43.214	24.393	68.345	1.00	26.25	AAAA	ATOM	2470	O	GLU	259	61.764	29.056	54.808	1.00	63.93	AAAA
ATOM	2308	C	GLY	243	43.355	22.887	69.056	1.00	37.15	AAAA	ATOM	2471	N	SER	260	65.565	28.409	52.910	1.00	67.17	AAAA
ATOM	2309	O	GLY	243	42.576	22.005	68.900	1.00	35.16	AAAA	ATOM	2473	CA	SER	260	62.927	29.733	52.345	1.00	68.69	AAAA
ATOM	2310	N	TRP	244	44.508	22.741	69.781	1.00	27.71	AAAA	ATOM	2474	CB	SER	260	63.983	29.606	51.221	1.00	71.38	AAAA
ATOM	2312	CA	TRP	244	44.782	21.512	70.491	1.00	27.68	AAAA	ATOM	2475	OG	SER	260	64.829	28.470	51.396	1.00	71.87	AAAA
ATOM	2313	CB	TRP	244	44.872	20.299	69.536	1.00	29.29	AAAA	ATOM	2477	C	SER	260	61.673	30.448	51.814	1.00	66.68	AAAA
ATOM	2314	CG	TRP	244	46.277	19.813	69.029	1.00	32.96	AAAA	ATOM	2478	O	SER	260	61.446	30.494	50.598	1.00	66.89	AAAA
ATOM	2315	CD2	TRP	244	47.164	18.860	69.666	1.00	30.71	AAAA	ATOM	2479	N	SER	261	60.872	31.013	52.719	1.00	64.98	AAAA
ATOM	2316	CE2	TRP	244	48.229	18.610	68.771	1.00	28.73	AAAA	ATOM	2481	CA	SER	261	59.633	31.704	52.325	1.00	60.28	AAAA
ATOM	2317	CE3	TRP	244	47.255	18.197	70.897	1.00	29.85	AAAA	ATOM	2482	CB	SER	261	58.468	31.259	52.910	1.00	58.86	AAAA
ATOM	2318	CD1	TRP	244	46.358	20.089	67.795	1.00	32.87	AAAA	ATOM	2483	OG	SER	261	57.857	30.098	52.687	1.00	60.47	AAAA
ATOM	2319	NE1	TRP	244	48.019	19.367	67.646	1.00	31.29	AAAA	ATOM	2485	C	SER	261	59.648	33.229	52.300	1.00	57.08	AAAA
ATOM	2321	CD2	TRP	244	49.260	17.735	69.074	1.00	27.63	AAAA	ATOM	2486	O	SER	261	59.867	33.878	53.319	1.00	54.48	AAAA
ATOM	2322	CD3	TRP	244	48.185	17.330	71.187	1.00	25.36	AAAA	ATOM	2487	N	ASP	262	59.395	33.775	51.110	1.00	55.02	AAAA
ATOM	2323	CH2	TRP	244	49.218	17.109	70.283	1.00	27.26	AAAA	ATOM	2489	CA	ASP	262	59.321	35.213	50.865	1.00	51.51	AAAA
ATOM	2324	C	TRP	244	45.992	21.523	71.338	1.00	27.89	AAAA	ATOM	2490	CB	ASP	262	59.256	35.469	49.364	1.00	57.33	AAAA
ATOM	2325	O	TRP	244	46.263	20.584	72.091	1.00	31.36	AAAA	ATOM	2491	CG	ASP	262	58.007	34.854	48.723	1.00	62.17	AAAA
ATOM	2326	N	ARG	245	45.831	22.546	71.211	1.00	22.84	AAAA	ATOM	2492	CD1	ASP	262	57.747	33.658	48.925	1.00	64.35	AAAA
ATOM	2328	CA	ARG	245	45.264	22.662	72.013	1.00	20.57	AAAA	ATOM	2493	OD2	ASP	262	57.287	35.564	47.973	1.00	68.42	AAAA
ATOM	2329	CB	ARG	245	49.103	21.677	71.356	1.00	22.33	AAAA	ATOM	2494	C	ASP	262	58.040	35.770	51.513	1.00	64.42	AAAA
ATOM	2330	CG	ARG	245	49.986	22.373	70.328	1.00	24.79	AAAA	ATOM	2495	O	ASP	262	57.778	36.985	51.485	1.00	46.29	AAAA
ATOM	2331	CD	ARG	245	51.678	21.398	69.848	1.00	24.78	AAAA	ATOM	2496	N	SER	263	57.233	34.876	52.075	1.00	42.62	AAAA
ATOM	2332	NE	ARG	245	52.003	21.923	68.948	1.00	28.79	AAAA	ATOM	2498	CA	SER	263	56.009	35.292	52.713	1.00	37.81	AAAA
ATOM	2334	CZ	ARG	245	52.494	21.179															

Figure 1 (cont'd)

ATOM	2578	CB	GLU	272	45.823	32.211	68.965	1.00	27.11	AAAA	ATOM	2739	O	MET	289	52.503	38.544	65.931	1.00	28.48	AAAA
ATOM	2579	CG	GLU	272	45.843	31.463	67.702	1.00	33.51	AAAA	ATOM	2740	N	TYR	290	53.445	40.597	65.744	1.00	28.07	AAAA
ATOM	2580	CD	GLU	272	44.495	30.900	67.390	1.00	37.10	AAAA	ATOM	2742	CA	TYR	290	52.377	41.409	66.314	1.00	28.86	AAAA
ATOM	2581	OE1	GLU	272	43.512	31.466	67.935	1.00	40.77	AAAA	ATOM	2743	CB	TYR	290	52.989	42.558	67.114	1.00	28.43	AAAA
ATOM	2582	OE2	GLU	272	44.426	29.906	66.617	1.00	34.64	AAAA	ATOM	2744	CG	TYR	290	53.700	42.094	68.358	1.00	28.56	AAAA
ATOM	2583	C	GLU	272	48.179	32.461	68.435	1.00	26.58	AAAA	ATOM	2745	CD1	TYR	290	55.046	41.761	68.336	1.00	26.85	AAAA
ATOM	2584	O	GLU	272	48.243	33.488	67.764	1.00	28.05	AAAA	ATOM	2746	CE1	TYR	290	55.681	41.334	69.466	1.00	23.77	AAAA
ATOM	2585	C	CYS	273	48.932	31.388	67.487	1.00	27.05	AAAA	ATOM	2747	CD2	TYR	290	53.601	41.012	68.600	1.00	23.41	AAAA
ATOM	2586	CA	CYS	273	49.923	31.301	67.165	1.00	27.37	AAAA	ATOM	2748	CE2	TYR	290	53.650	41.513	70.696	1.00	29.09	AAAA
ATOM	2587	C	CYS	273	49.202	30.847	65.897	1.00	29.56	AAAA	ATOM	2749	CZ	TYR	290	54.976	41.196	70.644	1.00	28.46	AAAA
ATOM	2588	O	CYS	273	48.665	29.717	65.816	1.00	27.40	AAAA	ATOM	2750	OH	TYR	290	55.604	40.788	71.794	1.00	34.58	AAAA
ATOM	2589	CB	CYS	273	51.022	30.284	67.516	1.00	26.43	AAAA	ATOM	2752	C	TYR	290	51.395	41.982	65.281	1.00	30.98	AAAA
ATOM	2590	SG	CYS	273	52.197	29.921	66.166	1.00	26.84	AAAA	ATOM	2753	O	TYR	290	51.783	42.644	64.298	1.00	27.04	AAAA
ATOM	2591	C	GLU	274	49.187	31.744	64.914	1.00	29.52	AAAA	ATOM	2754	N	CYS	291	50.116	41.742	65.560	1.00	22.25	AAAA
ATOM	2592	CA	MET	274	48.562	31.487	63.637	1.00	28.78	AAAA	ATOM	2756	CA	CYS	291	49.019	42.158	64.708	1.00	33.41	AAAA
ATOM	2593	CB	MET	274	47.160	32.129	63.586	1.00	30.44	AAAA	ATOM	2757	C	CYS	291	48.287	43.394	65.201	1.00	34.88	AAAA
ATOM	2594	CD	MET	274	47.028	31.654	63.820	1.00	34.33	AAAA	ATOM	2758	O	CYS	291	47.753	43.390	66.314	1.00	36.19	AAAA
ATOM	2595	SD	MET	274	45.573	34.073	64.843	1.00	33.21	AAAA	ATOM	2759	CB	CYS	291	48.007	41.025	64.623	1.00	34.25	AAAA
ATOM	2596	CE	MET	274	45.773	35.798	65.222	1.00	26.46	AAAA	ATOM	2760	SG	CYS	291	48.552	39.512	63.781	1.00	34.00	AAAA
ATOM	2597	C	MET	274	49.456	32.084	62.573	1.00	29.64	AAAA	ATOM	2761	N	ILE	292	48.228	44.446	64.397	1.00	26.12	AAAA
ATOM	2598	O	MET	274	50.165	33.038	62.815	1.00	31.59	AAAA	ATOM	2763	CA	ILE	292	47.485	45.645	64.792	1.00	38.93	AAAA
ATOM	2601	N	GLN	275	49.416	31.506	61.387	1.00	32.42	AAAA	ATOM	2764	CB	ILE	292	48.337	46.955	64.760	1.00	36.37	AAAA
ATOM	2603	CA	GLN	275	50.195	31.980	60.257	1.00	30.77	AAAA	ATOM	2765	CG2	ILE	292	49.341	46.976	65.887	1.00	33.01	AAAA
ATOM	2604	CB	GLN	275	50.110	30.968	59.109	1.00	36.00	AAAA	ATOM	2766	CG1	ILE	292	47.005	47.100	63.403	1.00	35.12	AAAA
ATOM	2605	CG	GLN	275	49.036	31.070	58.072	1.00	35.09	AAAA	ATOM	2767	CD1	ILE	292	49.331	48.009	63.079	1.00	32.69	AAAA
ATOM	2606	CD	GLN	275	49.158	30.416	56.816	1.00	43.67	AAAA	ATOM	2768	C	ILE	292	46.278	45.878	63.873	1.00	41.13	AAAA
ATOM	2607	OE1	GLN	275	49.623	29.266	56.864	1.00	43.42	AAAA	ATOM	2769	O	ILE	292	46.240	45.409	62.731	1.00	39.78	AAAA
ATOM	2608	NE2	GLN	275	48.737	30.978	55.676	1.00	45.11	AAAA	ATOM	2770	N	PRO	293	45.255	46.577	64.395	1.00	43.88	AAAA
ATOM	2611	C	GLN	275	49.796	33.333	59.720	1.00	27.95	AAAA	ATOM	2771	CD	PRO	293	45.175	47.032	65.791	1.00	45.83	AAAA
ATOM	2612	O	GLN	275	50.585	33.969	59.041	1.00	30.85	AAAA	ATOM	2772	CA	PRO	293	44.026	46.912	63.679	1.00	44.79	AAAA
ATOM	2613	N	GLU	276	48.583	33.779	60.000	1.00	29.24	AAAA	ATOM	2773	CB	PRO	293	43.135	47.546	64.754	1.00	43.03	AAAA
ATOM	2615	CA	GLU	276	48.129	35.061	59.478	1.00	31.66	AAAA	ATOM	2774	CG	PRO	293	43.686	47.101	66.025	1.00	45.02	AAAA
ATOM	2616	CB	GLU	276	47.239	34.818	58.243	1.00	34.69	AAAA	ATOM	2775	N	GLY	293	42.918	47.871	62.533	1.00	45.57	AAAA
ATOM	2617	CG	GLU	276	47.886	34.037	57.089	1.00	42.81	AAAA	ATOM	2776	O	PRO	293	45.081	48.782	62.622	1.00	47.35	AAAA
ATOM	2618	CD	GLU	276	48.801	34.898	56.183	1.00	49.14	AAAA	ATOM	2777	N	CYS	294	43.492	47.660	61.470	1.00	46.89	AAAA
ATOM	2619	OE1	GLU	276	48.924	36.124	56.452	1.00	49.17	AAAA	ATOM	2779	CA	CYS	294	43.539	48.481	60.275	1.00	49.04	AAAA
ATOM	2620	OE2	GLU	276	49.400	34.345	55.211	1.00	50.81	AAAA	ATOM	2780	C	CYS	294	42.490	49.582	60.346	1.00	53.79	AAAA
ATOM	2621	C	GLU	276	47.352	35.897	60.499	1.00	29.62	AAAA	ATOM	2781	O	CYS	294	41.280	49.282	60.359	1.00	57.25	AAAA
ATOM	2622	O	GLU	276	46.600	35.359	61.286	1.00	34.65	AAAA	ATOM	2782	CB	CYS	294	43.197	47.642	59.667	1.00	43.85	AAAA
ATOM	2623	N	CYS	277	47.513	37.209	60.488	1.00	28.03	AAAA	ATOM	2783	SG	CYS	294	44.523	46.608	58.442	1.00	39.70	AAAA
ATOM	2625	CA	CYS	277	46.773	38.035	61.419	1.00	32.17	AAAA	ATOM	2784	CG	CYS	294	42.918	47.871	62.533	1.00	45.57	AAAA
ATOM	2626	C	CYS	277	47.555	37.280	60.853	1.00	31.73	AAAA	ATOM	2785	CA	GLU	295	41.721	49.641	59.641	1.00	50.64	AAAA
ATOM	2627	O	CYS	277	44.871	37.405	60.112	1.00	40.78	AAAA	ATOM	2786	CB	GLU	295	42.610	53.279	60.393	1.00	58.06	AAAA
ATOM	2628	CB	CYS	277	47.060	39.492	61.177	1.00	34.42	AAAA	ATOM	2787	C	GLU	295	41.346	51.818	58.993	1.00	59.41	AAAA
ATOM	2629	SG	CYS	277	48.580	40.135	61.867	1.00	37.60	AAAA	ATOM	2788	O	GLU	295	41.976	52.216	58.005	1.00	62.05	AAAA
ATOM	2630	N	PRO	278	44.440	38.232	62.141	1.00	37.49	AAAA	ATOM	2790	N	GLY	296	40.165	51.218	58.903	1.00	58.10	AAAA
ATOM	2631	CD	PRO	278	44.717	38.787	63.475	1.00	37.99	AAAA	ATOM	2792	CA	GLY	296	39.529	52.069	57.609	1.00	55.31	AAAA
ATOM	2632	CA	PRO	278	43.011	38.077	61.908	1.00	37.70	AAAA	ATOM	2793	C	GLY	296	39.932	49.815	56.870	1.00	53.05	AAAA
ATOM	2633	CB	PRO	278	42.412	38.332	63.276	1.00	38.57	AAAA	ATOM	2794	O	GLY	296	40.566	49.936	57.456	1.00	52.72	AAAA
ATOM	2634	CG	PRO	278	42.173	39.232	63.943	1.00	36.22	AAAA	ATOM	2795	N	GLY	297	39.566	49.700	55.576	1.00	55.49	AAAA
ATOM	2635	C	PRO	278	42.609	39.151	60.915	1.00	40.24	AAAA	ATOM	2796	CD	PRO	297	38.828	50.681	54.766	1.00	50.86	AAAA
ATOM	2636	O	PRO	278	43.221	40.218	60.892	1.00	43.77	AAAA	ATOM	2797	CA	PRO	297	39.920	48.515	54.799	1.00	48.69	AAAA
ATOM	2637	N	SER	279	41.592	38.885	60.100	1.00	41.09	AAAA	ATOM	2798	CB	PRO	297	39.409	48.824	53.403	1.00	48.80	AAAA
ATOM	2639	CA	SER	279	41.128	39.861	59.109	1.00	41.70	AAAA	ATOM	2799	CG	PRO	297	38.356	49.865	53.611	1.00	51.79	AAAA
ATOM	2640	CB	SER	279	39.823	39.407	58.480	1.00	38.80	AAAA	ATOM	2800	C	PRO	297	41.416	48.431	54.849	1.00	48.25	AAAA
ATOM	2641	CG	SER	279	39.229	38.420	59.303	1.00	45.46	AAAA	ATOM	2801	O	PRO	297	42.115	49.423	54.607	1.00	47.82	AAAA
ATOM	2643	C	SER	279	40.886	41.162	59.820	1.00	42.34	AAAA	ATOM	2802	N	CYS	298	41.902	47.250	55.204	1.00	47.11	AAAA
ATOM	2644	O	SER	279	40.358	41.169	60.925	1.00	44.59	AAAA	ATOM	2804	CA	CYS	298	40.326	47.042	55.326	1.00	42.12	AAAA
ATOM	2645	N	GLY	280	41.272	42.263	59.187	1.00	42.13	AAAA	ATOM	2805	C	CYS	298	43.974	47.392	54.025	1.00	39.91	AAAA
ATOM	2647	CA	GLY	280	41.061	43.548	59.828	1.00	42.52	AAAA	ATOM	2806	O	CYS	298	43.397	47.184	52.959	1.00	41.44	AAAA
ATOM	2648	C	GLY	280	42.340	43.995	60.491	1.00	45.15	AAAA	ATOM	2807	CB	CYS	298	43.633	45.599	55.679	1.00	40.84	AAAA
ATOM	2649	O	GLY	280	42.490	45.185	60.810	1.00	46.25	AAAA	ATOM	2808	SG	CYS	298	43.470	45.212	57.443	1.00	39.73	AAAA
ATOM	2650	N	PHE	281	43.253	43.037	60.701	1.00	43.64	AAAA	ATOM	2809	N	PRO	299	45.210	47.894	54.094	1.00	36.17	AAAA
ATOM	2652	CA	PHE	281	44.559	43.317	61.304	1.00	39.52	AAAA	ATOM	2810	CD	PRO	299	46.030	48.054	55.299	1.00	34.26	AAAA
ATOM	2653	CB	PHE	281	44.802	42.442	62.515	1.00	39.18												

Figure 1 (cont'd)

ATOM	2895	CG1	THR	308	49.459	48.972	30.476	1.00	38.44	AAAA	ATOM	3058	CE1	PHE	326	46.546	46.196	38.844	1.00	14.79	AAAA
ATOM	2897	CG2	THR	308	47.095	48.443	30.419	1.00	33.84	AAAA	ATOM	3060	CE2	PHE	326	47.334	47.984	37.480	1.00	22.55	AAAA
ATOM	2898	C	THR	308	47.326	46.497	32.350	1.00	43.89	AAAA	ATOM	3062	CZ	PHE	326	47.090	46.628	37.677	1.00	16.82	AAAA
ATOM	2899	C	THR	308	47.192	45.578	31.534	1.00	45.00	AAAA	ATOM	3063	O	PHE	326	48.694	49.624	41.394	1.00	34.23	AAAA
ATOM	2900	N	LYS	309	46.486	46.711	33.353	1.00	43.85	AAAA	ATOM	3064	N	LYS	327	48.977	50.772	41.342	1.00	35.86	AAAA
ATOM	2901	CA	LYS	309	45.114	45.878	33.516	1.00	43.17	AAAA	ATOM	3065	CA	LYS	327	49.580	48.627	41.108	1.00	36.39	AAAA
ATOM	2902	CB	LYS	309	44.746	46.006	34.934	1.00	42.67	AAAA	ATOM	3066	CB	LYS	327	50.912	48.866	40.834	1.00	35.70	AAAA
ATOM	2903	CD	LYS	309	43.345	45.424	35.134	1.00	41.64	AAAA	ATOM	3067	CD	LYS	327	51.961	48.109	41.646	1.00	38.39	AAAA
ATOM	2904	CE	LYS	309	43.359	43.901	35.155	1.00	41.62	AAAA	ATOM	3068	CE	LYS	327	53.271	48.875	41.789	1.00	36.14	AAAA
ATOM	2905	CG	LYS	309	42.623	43.339	36.369	1.00	41.29	AAAA	ATOM	3069	CG	LYS	327	54.426	48.009	41.359	1.00	40.36	AAAA
ATOM	2906	CH	LYS	309	43.106	41.972	36.731	1.00	40.17	AAAA	ATOM	3070	CH	LYS	327	55.468	48.754	40.515	1.00	43.06	AAAA
ATOM	2907	NZ	LYS	309	44.383	46.513	32.512	1.00	41.76	AAAA	ATOM	3071	NZ	LYS	327	56.405	47.780	39.847	1.00	42.45	AAAA
ATOM	2911	C	LYS	309	44.230	47.735	32.497	1.00	41.14	AAAA	ATOM	3072	C	LYS	327	50.894	48.402	39.393	1.00	34.84	AAAA
ATOM	2912	C	LYS	309	43.806	45.694	31.645	1.00	41.60	AAAA	ATOM	3073	O	LYS	327	51.071	47.225	39.024	1.00	38.47	AAAA
ATOM	2913	CA	THR	310	42.879	46.196	30.643	1.00	42.53	AAAA	ATOM	3074	CA	LYS	328	50.641	49.357	38.524	1.00	31.65	AAAA
ATOM	2916	CB	THR	310	43.210	45.636	29.267	1.00	45.37	AAAA	ATOM	3075	CB	LYS	328	50.577	49.070	37.120	1.00	30.16	AAAA
ATOM	2917	CG1	THR	310	42.177	45.981	28.144	1.00	50.14	AAAA	ATOM	3076	CG1	THR	328	49.819	50.207	36.493	1.00	29.94	AAAA
ATOM	2919	CG2	THR	310	43.353	44.123	29.342	1.00	51.13	AAAA	ATOM	3077	O	GLY	328	49.781	51.325	37.003	1.00	29.12	AAAA
ATOM	2920	C	THR	310	42.475	45.781	31.041	1.00	38.93	AAAA	ATOM	3078	C	GLY	328	49.201	49.914	35.374	1.00	31.22	AAAA
ATOM	2921	C	THR	310	41.013	44.701	30.717	1.00	40.54	AAAA	ATOM	3079	CA	ASN	329	48.445	50.909	34.682	1.00	32.69	AAAA
ATOM	2922	N	ILE	311	40.821	46.659	31.775	1.00	36.14	AAAA	ATOM	3080	CB	ASN	329	49.107	51.188	33.336	1.00	32.90	AAAA
ATOM	2924	CA	ILE	311	39.483	46.452	32.276	1.00	32.23	AAAA	ATOM	3081	CB	ASN	329	50.614	51.466	33.459	1.00	31.09	AAAA
ATOM	2925	CB	ILE	311	39.262	47.395	33.443	1.00	33.40	AAAA	ATOM	3082	CD	ASN	329	51.058	52.197	34.350	1.00	28.29	AAAA
ATOM	2926	CG2	ILE	311	37.807	47.630	33.625	1.00	37.51	AAAA	ATOM	3083	CD	ASN	329	51.396	50.891	32.477	1.00	27.03	AAAA
ATOM	2927	CG1	ILE	311	39.940	46.853	34.701	1.00	28.43	AAAA	ATOM	3084	OD1	ASN	329	47.036	50.356	34.500	1.00	26.25	AAAA
ATOM	2928	CG1	ILE	311	37.770	47.775	35.890	1.00	27.87	AAAA	ATOM	3085	OD1	ASN	329	46.805	49.134	34.580	1.00	37.16	AAAA
ATOM	2929	C	ILE	311	38.527	46.786	31.142	1.00	32.17	AAAA	ATOM	3086	OD1	ASN	329	46.091	51.264	34.287	1.00	37.34	AAAA
ATOM	2930	O	ILE	311	38.463	47.916	30.689	1.00	27.60	AAAA	ATOM	3087	ND2	ASN	329	44.703	50.889	34.055	1.00	35.63	AAAA
ATOM	2931	N	ASP	312	37.790	45.789	30.681	1.00	36.21	AAAA	ATOM	3088	ND2	ASN	329	43.786	51.493	35.124	1.00	32.90	AAAA
ATOM	2933	CA	ASP	312	36.871	45.960	29.560	1.00	41.14	AAAA	ATOM	3089	CD1	LEU	330	43.134	51.789	37.518	1.00	33.50	AAAA
ATOM	2934	CB	ASP	312	37.416	45.228	28.330	1.00	43.85	AAAA	ATOM	3090	CD1	LEU	330	43.000	49.552	36.438	1.00	28.87	AAAA
ATOM	2935	CG	ASP	312	37.858	43.802	28.639	1.00	48.16	AAAA	ATOM	3091	CD1	LEU	330	44.291	51.117	32.692	1.00	35.17	AAAA
ATOM	2936	OD1	ASP	312	38.400	43.133	27.734	1.00	49.21	AAAA	ATOM	3092	O	LEU	330	44.554	52.583	32.342	1.00	37.46	AAAA
ATOM	2937	OD2	ASP	312	37.667	43.345	29.786	1.00	49.33	AAAA	ATOM	3093	O	LEU	330	43.773	50.583	31.898	1.00	34.64	AAAA
ATOM	2938	ASP	312	35.485	45.434	29.241	1.00	44.48	AAAA	ATOM	3094	CA	LEU	331	43.176	51.039	30.621	1.00	34.59	AAAA	
ATOM	2939	O	ASP	312	34.531	45.593	29.241	1.00	44.48	AAAA	ATOM	3095	CB	LEU	331	43.938	50.397	29.469	1.00	33.32	AAAA
ATOM	2940	N	SER	313	35.389	44.775	31.026	1.00	44.15	AAAA	ATOM	3096	CB	LEU	331	43.049	50.089	28.277	1.00	28.49	AAAA
ATOM	2942	CA	SER	313	34.125	44.264	31.476	1.00	44.15	AAAA	ATOM	3097	CD1	LEU	331	43.233	51.111	27.218	1.00	27.07	AAAA
ATOM	2943	CB	SER	313	34.114	42.746	31.390	1.00	44.77	AAAA	ATOM	3098	CD2	LEU	331	43.364	48.720	27.787	1.00	31.73	AAAA
ATOM	2944	OG	SER	313	35.007	42.177	32.332	1.00	49.37	AAAA	ATOM	3099	CD2	LEU	331	41.746	50.536	30.649	1.00	35.56	AAAA
ATOM	2946	C	SER	313	34.046	44.735	32.913	1.00	45.82	AAAA	ATOM	3100	O	LEU	331	41.511	49.399	30.389	1.00	38.42	AAAA
ATOM	2947	O	SER	313	34.702	45.696	33.291	1.00	48.10	AAAA	ATOM	3101	N	LEU	331	40.805	51.407	31.027	1.00	32.64	AAAA
ATOM	2948	N	VAL	314	33.239	44.060	33.713	1.00	48.85	AAAA	ATOM	3102	N	LEU	331	39.406	51.011	31.077	1.00	32.18	AAAA
ATOM	2950	CA	VAL	314	33.053	44.399	35.110	1.00	48.19	AAAA	ATOM	3103	CA	LEU	331	38.594	51.845	32.098	1.00	30.81	AAAA
ATOM	2951	CB	VAL	314	31.613	44.768	35.359	1.00	47.19	AAAA	ATOM	3104	CB	LEU	331	38.770	51.272	33.479	1.00	29.20	AAAA
ATOM	2952	CG1	VAL	314	30.801	43.504	35.379	1.00	46.68	AAAA	ATOM	3105	CG1	LEU	331	39.078	53.294	32.100	1.00	35.82	AAAA
ATOM	2953	CG2	VAL	314	31.467	45.548	36.441	1.00	46.68	AAAA	ATOM	3106	CD1	LEU	331	39.305	53.907	33.505	1.00	36.84	AAAA
ATOM	2954	C	VAL	314	33.346	43.097	35.806	1.00	50.62	AAAA	ATOM	3107	CD2	LEU	331	38.914	51.233	29.668	1.00	31.94	AAAA
ATOM	2955	O	VAL	314	33.328	43.002	37.022	1.00	53.04	AAAA	ATOM	3108	O	LEU	331	38.988	52.331	29.146	1.00	35.38	AAAA
ATOM	2956	N	THR	315	33.550	42.072	34.993	1.00	54.43	AAAA	ATOM	3109	C	LEU	331	38.446	50.168	29.045	1.00	32.73	AAAA
ATOM	2958	CA	THR	315	33.899	40.754	35.487	1.00	58.60	AAAA	ATOM	3110	N	ASN	333	37.991	50.209	27.673	1.00	35.72	AAAA
ATOM	2959	CB	THR	315	33.176	39.576	34.718	1.00	61.24	AAAA	ATOM	3111	CB	ASN	333	38.981	49.379	26.838	1.00	36.08	AAAA
ATOM	2960	CG1	THR	315	33.404	39.670	33.303	1.00	63.54	AAAA	ATOM	3112	CG	ASN	333	38.747	49.474	25.350	1.00	37.77	AAAA
ATOM	2962	CG2	THR	315	31.675	39.604	34.989	1.00	65.53	AAAA	ATOM	3113	OD1	ASN	333	39.422	50.224	24.653	1.00	38.86	AAAA
ATOM	2963	C	THR	315	35.397	40.679	35.244	1.00	60.77	AAAA	ATOM	3114	OD1	ASN	333	37.808	48.693	24.846	1.00	40.58	AAAA
ATOM	2964	O	THR	315	36.246	39.696	35.600	1.00	60.00	AAAA	ATOM	3115	ND2	ASN	333	36.611	49.569	27.667	1.00	39.80	AAAA
ATOM	2965	N	SER	316	35.341	41.725	34.625	1.00	55.55	AAAA	ATOM	3116	ND2	ASN	333	35.431	48.543	27.023	1.00	41.48	AAAA
ATOM	2967	CA	SER	316	37.370	41.764	34.374	1.00	53.19	AAAA	ATOM	3117	C	ASN	333	35.461	49.551	28.383	1.00	43.54	AAAA
ATOM	2968	CB	SER	316	37.679	42.350	32.990	1.00	51.31	AAAA	ATOM	3118	O	ILE	334	35.431	48.543	27.023	1.00	41.48	AAAA
ATOM	2969	OG	SER	316	37.536	43.754	32.962	1.00	47.38	AAAA	ATOM	3119	CD1	ILE	334	34.277	49.571	28.165	1.00	45.40	AAAA
ATOM	2971	C	SER	316	38.009	42.596	35.477	1.00	51.56	AAAA	ATOM	3120	CD1	ILE	334	34.639	49.626	29.854	1.00	42.84	AAAA
ATOM	2972	O	SER	316	39.212	42.538	35.696	1.00	52.97	AAAA	ATOM	3121	CA	ILE	334	34.548	49.280	30.938	1.00	41.69	AAAA
ATOM	2973	N	ALA	317	37.186	41.342	36.195	1.00	48.93	AAAA	ATOM	3122	CB	ILE	334	34.846	50.400	31.940	1.00	40.51	AAAA
ATOM	2975	CA	ALA	317	37.581	41.668	37.270	1.00	46.85	AAAA	ATOM	3123	CG	ILE	334	34.229	50.169	27.526	1.00	46.97	AAAA
ATOM	2976	CB	ALA	317	36.993	45.518	37.														

Figure 1 (cont'd)

ATOM	1225	O	LEU	344	34.029	48.845	40.528	1.00	40.28	AAAA	ATOM	1381	NE	ARG	361	41.669	51.942	23.519	1.00	42.75	AAAA
ATOM	1226	N	GLU	345	32.200	50.121	40.719	1.00	38.99	AAAA	ATOM	1382	CZ	ARG	361	42.595	51.905	23.684	1.00	42.28	AAAA
ATOM	1228	CA	GLU	345	32.626	50.715	41.973	1.00	41.32	AAAA	ATOM	1384	NH1	ARG	361	43.881	51.302	23.544	1.00	46.14	AAAA
ATOM	1229	CB	GLU	345	31.708	51.871	42.326	1.00	41.39	AAAA	ATOM	1387	NH2	ARG	361	42.236	49.769	23.982	1.00	41.80	AAAA
ATOM	1230	CC	GLU	345	31.979	52.524	43.650	1.00	48.84	AAAA	ATOM	1390	C	ARG	361	38.466	53.782	25.126	1.00	30.62	AAAA
ATOM	1231	CD	GLU	345	30.679	52.858	44.378	1.00	58.27	AAAA	ATOM	1391	O	ARG	361	38.621	52.579	25.252	1.00	34.58	AAAA
ATOM	1232	OE1	GLU	344	29.782	51.979	44.437	1.00	63.94	AAAA	ATOM	1392	HIS	362	37.465	54.333	24.479	1.00	32.74	AAAA	
ATOM	1233	OE2	GLU	345	30.537	53.996	44.887	1.00	62.15	AAAA	ATOM	1394	CA	HIS	362	36.402	53.518	23.879	1.00	34.44	AAAA
ATOM	1234	C	GLU	345	32.708	49.749	43.130	1.00	43.01	AAAA	ATOM	1395	CB	HIS	362	36.995	52.285	23.172	1.00	33.74	AAAA
ATOM	1235	O	GLU	345	33.257	50.085	44.166	1.00	45.52	AAAA	ATOM	1396	CG	HIS	362	37.843	52.621	23.986	1.00	33.42	AAAA
ATOM	1236	N	ASN	346	32.160	48.552	42.968	1.00	45.75	AAAA	ATOM	1397	CD2	HIS	362	38.038	53.793	23.338	1.00	35.37	AAAA
ATOM	1238	CA	ASN	346	32.208	47.529	44.022	1.00	48.31	AAAA	ATOM	1398	ND1	HIS	362	38.655	51.704	21.361	1.00	33.84	AAAA
ATOM	1239	CB	ASN	346	30.852	46.771	44.058	1.00	51.54	AAAA	ATOM	1400	CE1	HIS	362	39.313	52.296	20.383	1.00	32.38	AAAA
ATOM	1240	CG	ASN	346	30.989	52.244	44.244	1.00	55.38	AAAA	ATOM	1401	NE2	HIS	362	38.957	53.565	20.346	1.00	29.52	AAAA
ATOM	1241	OD1	ASN	346	31.208	44.748	45.165	1.00	54.89	AAAA	ATOM	1403	C	HIS	362	35.365	53.064	24.917	1.00	36.04	AAAA
ATOM	1242	ND2	ASN	346	30.832	44.495	43.141	1.00	55.35	AAAA	ATOM	1404	O	HIS	362	34.657	52.059	24.724	1.00	33.48	AAAA
ATOM	1244	O	ASN	346	33.408	46.689	43.713	1.00	49.78	AAAA	ATOM	1405	N	SER	363	35.285	53.817	26.013	1.00	37.14	AAAA
ATOM	1246	O	ASN	346	33.530	45.494	44.226	1.00	52.95	AAAA	ATOM	1407	CA	SER	363	34.360	53.527	27.103	1.00	38.68	AAAA
ATOM	1247	N	PHE	347	34.312	47.105	42.884	1.00	49.92	AAAA	ATOM	1408	CB	SER	363	34.997	53.891	28.442	1.00	37.92	AAAA
ATOM	1249	CA	PHE	347	35.505	46.366	42.500	1.00	47.86	AAAA	ATOM	1409	OG	SER	363	35.693	52.778	28.965	1.00	35.98	AAAA
ATOM	1250	CB	PHE	347	35.314	45.747	41.131	1.00	46.22	AAAA	ATOM	1411	C	SER	363	33.061	54.291	26.927	1.00	40.17	AAAA
ATOM	1251	CG	PHE	347	34.523	44.492	41.122	1.00	45.82	AAAA	ATOM	1412	O	SER	363	32.774	55.265	27.644	1.00	38.73	AAAA
ATOM	1252	CD1	PHE	347	33.374	44.401	40.350	1.00	47.18	AAAA	ATOM	1413	N	HIS	364	32.281	53.843	25.951	1.00	43.35	AAAA
ATOM	1253	CD2	PHE	347	33.975	43.367	41.783	1.00	49.19	AAAA	ATOM	1415	CB	HIS	364	34.955	54.464	25.645	1.00	45.91	AAAA
ATOM	1254	CE1	PHE	347	32.687	43.707	40.225	1.00	48.19	AAAA	ATOM	1416	CB	HIS	364	30.506	53.943	24.315	1.00	42.21	AAAA
ATOM	1255	CE2	PHE	347	34.302	42.161	41.669	1.00	48.72	AAAA	ATOM	1417	CG	HIS	364	31.547	54.027	23.252	1.00	40.39	AAAA
ATOM	1256	CZ	PHE	347	33.153	42.080	40.885	1.00	50.09	AAAA	ATOM	1418	CD2	HIS	364	32.104	53.087	22.454	1.00	37.68	AAAA
ATOM	1257	C	PHE	347	36.690	47.336	42.397	1.00	49.37	AAAA	ATOM	1419	ND1	HIS	364	32.202	55.208	22.973	1.00	36.18	AAAA
ATOM	1258	O	PHE	347	37.847	46.913	42.438	1.00	52.13	AAAA	ATOM	1421	CE1	HIS	364	33.117	54.992	22.050	1.00	37.88	AAAA
ATOM	1259	N	MET	348	36.405	48.633	42.258	1.00	46.83	AAAA	ATOM	1422	NE2	HIS	364	33.080	53.713	21.716	1.00	39.37	AAAA
ATOM	1261	CA	MET	348	37.468	49.615	42.092	1.00	43.67	AAAA	ATOM	1424	C	HIS	364	29.974	54.223	26.745	1.00	46.53	AAAA
ATOM	1262	CB	MET	348	37.525	50.076	40.637	1.00	46.70	AAAA	ATOM	1425	CB	HIS	364	29.200	55.113	27.096	1.00	50.08	AAAA
ATOM	1263	CG	MET	348	37.435	48.955	39.618	1.00	41.47	AAAA	ATOM	1426	N	ALA	365	30.005	54.182	31.586	1.00	44.46	AAAA
ATOM	1264	SD	MET	348	37.816	49.499	37.940	1.00	45.56	AAAA	ATOM	1428	CA	ALA	365	29.102	52.673	28.383	1.00	42.94	AAAA
ATOM	1265	CE	MET	348	37.633	51.237	38.105	1.00	45.81	AAAA	ATOM	1429	CB	ALA	365	29.247	51.212	28.675	1.00	42.23	AAAA
ATOM	1266	C	MET	348	37.346	50.835	42.954	1.00	43.10	AAAA	ATOM	1430	C	ALA	365	29.283	53.472	29.687	1.00	42.62	AAAA
ATOM	1267	O	MET	348	38.237	51.679	42.971	1.00	45.33	AAAA	ATOM	1431	O	ALA	365	28.682	53.100	30.700	1.00	47.27	AAAA
ATOM	1268	N	GLY	349	36.242	50.944	43.667	1.00	45.75	AAAA	ATOM	1432	N	LEU	366	30.079	54.547	29.694	1.00	37.83	AAAA
ATOM	1270	CA	GLY	349	36.024	52.117	44.504	1.00	47.20	AAAA	ATOM	1434	CA	LEU	366	30.301	55.297	30.935	1.00	34.43	AAAA
ATOM	1271	C	GLY	349	37.063	52.367	45.571	1.00	46.70	AAAA	ATOM	1435	CB	LEU	366	31.782	55.347	31.330	1.00	35.50	AAAA
ATOM	1272	O	GLY	349	37.211	53.501	46.038	1.00	45.69	AAAA	ATOM	1436	CG	LEU	366	32.748	54.862	32.474	1.00	37.95	AAAA
ATOM	1273	N	LEU	350	37.786	51.312	45.944	1.00	47.30	AAAA	ATOM	1437	CD1	LEU	366	34.151	54.791	31.721	1.00	39.21	AAAA
ATOM	1275	CA	LEU	350	38.797	51.417	46.982	1.00	48.38	AAAA	ATOM	1438	CD2	LEU	366	32.391	53.388	32.835	1.00	33.39	AAAA
ATOM	1276	CB	LEU	350	38.826	50.150	47.817	1.00	47.67	AAAA	ATOM	1439	C	LEU	366	29.872	56.711	30.782	1.00	34.59	AAAA
ATOM	1277	CG	LEU	350	38.226	50.472	49.189	1.00	50.77	AAAA	ATOM	1440	O	LEU	366	29.887	57.226	29.681	1.00	39.48	AAAA
ATOM	1278	CD1	LEU	350	37.770	49.183	49.823	1.00	49.78	AAAA	ATOM	1441	N	VAL	367	29.552	57.357	31.901	1.00	34.62	AAAA
ATOM	1279	CD2	LEU	350	39.259	51.213	50.078	1.00	51.01	AAAA	ATOM	1443	CA	VAL	367	29.131	58.754	31.905	1.00	34.89	AAAA
ATOM	1280	C	LEU	350	40.214	51.767	46.568	1.00	48.86	AAAA	ATOM	1444	CB	VAL	367	27.646	58.863	32.297	1.00	36.40	AAAA
ATOM	1281	O	LEU	350	52.111	52.141	47.433	1.00	51.97	AAAA	ATOM	1445	CG1	VAL	367	60.210	60.320	32.474	1.00	38.87	AAAA
ATOM	1282	N	ILE	351	40.531	51.655	45.274	1.00	46.49	AAAA	ATOM	1446	CG2	VAL	367	26.819	59.229	31.224	1.00	36.70	AAAA
ATOM	1284	CA	ILE	351	41.874	51.996	44.800	1.00	41.30	AAAA	ATOM	1447	C	VAL	367	29.981	59.577	32.872	1.00	34.18	AAAA
ATOM	1285	CB	ILE	351	42.026	51.782	43.279	1.00	36.48	AAAA	ATOM	1448	O	VAL	367	29.994	60.813	32.833	1.00	30.93	AAAA
ATOM	1286	CG2	ILE	351	43.289	52.458	42.769	1.00	34.25	AAAA	ATOM	1449	N	SER	368	30.687	58.873	33.747	1.00	37.44	AAAA
ATOM	1287	CG1	ILE	351	42.118	50.299	42.975	1.00	31.15	AAAA	ATOM	1451	CA	SER	368	31.563	59.502	34.733	1.00	40.82	AAAA
ATOM	1288	CD1	ILE	351	41.280	49.917	41.829	1.00	30.14	AAAA	ATOM	1452	CB	SER	368	30.780	60.006	35.934	1.00	44.22	AAAA
ATOM	1289	C	ILE	351	42.134	53.461	45.135	1.00	41.17	AAAA	ATOM	1453	OG	SER	368	31.610	60.030	37.088	1.00	51.43	AAAA
ATOM	1290	O	ILE	351	54.134	54.337	44.780	1.00	44.80	AAAA	ATOM	1454	C	SER	368	32.510	58.464	35.230	1.00	40.25	AAAA
ATOM	1291	N	GLU	352	43.236	53.721	45.834	1.00	37.12	AAAA	ATOM	1456	O	SER	368	32.236	57.276	35.171	1.00	40.17	AAAA
ATOM	1293	CA	GLU	352	43.551	55.067	46.221	1.00	36.72	AAAA	ATOM	1457	N	LEU	369	33.674	58.911	35.726	1.00	40.76	AAAA
ATOM	1294	CB	GLU	352	43.958	55.093	47.663	1.00	40.63	AAAA	ATOM	1459	CA	LEU	369	34.672	57.988	36.248	1.00	43.36	AAAA
ATOM	1295	CG	GLU	352	42.882	55.428	48.601	1.00	44.83	AAAA	ATOM	1460	CB	LEU	369	36.051	58.431	35.790	1.00	42.84	AAAA
ATOM	1296	CD	GLU	352	43.204	54.868	49.935	1.00	48.53	AAAA	ATOM	1461	CG	LEU	369	36.153	58.281	34.281	1.00	40.88	AAAA
ATOM	1297	OE1	GLU	352	44.020	55.518	50.640	1.00	49.86	AAAA	ATOM	1462	CD1	LEU	369	37.404	58.982	33.798	1.00	40.40	AAAA
ATOM	1298	OE2	GLU	352	42.659	53.780	50.252	1.00	49.21	AAAA	ATOM	1463	CD2	LEU	369	36.155	56.801	33.922	1.00	38.47	AAAA
ATOM	1300	O	GLU	352	44.712	56.601	51.640	1.0													

Figure 1 (cont'd)

ATOM 3551 CD2 LEU 177	48.524	59.490	46.127	1.00	35.51	AAAA	ATOM 3706 CD2 LEU 393	41.712	57.482	21.327	1.00	25.78	AAAA
ATOM 3552 C LEU 177	46.966	59.205	42.276	1.00	35.54	AAAA	ATOM 3707 C LEU 393	38.907	58.277	23.572	1.00	31.54	AAAA
ATOM 3553 O LEU 177	46.792	57.988	42.342	1.00	32.60	AAAA	ATOM 3708 O LEU 393	38.885	57.086	23.916	1.00	30.34	AAAA
ATOM 3554 N ILE 178	47.601	59.800	41.267	1.00	37.18	AAAA	ATOM 3709 N ASP 394	38.021	58.812	22.726	1.00	32.68	AAAA
ATOM 3556 CA ILE 178	48.258	59.087	40.169	1.00	37.18	AAAA	ATOM 3711 CA ASP 394	36.991	57.993	22.095	1.00	34.14	AAAA
ATOM 3557 CB ILE 178	47.621	59.424	38.797	1.00	36.92	AAAA	ATOM 3712 CB ASP 394	37.681	56.856	21.355	1.00	39.38	AAAA
ATOM 3558 CG2 ILE 178	48.421	58.788	37.675	1.00	31.15	AAAA	ATOM 3713 CG2 ASP 394	36.891	55.716	20.184	1.00	40.62	AAAA
ATOM 3559 CG1 ILE 178	46.186	58.078	38.737	1.00	36.11	AAAA	ATOM 3714 OD1 ASP 394	37.510	56.556	21.339	1.00	43.56	AAAA
ATOM 3560 CD1 ILE 178	45.373	59.423	37.585	1.00	33.62	AAAA	ATOM 3715 OD2 ASP 394	35.665	56.592	20.190	1.00	39.44	AAAA
ATOM 3561 C ILE 178	49.727	59.543	40.194	1.00	39.52	AAAA	ATOM 3716 C ASP 394	36.051	57.428	23.131	1.00	28.78	AAAA
ATOM 3562 O ILE 178	50.032	60.736	40.099	1.00	40.21	AAAA	ATOM 3717 O ASP 394	35.968	56.205	23.339	1.00	34.65	AAAA
ATOM 3563 N LEU 179	50.628	58.577	40.445	1.00	40.05	AAAA	ATOM 3718 N ASN 395	34.407	57.984	24.819	1.00	36.47	AAAA
ATOM 3565 CA LEU 179	52.055	58.842	40.447	1.00	38.95	AAAA	ATOM 3720 CA ASN 395	34.991	58.434	26.157	1.00	35.66	AAAA
ATOM 3566 CB LEU 179	52.739	57.650	41.129	1.00	38.74	AAAA	ATOM 3722 CB ASN 395	36.086	57.502	26.646	1.00	35.14	AAAA
ATOM 3567 CG LEU 179	52.095	57.212	42.463	1.00	37.60	AAAA	ATOM 3723 OD1 ASN 395	35.902	56.277	26.740	1.00	37.26	AAAA
ATOM 3568 CD1 LEU 179	52.835	56.032	43.029	1.00	37.60	AAAA	ATOM 3724 ND2 ASN 395	37.219	58.080	26.956	1.00	37.41	AAAA
ATOM 3569 OD1 LEU 179	52.359	56.359	43.082	1.00	36.85	AAAA	ATOM 3727 C ASN 395	33.032	58.589	24.567	1.00	36.53	AAAA
ATOM 3570 C LEU 179	52.733	59.165	39.128	1.00	40.01	AAAA	ATOM 3728 O ASN 395	32.695	59.670	25.091	1.00	37.94	AAAA
ATOM 3571 O LEU 179	53.593	60.044	39.076	1.00	43.96	AAAA	ATOM 3729 N GLN 396	32.245	57.880	23.770	1.00	38.09	AAAA
ATOM 3572 N GLY 380	52.364	58.463	38.064	1.00	36.71	AAAA	ATOM 3731 CA GLN 396	30.911	58.302	23.339	1.00	37.03	AAAA
ATOM 3574 CA GLY 380	52.956	58.755	36.778	1.00	34.15	AAAA	ATOM 3732 CB GLN 396	30.173	57.106	22.740	1.00	37.03	AAAA
ATOM 3575 C GLY 380	54.229	57.991	36.485	1.00	35.78	AAAA	ATOM 3733 CG GLN 396	29.991	57.148	21.246	1.00	35.68	AAAA
ATOM 3576 G GLY 380	54.771	58.120	35.401	1.00	36.80	AAAA	ATOM 3734 CD GLN 396	29.509	55.815	20.696	1.00	41.00	AAAA
ATOM 3577 N GLU 381	54.705	57.186	37.433	1.00	36.39	AAAA	ATOM 3736 OE1 GLN 396	28.688	55.145	21.310	1.00	47.25	AAAA
ATOM 3579 CA GLU 381	55.936	56.393	37.259	1.00	36.23	AAAA	ATOM 3736 NE2 GLN 396	29.012	55.429	21.535	1.00	42.50	AAAA
ATOM 3580 CB GLU 381	56.021	55.336	38.367	1.00	32.64	AAAA	ATOM 3739 C GLN 396	29.959	59.039	24.299	1.00	38.09	AAAA
ATOM 3581 CG GLU 381	55.424	55.837	39.673	1.00	31.08	AAAA	ATOM 3740 O GLN 396	29.418	60.055	23.913	1.00	39.65	AAAA
ATOM 3582 CD GLU 381	56.134	55.337	40.915	1.00	30.26	AAAA	ATOM 3741 N ASN 397	29.767	58.526	25.516	1.00	40.01	AAAA
ATOM 3583 OE1 GLU 381	56.193	54.097	41.090	1.00	25.59	AAAA	ATOM 3743 CA ASN 397	28.794	59.115	26.467	1.00	38.64	AAAA
ATOM 3584 OE2 GLU 381	56.613	56.192	41.718	1.00	28.20	AAAA	ATOM 3744 CB ASN 397	27.795	58.035	26.846	1.00	35.69	AAAA
ATOM 3585 C GLU 381	56.095	55.739	35.867	1.00	38.63	AAAA	ATOM 3745 CG ASN 397	27.182	57.370	25.624	1.00	39.95	AAAA
ATOM 3586 O GLU 381	57.201	55.342	35.467	1.00	39.43	AAAA	ATOM 3746 OD1 ASN 397	26.447	58.014	24.865	1.00	42.85	AAAA
ATOM 3587 N GLU 381	55.492	55.637	35.133	1.00	36.87	AAAA	ATOM 3747 C ASN 397	27.484	56.084	25.417	1.00	36.45	AAAA
ATOM 3589 CA GLU 382	54.999	55.074	33.800	1.00	36.67	AAAA	ATOM 3750 C ASN 397	28.254	59.795	27.750	1.00	38.08	AAAA
ATOM 3590 CB GLU 382	54.645	53.613	33.835	1.00	33.71	AAAA	ATOM 3751 O ASN 397	28.440	60.235	28.555	1.00	35.86	AAAA
ATOM 3591 CG GLU 382	54.998	52.888	32.572	1.00	44.29	AAAA	ATOM 3752 N LEU 398	30.580	59.873	27.923	1.00	38.81	AAAA
ATOM 3592 CD GLU 382	55.450	51.444	32.833	1.00	52.79	AAAA	ATOM 3754 CA LEU 398	31.208	60.452	29.099	1.00	38.24	AAAA
ATOM 3593 OE1 GLU 382	55.776	51.157	33.946	1.00	51.32	AAAA	ATOM 3755 CB LEU 398	32.725	60.276	28.995	1.00	38.47	AAAA
ATOM 3594 OE2 GLU 382	55.278	50.597	31.914	1.00	57.67	AAAA	ATOM 3756 CG LEU 398	33.536	59.791	30.199	1.00	33.50	AAAA
ATOM 3595 C GLU 382	53.919	55.851	33.098	1.00	37.67	AAAA	ATOM 3757 CD1 LEU 398	34.095	58.432	29.882	1.00	35.71	AAAA
ATOM 3596 O GLU 382	52.850	56.075	33.667	1.00	40.34	AAAA	ATOM 3758 CD2 LEU 398	34.660	60.760	30.498	1.00	32.89	AAAA
ATOM 3597 N GLU 383	54.205	54.286	31.849	1.00	39.42	AAAA	ATOM 3759 C LEU 398	30.879	61.912	29.361	1.00	40.51	AAAA
ATOM 3599 CA GLN 383	53.274	57.090	31.073	1.00	34.77	AAAA	ATOM 3760 CB LEU 398	31.307	62.806	28.620	1.00	41.09	AAAA
ATOM 3600 CB GLN 383	53.599	58.572	31.212	1.00	29.45	AAAA	ATOM 3761 N GLN 399	30.159	62.150	30.458	1.00	44.82	AAAA
ATOM 3601 CG GLN 383	54.361	58.896	32.453	1.00	24.55	AAAA	ATOM 3763 CA GLN 399	29.743	63.498	30.845	1.00	46.98	AAAA
ATOM 3602 CD GLN 383	54.918	60.263	32.392	1.00	24.86	AAAA	ATOM 3764 CB GLN 399	28.298	63.458	31.344	1.00	51.82	AAAA
ATOM 3603 OE1 GLN 383	55.179	60.753	31.316	1.00	29.07	AAAA	ATOM 3765 CG GLN 399	27.287	63.807	30.259	1.00	58.43	AAAA
ATOM 3604 NE2 GLN 383	55.097	60.906	33.541	1.00	28.28	AAAA	ATOM 3766 CD GLN 399	26.083	62.897	30.283	1.00	63.27	AAAA
ATOM 3607 C GLN 383	53.335	56.734	29.604	1.00	34.72	AAAA	ATOM 3767 OE1 GLN 399	24.992	63.325	30.658	1.00	65.70	AAAA
ATOM 3608 O GLN 383	53.337	56.216	29.126	1.00	37.41	AAAA	ATOM 3768 NE2 GLN 399	26.271	61.627	29.888	1.00	63.89	AAAA
ATOM 3609 N LEU 384	52.355	57.029	26.891	1.00	43.39	AAAA	ATOM 3771 C GLN 399	30.621	64.227	31.870	1.00	44.75	AAAA
ATOM 3611 CA LEU 384	52.189	56.746	27.475	1.00	31.41	AAAA	ATOM 3772 O GLN 399	30.915	64.095	31.727	1.00	43.25	AAAA
ATOM 3612 CB LEU 384	50.753	56.864	26.967	1.00	31.41	AAAA	ATOM 3773 N GLN 400	31.012	63.525	32.422	1.00	42.85	AAAA
ATOM 3613 CG LEU 384	49.706	56.196	27.848	1.00	29.58	AAAA	ATOM 3775 CA GLN 400	31.866	64.126	33.921	1.00	43.95	AAAA
ATOM 3614 CD1 LEU 384	48.674	57.221	28.230	1.00	29.04	AAAA	ATOM 3776 CB GLN 400	31.075	64.482	35.191	1.00	43.36	AAAA
ATOM 3615 CD2 LEU 384	49.069	55.045	27.117	1.00	29.25	AAAA	ATOM 3777 CG GLN 400	30.463	63.264	35.901	1.00	45.91	AAAA
ATOM 3616 C LEU 384	53.089	57.715	26.737	1.00	33.00	AAAA	ATOM 3778 CD GLN 400	30.460	63.351	37.440	1.00	46.70	AAAA
ATOM 3617 O LEU 384	53.484	58.755	27.254	1.00	28.69	AAAA	ATOM 3779 OE1 GLN 400	30.489	64.443	38.023	1.00	46.78	AAAA
ATOM 3618 N GLU 385	53.411	57.356	25.510	1.00	37.90	AAAA	ATOM 3780 NE2 GLN 400	30.410	62.288	38.095	1.00	43.73	AAAA
ATOM 3620 CA GLU 385	54.424	59.665	27.708	1.00	42.84	AAAA	ATOM 3781 C GLN 400	32.992	63.178	34.292	1.00	44.99	AAAA
ATOM 3621 CB GLU 385	54.475	57.507	33.337	1.00	49.19	AAAA	ATOM 3782 O GLN 400	32.903	63.592	34.313	1.00	45.39	AAAA
ATOM 3622 CG GLU 385	55.273	56.189	33.393	1.00	53.13	AAAA	ATOM 3785 N LEU 401	34.059	63.751	34.823	1.00	43.93	AAAA
ATOM 3623 CD GLU 385	54.464	54.951	33.875	1.00	58.73	AAAA	ATOM 3787 CA LEU 401	35.177	62.941	35.243	1.00	41.07	AAAA
ATOM 3624 OE1 GLU 385	53.470	55.066	24.640	1.00	57.99	AAAA	ATOM 3788 CB LEU 401	36.445	63.813	35.231	1.00	38.05	AAAA
ATOM 3625 OE2 GLU 385	54.845	53.826	23.478	1.00	62.33	AAAA	ATOM 3789 CG LEU 401	36.800	64.425	33.857	1.00	31.72	AAAA
ATOM 3626 C GLU 385	53.680	59.551	24.625	1.00	42.49	AAAA							
ATOM 3627 O GLU 385	52.469	59.694	24.402	1.00	44.93	AAAA	ATOM 3790 CD1 LEU 401	37.977	65.370	33.971	1.00	28.74	AAAA
ATOM 3628 N GLY 386	54.521	60.561	24.827	1.00	39.27	AAAA	ATOM 3791 CD2 LEU 401	37.123	63.330	32.887	1.00	27.88	AAAA
ATOM 3630 CA GLY 386	54.062	61.936	24.819	1.00	36.93	AAAA	ATOM 3792 C LEU 401	34.806	62.415	36.639	1.00	40.74	AAAA
ATOM 3631 C GLY 386	54.135	62.325	26.270	1.00	36.18	AAAA	ATOM 3793 O LEU 401	34.224	61.335	36.773	1.00	39.04	AAAA
ATOM 3632 O GLY 386	63.402	62.490	26.685	1.00	34.95	AAAA	ATOM 3794 N TRP 402	35.128	61.203	37.659	1.00	43.32	AAAA
ATOM 3633 N ASN 387	54.353	61.289	27.056	1.00	37.01	AAAA	ATOM 3795 CA TRP 402	34.828	62.899	39.061	1.00	48.15	AAAA
ATOM 3635 CA ASN 387	54.458	61.424	28.497	1.00	38.05	AAAA	ATOM 3797 CB TRP 402	36.108	62.528	39.888	1.00	46.87	AAAA
ATOM 3636 CB ASN 387	55.608	62.377	28.828	1.00	39.59	AAAA	ATOM 3798 CG TRP 402	36.784	61.226	39.735	1.00	43.99	AAAA
ATOM 3637 CG ASN 387	56.980	61.681	28.685	1.00	45.31	AAAA	ATOM 3799 CD2 TRP 402	37.854	60.898	38.833	1.00	44.35	AAAA
ATOM 3638 OD1 ASN 387	57.057	60.430	28.596	1.00	44.06	AAAA	ATOM 3800 CE2 TRP 402	38.201	59.545	39.072	1.00	45.95	AAAA
ATOM 3639 ND2 ASN 387	58.057	62.475	28.650	1.00	44.74	AAAA	ATOM 3801 CE3 TRP 402	38.553	61.609	37.846	1.00	39.33	AAAA
ATOM 3642 C ASN 387	53.123	61.880	29.055	1.00	37.24	AAAA	ATOM 3802 CD1 TRP 402	36.516	60.091	40.462	1.00	44.07	AAAA
ATOM 3643 O ASN 387	52.972	63.0											

Figure 1 (cont'd)

ATOM	1872	C	LEU	409	44.211	64.611	42.172	1.00	49.41	AAAA	ATOM	4031	O	VAL	426	37.080	72.084	30.473	1.00	45.01	AAAA
ATOM	1873	O	LEU	409	44.192	65.839	42.316	1.00	48.73	AAAA	ATOM	4032	N	SER	427	34.946	72.579	30.000	1.00	46.62	AAAA
ATOM	1874	N	THR	410	45.117	61.891	42.198	1.00	48.82	AAAA	ATOM	4034	CA	SER	427	34.695	73.194	31.293	1.00	48.46	AAAA
ATOM	1876	CA	THR	410	46.605	64.527	41.952	1.00	46.46	AAAA	ATOM	4035	CB	SER	427	37.184	73.447	31.456	1.00	49.20	AAAA
ATOM	1877	CB	THR	410	47.364	64.730	43.270	1.00	47.45	AAAA	ATOM	4036	OG	SER	427	32.597	73.899	30.229	1.00	51.43	AAAA
ATOM	1878	CG1	THR	410	46.860	65.897	43.940	1.00	48.99	AAAA	ATOM	4038	C	SER	427	35.237	72.316	32.421	1.00	48.91	AAAA
ATOM	1880	CG2	THR	410	46.884	64.881	42.997	1.00	47.91	AAAA	ATOM	4039	O	SER	427	35.928	72.794	33.118	1.00	49.65	AAAA
ATOM	1881	C	THR	410	47.506	63.740	41.009	1.00	43.42	AAAA	ATOM	4040	N	GLU	428	34.946	73.025	31.370	1.00	50.84	AAAA
ATOM	1882	O	THR	410	47.716	62.551	41.191	1.00	42.61	AAAA	ATOM	4042	CA	GLU	428	35.425	70.123	31.402	1.00	51.24	AAAA
ATOM	1883	N	ILE	411	48.035	64.406	39.995	1.00	41.25	AAAA	ATOM	4043	CB	GLU	428	34.693	68.787	33.309	1.00	53.24	AAAA
ATOM	1885	CA	ILE	411	48.934	63.735	39.070	1.00	41.84	AAAA	ATOM	4044	CG	GLU	428	33.197	68.875	33.541	1.00	60.59	AAAA
ATOM	1886	CB	ILE	411	48.565	64.010	37.569	1.00	36.01	AAAA	ATOM	4045	CD	GLU	428	32.829	69.284	34.969	1.00	65.08	AAAA
ATOM	1887	CG2	ILE	411	49.253	63.011	36.662	1.00	31.64	AAAA	ATOM	4046	OE1	GLU	428	32.667	68.380	35.839	1.00	66.54	AAAA
ATOM	1888	CG1	ILE	411	47.050	63.889	37.375	1.00	36.00	AAAA	ATOM	4047	OE2	GLU	428	32.696	70.514	35.209	1.00	68.29	AAAA
ATOM	1889	CD1	ILE	411	46.594	62.884	36.336	1.00	31.18	AAAA	ATOM	4048	C	GLU	428	36.921	69.886	33.234	1.00	51.94	AAAA
ATOM	1890	C	ILE	411	50.338	64.265	39.407	1.00	45.51	AAAA	ATOM	4049	O	GLU	428	37.706	69.999	34.168	1.00	56.08	AAAA
ATOM	1891	O	ILE	411	50.730	65.378	38.003	1.00	46.25	AAAA	ATOM	4050	N	ILE	429	37.315	69.552	32.022	1.00	50.82	AAAA
ATOM	1892	N	LVS	412	51.061	63.468	40.195	1.00	43.71	AAAA	ATOM	4052	CA	ILE	429	38.697	69.280	31.750	1.00	48.09	AAAA
ATOM	1894	CA	LVS	412	52.419	63.769	40.634	1.00	41.26	AAAA	ATOM	4053	CB	ILE	429	38.897	69.136	30.240	1.00	45.91	AAAA
ATOM	1895	CB	LVS	412	52.972	62.563	41.369	1.00	42.01	AAAA	ATOM	4054	CG2	ILE	429	40.359	69.094	29.893	1.00	43.32	AAAA
ATOM	1896	CG	LVS	412	53.111	62.833	42.746	1.00	49.93	AAAA	ATOM	4055	CG1	ILE	429	38.214	67.854	29.785	1.00	42.83	AAAA
ATOM	1898	C	LVS	412	53.350	64.073	39.483	1.00	41.15	AAAA	ATOM	4056	CD1	ILE	429	38.390	67.553	28.335	1.00	44.47	AAAA
ATOM	1899	O	LVS	412	54.195	64.959	39.560	1.00	40.96	AAAA	ATOM	4057	C	ILE	429	39.645	70.319	32.332	1.00	50.57	AAAA
ATOM	1900	N	ALA	413	53.192	63.109	38.411	1.00	42.72	AAAA	ATOM	4058	O	ILE	429	40.722	69.985	32.817	1.00	50.48	AAAA
ATOM	1901	CA	ALA	413	54.043	63.445	37.255	1.00	40.95	AAAA	ATOM	4059	N	ALA	430	39.249	71.596	32.120	1.00	57.04	AAAA
ATOM	1903	CB	ALA	413	55.277	62.614	37.471	1.00	44.44	AAAA	ATOM	4061	CA	TYR	430	40.189	72.608	32.854	1.00	55.94	AAAA
ATOM	1904	C	ALA	413	53.366	63.028	35.959	1.00	40.18	AAAA	ATOM	4062	CB	TYR	430	40.000	74.001	32.217	1.00	59.37	AAAA
ATOM	1905	O	ALA	413	52.599	62.071	35.929	1.00	40.51	AAAA	ATOM	4063	CG	TYR	430	39.958	74.055	30.692	1.00	61.64	AAAA
ATOM	1906	N	GLY	414	53.694	63.729	34.884	1.00	39.10	AAAA	ATOM	4064	CD1	TYR	430	39.656	72.923	29.943	1.00	61.92	AAAA
ATOM	1908	CA	GLY	414	53.086	63.434	33.603	1.00	40.43	AAAA	ATOM	4065	CD2	TYR	430	39.447	72.997	28.883	1.00	61.98	AAAA
ATOM	1909	C	GLY	414	51.983	64.453	33.371	1.00	41.42	AAAA	ATOM	4066	CE1	TYR	430	40.073	75.277	30.013	1.00	62.01	AAAA
ATOM	1910	O	GLY	414	51.870	64.415	34.131	1.00	42.72	AAAA	ATOM	4067	CE2	TYR	430	39.853	75.356	28.641	1.00	61.34	AAAA
ATOM	1911	N	LVS	415	55.161	64.247	32.047	1.00	40.85	AAAA	ATOM	4068	CG	TYR	430	39.539	74.206	27.935	1.00	61.30	AAAA
ATOM	1913	CA	LVS	415	50.065	65.173	32.040	1.00	38.88	AAAA	ATOM	4069	OH	TYR	430	39.226	74.254	26.595	1.00	62.51	AAAA
ATOM	1914	CB	LVS	415	50.556	66.241	31.084	1.00	39.32	AAAA	ATOM	4071	C	TYR	430	40.068	72.726	34.366	1.00	55.10	AAAA
ATOM	1915	CG	LVS	415	50.938	65.625	29.744	1.00	40.34	AAAA	ATOM	4072	O	TYR	430	40.961	73.266	35.026	1.00	56.49	AAAA
ATOM	1916	CD	LVS	415	51.401	66.633	28.746	1.00	38.47	AAAA	ATOM	4073	N	ARG	431	38.964	72.229	34.908	1.00	52.81	AAAA
ATOM	1917	CE	LVS	415	52.618	66.153	28.048	1.00	35.12	AAAA	ATOM	4075	CA	ARG	431	38.776	72.253	36.346	1.00	52.51	AAAA
ATOM	1918	NZ	LVS	415	52.669	66.925	26.813	1.00	43.52	AAAA	ATOM	4076	CB	ARG	431	37.377	71.782	36.742	1.00	54.84	AAAA
ATOM	1922	C	LVS	415	48.885	64.446	31.383	1.00	37.63	AAAA	ATOM	4077	CG	ARG	431	37.349	71.386	38.106	1.00	57.04	AAAA
ATOM	1923	N	LVS	415	48.834	63.201	31.334	1.00	37.93	AAAA	ATOM	4079	C	MET	431	39.770	71.331	36.836	1.00	51.03	AAAA
ATOM	1924	N	MET	416	47.962	65.234	30.834	1.00	35.60	AAAA	ATOM	4080	O	ARG	431	40.703	71.582	37.554	1.00	49.10	AAAA
ATOM	1926	CA	MET	416	46.759	64.690	30.204	1.00	30.58	AAAA	ATOM	4081	N	MET	432	39.536	70.009	36.427	1.00	52.81	AAAA
ATOM	1927	CB	MET	416	45.548	65.090	31.032	1.00	31.10	AAAA	ATOM	4083	CA	MET	432	40.376	68.866	36.766	1.00	55.38	AAAA
ATOM	1928	CG	MET	416	45.916	65.605	32.386	1.00	32.69	AAAA	ATOM	4084	CB	MET	432	40.049	67.712	35.823	1.00	52.88	AAAA
ATOM	1929	SD	MET	416	44.485	65.721	33.413	1.00	39.47	AAAA	ATOM	4085	CG	MET	432	40.286	66.348	35.402	1.00	50.22	AAAA
ATOM	1930	CE	MET	416	44.258	67.553	33.433	1.00	33.89	AAAA	ATOM	4086	SD	MET	432	41.440	65.457	35.165	1.00	47.27	AAAA
ATOM	1931	C	MET	416	46.475	65.038	28.755	1.00	26.23	AAAA	ATOM	4087	CE	MET	432	40.515	63.999	34.999	1.00	52.56	AAAA
ATOM	1932	O	MET	416	46.851	66.096	28.258	1.00	24.44	AAAA	ATOM	4088	C	MET	432	41.852	67.232	36.642	1.00	58.02	AAAA
ATOM	1933	N	TYR	417	45.797	64.114	28.090	1.00	24.40	AAAA	ATOM	4089	O	MET	432	42.170	68.694	37.346	1.00	59.47	AAAA
ATOM	1935	CA	TYR	417	45.375	64.284	26.710	1.00	26.54	AAAA	ATOM	4090	N	GLU	433	42.131	70.161	35.732	1.00	59.97	AAAA
ATOM	1936	CB	TYR	417	46.137	63.349	25.779	1.00	25.59	AAAA	ATOM	4092	CA	GLU	433	43.480	70.643	35.483	1.00	60.19	AAAA
ATOM	1937	CG	TYR	417	45.660	63.489	24.350	1.00	29.15	AAAA	ATOM	4093	CB	GLU	433	43.447	71.636	34.349	1.00	60.90	AAAA
ATOM	1938	CD1	TYR	417	45.859	64.686	23.659	1.00	27.78	AAAA	ATOM	4094	CG	GLU	433	43.854	71.206	33.021	1.00	62.57	AAAA
ATOM	1939	CE1	TYR	417	45.383	64.867	22.368	1.00	28.95	AAAA	ATOM	4095	CD	GLU	433	44.322	72.242	32.159	1.00	64.14	AAAA
ATOM	1940	CD2	TYR	417	44.967	62.453	21.997	1.00	29.10	AAAA	ATOM	4096	OE1	GLU	433	44.681	73.282	32.742	1.00	64.05	AAAA
ATOM	1941	CE2	TYR	417	44.479	62.622	22.382	1.00	26.46	AAAA	ATOM	4097	OE2	GLU	433	44.032	72.117	30.920	1.00	66.58	AAAA
ATOM	1942	CH	TYR	417	44.692	63.844	21.735	1.00	28.87	AAAA	ATOM	4098	C	GLU	433	44.032	71.348	36.710	1.00	60.66	AAAA
ATOM	1943	OH	TYR	417	44.180	64.116	20.474	1.00	38.55	AAAA	ATOM	4099	O	GLU	433	45.232	71.399	36.996	1.00	62.11	AAAA
ATOM	1945	C	TYR	417	43.877	63.956	26.616	1.00	29.17	AAAA	ATOM	4100	N	GLU	434	43.147	72.035	37.416	1.00	60.35	AAAA
ATOM	1946	O	TYR	417	43.443	62.845	26.953	1.00	30.30	AAAA	ATOM	4102	CA	GLU	434	43.541	72.770	38.603	1.00	60.47	AAAA
ATOM	1947	N	PHE	418	43.081	64.917	26.173	1.00	30.27	AAAA	ATOM	4103	CB	GLU	434	42.502	73.838	38.923	1.00	63.91	AAAA
ATOM	1949	CA	PHE	418	41.645	64.685	26.048	1.00	33.08	AAAA	ATOM	4104	CG	GLU	434	41.554	74.150	37.800	1.00	67.95	AAAA
ATOM	1950	CB	PHE	418	40.865	65.640	25.838	1.00	34.47	AAAA	ATOM	4105	CD	GLU	434	42.380	73.580	37.427	1.00	67.47	AAAA
ATOM	1951	CG	PHE	418	40.830	65.224	28.393	1.00													

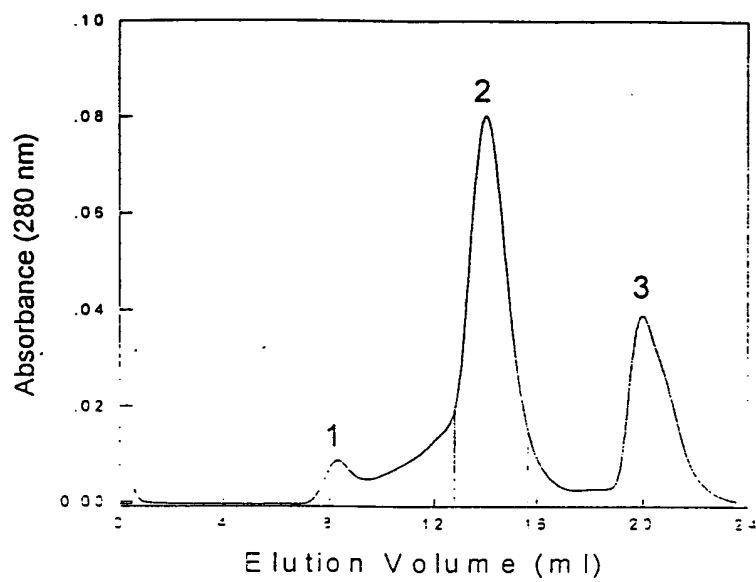
Figure 1 (cont'd)

ATOM	4194	O	LYS	444	45.682	71.789	22.581	1.00	65.94	AAAA	ATOM	4190	C3	NAG	105B	30.646	22.028	75.808	1.00	65.10	BBBB
ATOM	4195	N	GLY	445	47.086	70.421	21.445	1.00	63.29	AAAA	ATOM	4192	O3	NAG	105B	29.991	23.215	75.373	1.00	72.19	BBBB
ATOM	4197	CA	GLY	445	46.066	69.421	21.117	1.00	59.11	AAAA	ATOM	4194	C4	NAG	105B	29.721	21.268	76.792	1.00	65.51	BBBB
ATOM	4198	C	GLY	445	45.622	68.522	22.247	1.00	55.66	AAAA	ATOM	4196	O4	NAG	105B	29.607	21.997	78.009	1.00	65.49	BBBB
ATOM	4199	O	GLY	445	44.792	67.610	22.064	1.00	52.13	AAAA	ATOM	4198	C5	NAG	105B	30.330	19.889	77.043	1.00	64.84	BBBB
ATOM	4200	N	ASP	446	46.199	68.806	23.414	1.00	53.96	AAAA	ATOM	4200	O5	NAG	105B	30.415	19.199	75.797	1.00	62.03	BBBB
ATOM	4202	CA	ASP	446	45.932	68.099	24.668	1.00	49.82	AAAA	ATOM	4201	C6	NAG	105B	29.654	18.950	78.009	1.00	67.03	BBBB
ATOM	4203	CB	ASP	446	46.726	68.767	25.819	1.00	46.80	AAAA	ATOM	4204	O6	NAG	105B	30.323	17.684	78.007	1.00	67.68	BBBB
ATOM	4204	CG	ASP	446	48.109	68.141	26.059	1.00	43.77	AAAA	ATOM	4206	C1	NAG	284A	55.225	45.501	66.582	1.00	40.21	BBBB
ATOM	4205	OD1	ASP	446	48.659	67.456	25.174	1.00	39.45	AAAA	ATOM	4208	C2	NAG	284A	56.197	46.668	66.564	1.00	43.05	BBBB
ATOM	4206	OD2	ASP	446	48.655	68.350	27.162	1.00	45.58	AAAA	ATOM	4210	N2	NAG	284A	56.745	46.859	65.232	1.00	43.34	BBBB
ATOM	4207	C	ASP	446	44.425	68.197	24.955	1.00	46.31	AAAA	ATOM	4212	C7	NAG	284A	56.896	48.078	64.713	1.00	39.06	BBBB
ATOM	4208	O	ASP	446	43.707	67.202	25.011	1.00	43.03	AAAA	ATOM	4213	O7	NAG	284A	56.610	49.107	65.308	1.00	42.15	BBBB
ATOM	4209	N	ILE	447	43.957	69.423	25.115	1.00	43.79	AAAA	ATOM	4214	C8	NAG	284A	57.462	48.164	63.316	1.00	39.13	BBBB
ATOM	4211	CA	ILE	447	42.574	69.646	25.407	1.00	44.45	AAAA	ATOM	4218	C3	NAG	284A	57.273	46.321	67.579	1.00	39.39	BBBB
ATOM	4212	CB	ILE	447	42.435	70.389	26.758	1.00	42.60	AAAA	ATOM	4220	O3	NAG	284A	58.265	47.338	67.593	1.00	41.03	BBBB
ATOM	4213	CG2	ILE	447	40.978	70.386	27.225	1.00	41.47	AAAA	ATOM	4222	C4	NAG	284A	58.630	46.124	68.862	1.00	39.94	BBBB
ATOM	4214	CG1	ILE	447	42.308	69.698	29.014	1.00	41.98	AAAA	ATOM	4224	O4	NAG	284A	57.613	45.720	69.912	1.00	36.85	BBBB
ATOM	4215	CD1	ILE	447	43.069	70.172	29.231	1.00	37.75	AAAA	ATOM	4225	C5	NAG	284A	55.474	45.177	68.945	1.00	41.49	BBBB
ATOM	4216	C	ILE	447	41.916	70.438	24.292	1.00	46.85	AAAA	ATOM	4227	O5	NAG	284A	54.559	45.483	67.862	1.00	45.44	BBBB
ATOM	4217	O	ILE	447	41.633	71.632	24.450	1.00	49.18	AAAA	ATOM	4228	C6	NAG	284A	54.668	45.071	70.260	1.00	42.30	BBBB
ATOM	4218	N	ASN	448	41.662	69.790	23.156	1.00	50.47	AAAA	ATOM	4231	O6	NAG	284A	53.894	46.229	70.570	1.00	42.86	BBBB
ATOM	4219	CA	ASN	448	41.019	70.516	22.056	1.00	55.78	AAAA	ATOM	4233	C1	NAG	284B	57.951	46.656	70.868	1.00	33.26	BBBB
ATOM	4221	CB	ASN	448	40.756	69.766	20.718	1.00	59.98	AAAA	ATOM	4235	C2	NAG	284B	58.542	45.990	72.101	1.00	30.90	BBBB
ATOM	4222	CG	ASN	448	40.575	68.378	20.800	1.00	65.00	AAAA	ATOM	4237	N2	NAG	284B	57.525	45.115	72.646	1.00	27.21	BBBB
ATOM	4223	OD1	ASN	448	41.207	67.421	20.347	1.00	71.20	AAAA	ATOM	4239	C7	NAG	284B	57.806	43.866	72.976	1.00	27.34	BBBB
ATOM	4224	ND2	ASN	448	39.408	68.246	21.395	1.00	72.69	AAAA	ATOM	4240	O7	NAG	284B	58.931	43.376	72.855	1.00	20.62	BBBB
ATOM	4227	C	ASN	448	39.572	70.820	22.370	1.00	55.89	AAAA	ATOM	4241	C8	NAG	284B	56.655	43.038	73.531	1.00	33.39	BBBB
ATOM	4228	O	ASN	448	38.872	70.044	23.027	1.00	56.11	AAAA	ATOM	4245	C3	NAG	284B	58.883	47.107	73.114	1.00	33.39	BBBB
ATOM	4229	N	THR	449	39.134	71.970	21.892	1.00	55.64	AAAA	ATOM	4247	O3	NAG	284B	59.643	46.597	74.200	1.00	35.36	BBBB
ATOM	4231	CA	THR	449	37.781	72.433	22.128	1.00	55.58	AAAA	ATOM	4249	C4	NAG	284B	59.673	48.263	72.497	1.00	34.34	BBBB
ATOM	4232	CB	THR	449	37.815	73.914	22.423	1.00	57.21	AAAA	ATOM	4251	O4	NAG	284B	59.645	49.360	73.422	1.00	38.30	BBBB
ATOM	4233	CG1	THR	449	38.125	74.629	21.212	1.00	59.40	AAAA	ATOM	4252	C5	NAG	284B	59.108	48.710	71.134	1.00	32.15	BBBB
ATOM	4235	CG2	THR	449	38.913	74.198	21.479	1.00	59.18	AAAA	ATOM	4254	O5	NAG	284B	58.880	47.565	70.294	1.00	32.44	BBBB
ATOM	4236	C	THR	449	36.981	72.185	20.880	1.00	53.18	AAAA	ATOM	4255	C6	NAG	284B	60.065	49.586	70.360	1.00	30.53	BBBB
ATOM	4237	O	THR	449	35.785	72.445	20.812	1.00	52.99	AAAA	ATOM	4258	O6	NAG	284B	60.325	50.803	73.022	1.00	36.64	BBBB
ATOM	4238	N	ARG	450	37.658	71.655	19.885	1.00	52.04	AAAA	ATOM	4259	C7	NAG	284C	60.861	49.875	73.828	1.00	47.37	BBBB
ATOM	4240	CA	ARG	450	37.009	71.401	18.632	1.00	52.59	AAAA	ATOM	4260	C1	MAN	284C	60.651	51.284	74.321	1.00	52.82	BBBB
ATOM	4241	CB	ARG	450	37.957	71.755	17.483	1.00	51.20	AAAA	ATOM	4262	C2	MAN	284C	59.718	51.284	75.398	1.00	56.09	BBBB
ATOM	4242	CG	ARG	450	38.604	70.561	16.801	1.00	48.17	AAAA	ATOM	4263	O2	MAN	284C	62.000	51.840	74.779	1.00	54.95	BBBB
ATOM	4243	CD	ARG	450	39.869	70.955	16.073	1.00	46.87	AAAA	ATOM	4266	C3	MAN	284C	61.827	53.180	75.284	1.00	61.94	BBBB
ATOM	4244	NE	ARG	450	41.030	70.821	16.942	1.00	47.57	AAAA	ATOM	4268	O3	MAN	284C	62.581	50.949	75.858	1.00	50.28	BBBB
ATOM	4246	CG	ARG	450	41.612	69.664	17.217	1.00	43.50	AAAA	ATOM	4271	O4	MAN	284C	63.871	51.404	76.227	1.00	49.51	BBBB
ATOM	4247	NH1	ARG	450	41.143	68.530	16.713	1.00	45.69	AAAA	ATOM	4273	C5	MAN	284C	62.673	49.529	75.341	1.00	48.06	BBBB
ATOM	4248	NH2	ARG	450	42.654	69.643	18.051	1.00	46.29	AAAA	ATOM	4275	O5	MAN	284C	61.374	49.072	74.897	1.00	49.19	BBBB
ATOM	4253	C	ARG	450	36.633	69.958	18.546	1.00	53.95	AAAA	ATOM	4276	C6	MAN	284C	63.121	48.619	76.445	1.00	47.11	BBBB
ATOM	4254	O	ARG	450	35.498	69.603	18.214	1.00	52.79	AAAA	ATOM	4279	O6	MAN	284C	62.888	49.221	77.701	1.00	46.51	BBBB
ATOM	4255	N	ASN	451	37.601	69.118	18.877	1.00	57.18	AAAA	ATOM	4281	C1	MAN	284D	62.583	54.175	74.656	1.00	67.56	BBBB
ATOM	4257	CA	ASN	451	37.386	67.696	18.757	1.00	60.27	AAAA	ATOM	4283	C2	MAN	284D	61.843	55.516	74.763	1.00	71.37	BBBB
ATOM	4258	CB	ASN	451	38.695	66.945	18.781	1.00	58.04	AAAA	ATOM	4284	O2	MAN	284D	62.728	56.582	74.422	1.00	74.58	BBBB
ATOM	4259	CG	ASN	451	38.527	65.577	18.288	1.00	59.45	AAAA	ATOM	4287	C3	MAN	284D	60.603	55.514	73.831	1.00	72.21	BBBB
ATOM	4260	OD1	ASN	451	38.273	64.657	19.033	1.00	65.58	AAAA	ATOM	4289	O3	MAN	284D	59.978	56.792	73.836	1.00	74.67	BBBB
ATOM	4261	ND2	ASN	451	38.612	65.422	16.955	1.00	57.87	AAAA	ATOM	4291	C4	MAN	284D	60.988	55.115	72.388	1.00	71.55	BBBB
ATOM	4264	C	ASN	451	36.448	67.028	19.715	1.00	61.89	AAAA	ATOM	4293	O4	MAN	284D	59.823	55.021	71.573	1.00	71.37	BBBB
ATOM	4265	O	ASN	451	35.991	67.626	20.686	1.00	64.98	AAAA	ATOM	4295	C5	MAN	284D	61.695	53.760	72.450	1.00	68.81	BBBB
ATOM	4266	N	ASN	452	36.173	65.767	19.418	1.00	64.59	AAAA	ATOM	4297	O5	MAN	284D	62.865	53.867	73.288	1.00	48.38	BBBB
ATOM	4268	CA	ASN	452	35.275	64.944	20.208	1.00	68.31	AAAA	ATOM	4298	C6	MAN	284D	62.122	53.130	71.131	1.00	67.04	BBBB
ATOM	4269	CB	ASN	452	35.153	65.499	21.644	1.00	67.97	AAAA	ATOM	4301	O6	MAN	284D	62.724	51.856	71.334	1.00	56.70	BBBB
ATOM	4270	C	ASN	452	33.910	64.932	19.507	1.00	69.96	AAAA	END										
ATOM	4271	CG	ASN	452	33.104	63.867	19.236	1.00	70.39	AAAA											
ATOM	4272	N	GLY	453	33.469	66.127	19.126	1.00	70.80	AAAA											
ATOM	4274	CA	GLY	453	32.193	66.300	18.455	1.00	72.63	AAAA											
ATOM	4275	C	GLY	453	31.570	67.493	19.138	1.00	74.27	AAAA											
ATOM	4276	O	GLY	453	30.681	68.147	18.500	1.00	74.08	AAAA											
ATOM	4277	N	GLU	454	32.052	67.755	20.351	1.00	76.62	AAAA											
ATOM	4279	CA	GLU	454	31.588	68.878	21.159	1.00	78.96	AAAA											
ATOM	4280	CB	GLU	454	32.124	7															

Face 1	Cleft 1	Face 2	Cleft 2	Face 3
(12D) 11N 10R				
(35S) 33L 32L 8D (6G) 5P	259E 261S 262D			
(61A) 59R 58F 56L 54Y 28Y (27G) 26E 255I	256L 263S 264E			
91E 90F 82F (88V) 83Y 80K 79W	266F 275Q (283R) 282I 300K			
115K 114E 112R 85Y 84N 108R	272E 279S 298C (322G) 321Q 347F			
(140V) 138Y	240R 270D			
		305E		
			309K 312D 335R	
			(316S) 313S 336R	
			315T 314V (344V) 338N	
			310T 318Q 346Q 343E	

Figure 2

(a)



(b)

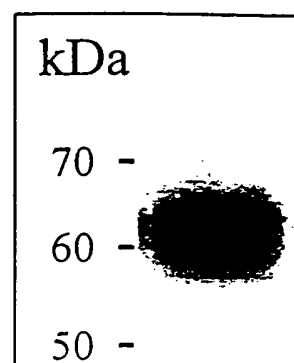
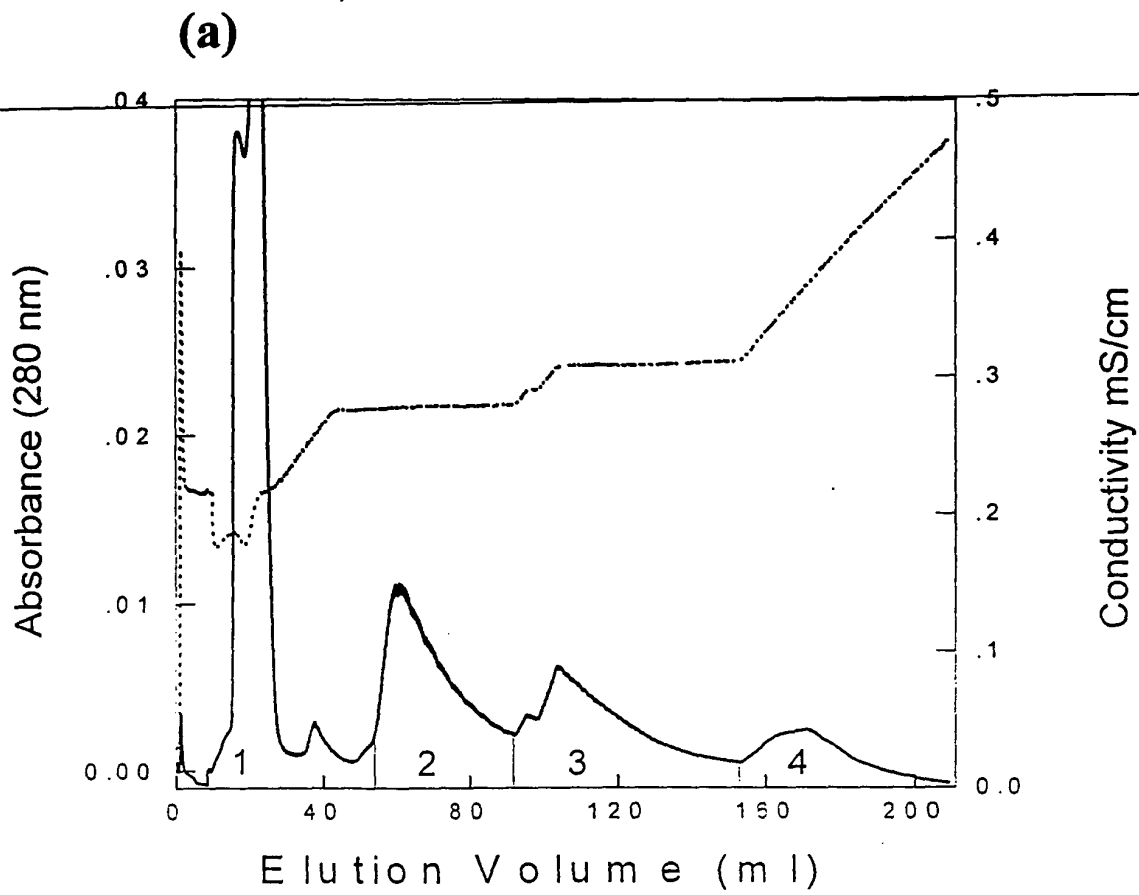
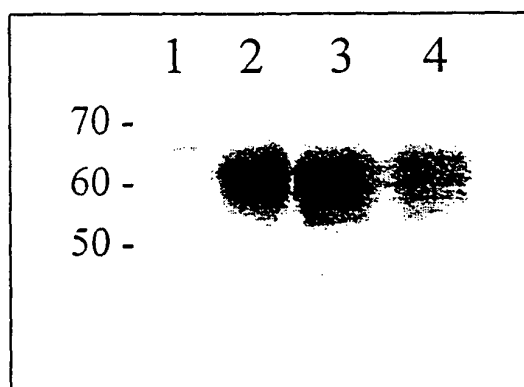


Figure 3



(b)



(c)

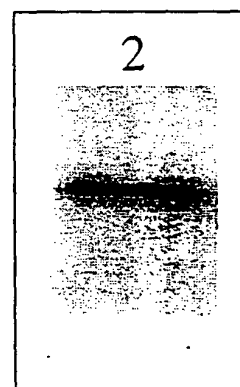


Figure 4

Figure 5

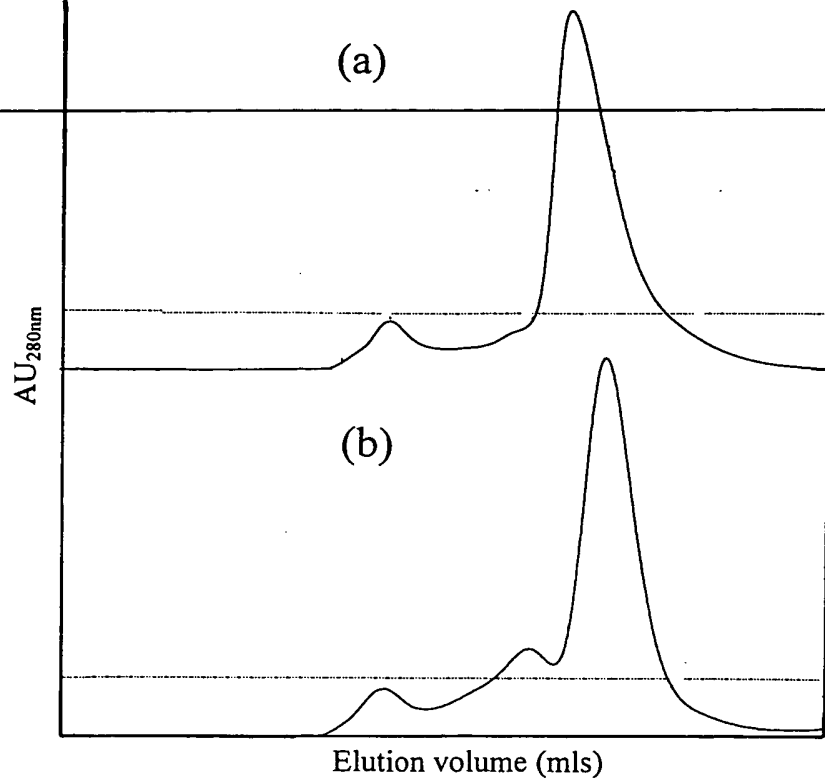
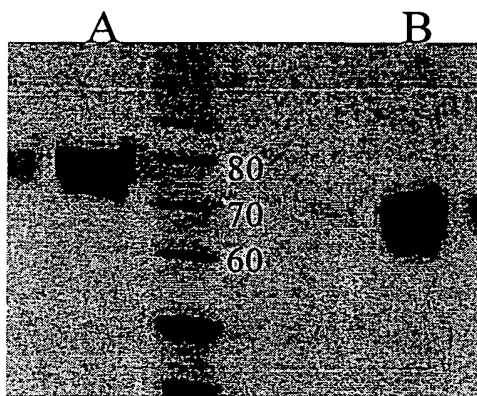
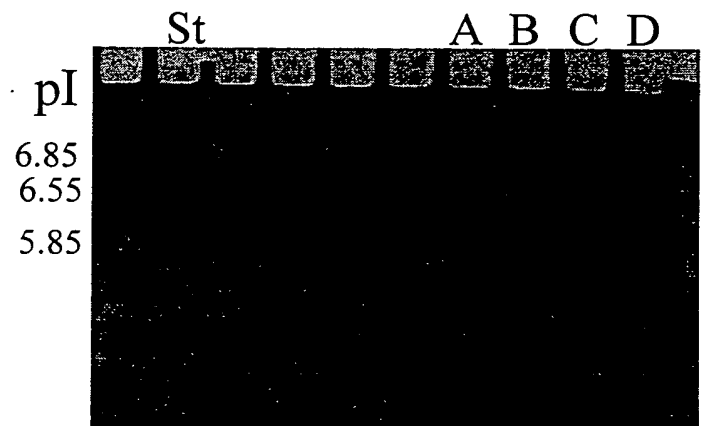


Figure 6

(a) SDS PAGE



(b) IEF pH3-7



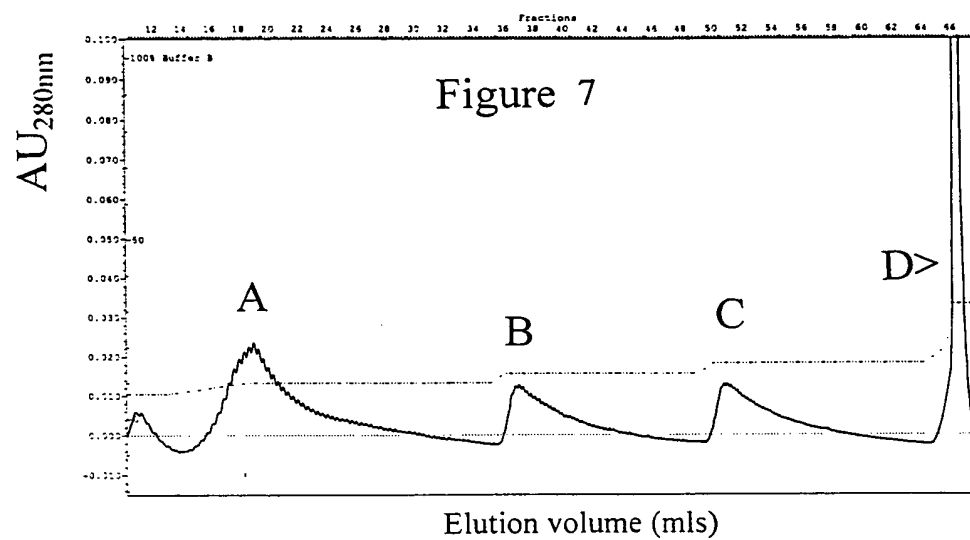




Figure 8

IGF1R L1 I H L Y P E K I C G P G I D I R N N D N L S Y Q R L K R L E N C C T S V I E G G N D L L I S K A R P E D Y R S 42
IR L1 L L E E K K V C C Q G M D I R N N D N L S Y Q R L K R L E N C C T S V I E G G N D L L I S K A R P E D Y R S 48
EGFR L1 I L L E E K K V C C Q G M D I R N N D N L S Y Q R L K R L E N C C T S V I E G G N D L L I S K A R P E D Y R S 51
EGFR L2 311 I L L E E K K V C C Q G M D I R N N D N L S Y Q R L K R L E N C C T S V I E G G N D L L I S K A R P E D Y R S 367
IR L2 310 I L L E E K K V C C Q G M D I R N N D N L S Y Q R L K R L E N C C T S V I E G G N D L L I S K A R P E D Y R S 355
IGF1R L2 300 I L L E E K K V C C Q G M D I R N N D N L S Y Q R L K R L E N C C T S V I E G G N D L L I S K A R P E D Y R S 345

Y R S S F F L L G L L I T L I Q K E V V T G Y Y K I L L L L I A L W P A R H S A L V S L S L L E S L L G K H P A F F L K N L L Q E R L L T T V I I R R G G E T L L G W K R L Y F F Y N Y A L L V I F E M T N H L A N T S K D L K E K L W D 100
L S S F L L G L L I T L I Q K E V V T G Y Y K I L L L L I A L W P A R H S A L V S L S L L E S L L G K H P A F F L K N L L Q E R L L T T V I I R R G G E T L L G W K R L Y F F Y N Y A L L V I F E M T N H L A N T S K D L K E K L W D 106
L S S F L L G L L I T L I Q K E V V T G Y Y K I L L L L I A L W P A R H S A L V S L S L L E S L L G K H P A F F L K N L L Q E R L L T T V I I R R G G E T L L G W K R L Y F F Y N Y A L L V I F E M T N H L A N T S K D L K E K L W D 108
L S S F L L G L L I T L I Q K E V V T G Y Y K I L L L L I A L W P A R H S A L V S L S L L E S L L G K H P A F F L K N L L Q E R L L T T V I I R R G G E T L L G W K R L Y F F Y N Y A L L V I F E M T N H L A N T S K D L K E K L W D 426
L S S F L L G L L I T L I Q K E V V T G Y Y K I L L L L I A L W P A R H S A L V S L S L L E S L L G K H P A F F L K N L L Q E R L L T T V I I R R G G E T L L G W K R L Y F F Y N Y A L L V I F E M T N H L A N T S K D L K E K L W D 413
L S S F L L G L L I T L I Q K E V V T G Y Y K I L L L L I A L W P A R H S A L V S L S L L E S L L G K H P A F F L K N L L Q E R L L T T V I I R R G G E T L L G W K R L Y F F Y N Y A L L V I F E M T N H L A N T S K D L K E K L W D 403

K E L P M R S S L N N N L Y N R S H R W S K H R W T T K A G G K M Y F F E H A N N P K K L L C C Y Y A S E I H K M E E V V T G T K G R Q S A S D G Q R K N D S T N G I I M S I A L T R N K K F R N G G D D C C C C C C G G D D 150
K E L P M R S S L N N N L Y N R S H R W S K H R W T T K A G G K M Y F F E H A N N P K K L L C C Y Y A S E I H K M E E V V T G T K G R Q S A S D G Q R K N D S T N G I I M S I A L T R N K K F R N G G D D C C C C C C G G D D 157
K E L P M R S S L N N N L Y N R S H R W S K H R W T T K A G G K M Y F F E H A N N P K K L L C C Y Y A S E I H K M E E V V T G T K G R Q S A S D G Q R K N D S T N G I I M S I A L T R N K K F R N G G D D C C C C C C G G D D 165
K E L P M R S S L N N N L Y N R S H R W S K H R W T T K A G G K M Y F F E H A N N P K K L L C C Y Y A S E I H K M E E V V T G T K G R Q S A S D G Q R K N D S T N G I I M S I A L T R N K K F R N G G D D C C C C C C G G D D 477
K E L P M R S S L N N N L Y N R S H R W S K H R W T T K A G G K M Y F F E H A N N P K K L L C C Y Y A S E I H K M E E V V T G T K G R Q S A S D G Q R K N D S T N G I I M S I A L T R N K K F R N G G D D C C C C C C G G D D 470
K E L P M R S S L N N N L Y N R S H R W S K H R W T T K A G G K M Y F F E H A N N P K K L L C C Y Y A S E I H K M E E V V T G T K G R Q S A S D G Q R K N D S T N G I I M S I A L T R N K K F R N G G D D C C C C C C G G D D 460

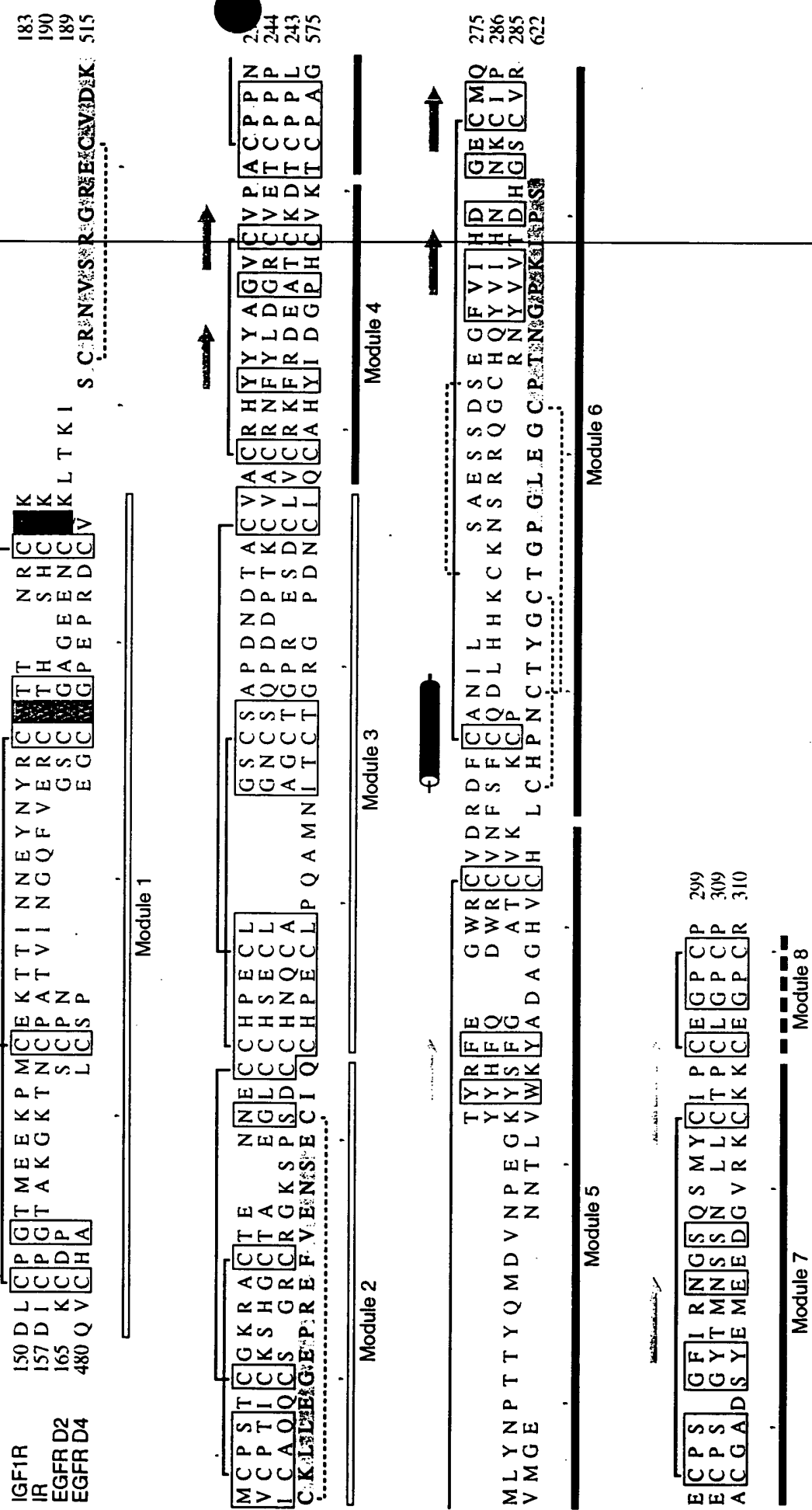


Figure 9b

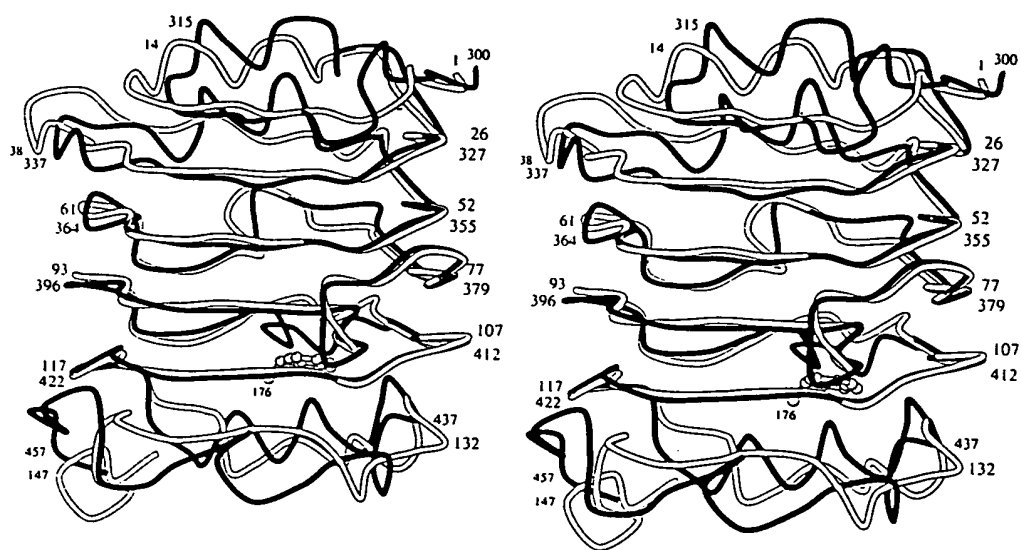


Figure 10

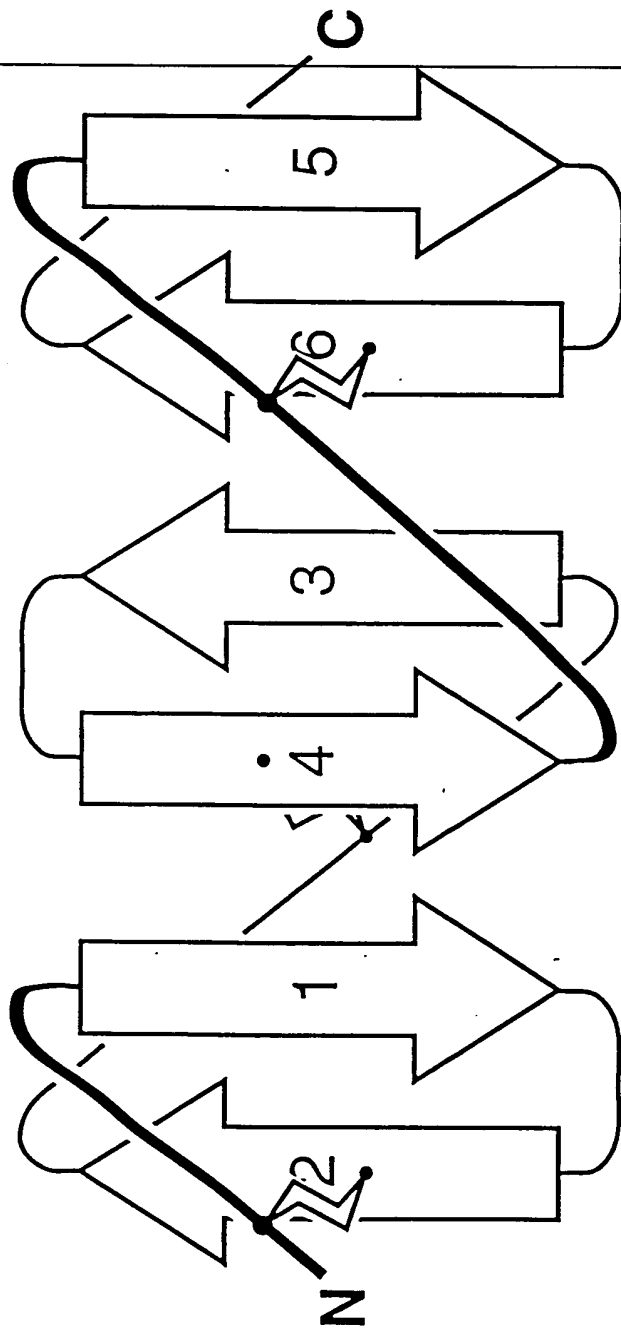


Figure 11

15/11

12/8

14/10

34/30

36/32

30/24

64/58

87/81

96/90

80/74

91/85

Figure 12

Figure 13: Sequence Alignment of hIGF-1R, hIR and hIRR ectodomains.

Derived by use of the PileUp program in the software package of the Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA.

Symbol Comparison table: GenRunData:FileUpPep.Cmp CompCheck: 1254

GapWeight: 3.0
GapLengthWeight: 0.1

Name: Higflr	Len: 972	Check: 1781	Weight: 1.00
Name: Hir	Len: 972	Check: 2986	Weight: 1.00
Name: Hirr	Len: 972	Check: 9819	Weight: 1.00

HigflrEICGP	GIDIRNDYQQ	LKRLENCTVI	EGYLGHILLIS	K..AEDYRSY	43
Hir	HLYPGEVC.P	GMDIRNNLTR	LHELENCSTVI	EGHLQILLMF	KTRPEDFRDL	49
Hirr	...MNVC.P	SLDIRSEVAE	LRQLENCSTVV	EGHLQILLMF	TATGEDFRGL	45

Higflr	RFPKLTIVITE	YLLLFRVAGL	ESLGDLFPNL	TVIRGWKLFY	NYALVIFEMT	93
Hir	SFPKLIMITD	YLLLFRVYGL	ESLKDLFPNL	TVIRGSRLFF	NYALVIFEMV	99
Hirr	SFPRLTQVTD	YLLLFRVYGL	ESLRDLFPNL	AVIRGTRLFL	GYALVIFEMP	95

Higflr	NLKDIGLYNL	RNITRGAIRI	EKNADLCYLS	TVDWSLILDA	VSNNYIVGNK	143
Hir	HLKELGLYNL	MNITRGSVRI	EKNNELCYLA	TIDWSRILDS	VEDNYIVLNK	149
Hirr	HLRDVALPAL	GAVLRGAVRV	EKNQELCHLS	TIDWGLLQPA	PGANHIVGNK	145

Higflr	PPK.ECGDLC	PGTMEEKPM.	CEKTTINNEY	NYRCWTTNRC	QKMCPTSTCGK	191
Hir	DDNEECGDIC	PGTAKGKTN.	CPATVINGQF	VERCWTHSHC	QKVCPTICKS	198
Hirr	LG.EECADV	PGVLGAAGEP	CAKTTFSGHT	DYRCWTSSHC	QRCVCPCHPG.	193

Higflr	RACTENNECC	HPECLGSCSA	PDNDTACVAC	RHYYYAGVCV	PACPPNTYRF	241
Hir	HGCTAEGLCC	HSECLGNCSQ	PDDPTKCVAC	RNFYLDGRCV	ETCPPPYHYF	248
Hirr	MACTARGECC	HTECLGGCSQ	PEDPRACVAC	RHLYFQGA	CLWACPPGTYQY	243

Higflr	EGWRCVDRDF	CANILSAES.	...SDSEGFV	IHDGECMQEC	PSGFIRNGSQ	287
Hir	QDWRVCNFSF	CQDLHHKCKN	SRROGCHQYV	IHNKNCIPEC	PSGYTMNSSN	298
Hirr	ESWRCVTAER	CASLHSVPG.RASTFG	IHQGSCLAQC	PSGFTRNSS.	287

Higflr	SMYCIPCEGP	CPKVCEEEKK	TKTIDSVTSA	QMLQGCTIFK	GNLLINIRRG	337
Hir	.LLCTPCLGP	CPKVCHLLEG	EKTIDSVTSA	QELRGCTVIN	GSLIINIRGG	347
Hirr	SIFCHKCEGL	CPKECKV..G	TKTIDSIQAA	QDLVGCTHVE	GSLILNLRQG	335

Higflr	NNIASELENF	MGLIEVVTGY	VKIRSHALV	SLSFLKNLRL	ILGEEQLEGN	387
Hir	NNLAAELEAN	LGLIEEISGY	LKIRRSYALV	SLSFFRKLRL	IRGETLEIGN	397
Hirr	YNLEPQLQHS	LGLVETITGF	LKIKHSFALV	SLGFFKNLKL	IRGDAMVDGN	385

Higflr	YSFYVLDNQN	LQQLWDWDHR	NLTIKAGKMY	FAFNPKLCVS	EIYRMEEVTG	437
Hir	YSFYALDNQN	LRQLWDWSKH	NLTITQGKLF	FHYNPKLCLS	EIHKMEEVSG	447
Hirr	YTLYVLDNQN	LQQLGSWVAA	GLTIPVGKIY	FAFNPRLCLE	HIYRLEEVTG	435

Higflr	TKGRQSKGDI	NTRNNGERAS	CESDV	LHFTS	TTTSKNRIII	TWHRYRPPDY	487
Hir	TKGRQERNDI	ALKTNGDQAS	CENEL	LKFSY	IRTSFDKILL	RWEPYWPPDF	497
Hirr	TRGRQNKAIEI	NPRTNGDRAA	CQTRT	LRFVS	NVTEADRILL	RWEREYPLEA	485

				*			
Higflr	RDLISFTVYY	KEAPFKNVTE	YDQDAGCSN	SWNMVDVDLPPNKDV	532	
Hir	RDLLGFMLFY	KEAPYQNVTE	FDGQDAGCSN	SWTVVDIDPP	LRSNDPKSQN	547	
Hirr	RDLLSFIVYY	KESPFONATE	HVGPDACGTQ	SWNLLDVLP	L.....SRTQ	530	
Higflr	EPGILLHGLK	PWTQYAVYVK	AVTLTMVEND	HIRGAKSEIL	YIRTNASVPS	582	
Hir	HPGWLMRGLK	PWTQYAIQVK	TL.VTFSDER	RTYGAKSDII	YVQTDATNPS	596	
Hirr	EPGVTLASLK	PWTQYAVFVR	AITLTTEEDS	PHQGAQSPIV	YLRTLPAAPT	580	
Higflr	IPLDVLASN	SSSQLIVKWN	PPSLPNGNLS	YYIVRWQROP	QDGYLYRHNY	632	
Hir	VPLDPISVSN	SSSQIILKWK	PPSDPNGNIT	HYLVFWERQA	EDSELFELDY	646	
Hirr	VPQDVISTSN	SSSHLLVRWK	PPTQRNGNLT	YYLVWLQRLA	EDGDLYLNDY	630	
				*	** ** *		
Higflr	CSKD.KIPIR	KYADGTIDIE	EV TENPKTEV	CGGEKGPCCA	C...PKTEAE	678	
Hir	CLKGLKLPSR	TWS.PPFESE	DSQKHNOSE.	YEDSAGECCS	C...PKTDSQ	691	
Hirr	CHRGRLRLPTS	N.NDPRFDGE	DGDPEAEME.SDCCP	CQHPPPGQVL	673	
				α -----><----- β			
Higflr	KQAEKEEAAY	RKVFENFLHN	SIFVPRPERK	RRDVMQVANT	TMSSRSRNTT	728	
Hir	ILKELEESSF	RKTFEDYLHN	VVFVPRPSRK	RRSLGDVGNV	TVAVP...TV	738	
Hirr	PPLAQEASF	QKKFENFLHN	AITIPISPKW	VTSINKSPQR	D.SGRHRRAA	722	
				*			
Higflr	AA..DTYNIT	DPEELETEYP	FFESRVDNKE	RTVISNLRPF	TLYRIDIHSC	776	
Hir	AAFPNTSSTS	VPTSPEEHRP	F..EKVVNKE	SLVISGLRHF	TGYRIELQAC	786	
Hirr	GPLRLGNGSS	DFEIQEDKVPRE	RAVL SGLRHF	TEYRIDIHAC	764	
				*			
Higflr	NHEAEKLGCS	ASN FVFARTM	PAEGADDIPG	PVTWEPRPEN	SIFLKWPEPE	826	
Hir	NQDTPEERCS	VAA YVSARTM	PEAKADDIVG	PVTHEIFENN	VVHLMWQEPK	836	
Hirr	NHAAHTVGCS	AATFVFARTM	PHREADGIPG	KVAWEASSKN	SVLLRWLEPP	814	
				*			
Higflr	NPNGLILMYE	IKYGS.QVED	QRECVSRQEY	RKYGGAKLNR	LNPGNYTARI	875	
Hir	EPNGLIVLYE	VS YRRYGDEE	LHLCVSRKHF	ALERG CRLRG	LSPGNYSVRI	886	
Hirr	DPNGLILKYE	IKYRRLGEEA	TVLCVSRRLRY	AKFGGVHLAL	LPPGNY SARV	864	
Higflr	QATSLSGNGS	WTD PVFFYVQ	AKTGYENFIH	L		906	
Hir	RATSLAGNGS	WTEPTYFYVT	DYLDVPSNIA	K		917	
Hirr	RATSLAGNGS	WTDSVAFYIL	GPEEDAGGL	H		895	

Figure 14: Sequence Alignment of EGFR, ErbB2, ErbB3 and ErbB4 Ectodomains.

[For alignment on the IGF-1R fragment see Fig. 9]

Derived by use of the PileUp program in the software package of the Genetics Computer Group, 575 Science Drive, Madison, Wisconsin, USA.

Symbol comparison table: GenRunData.Pileuppp.Cmp CompCheck: 1254

GapWeight: 3.000
GapLengthWeight: 0.100

Name: Erb3	Len: 649	Check: 4625	Weight: 1.00
Name: Erb4	Len: 649	Check: 790	Weight: 1.00
Name: Egfr	Len: 649	Check: 2381	Weight: 1.00
Name: Erb2	Len: 649	Check: 8174	Weight: 1.00

	1				50
Erb3	SEVGNSQAVC	PGTLNGLSVT	GDAENQYQTL	YKLYERCEVV	MGNLEIVLTG
Erb4	...SDSQSVC	AGTENKLSSL	SDLEQQYRAL	RKYENCEVV	MGNLEITSIE
Egfr	...LEEKKVC	QGTSNKLTQL	GTTFEDHFLSL	QRMFNCEVV	LGNLEITYVQ
Erb2STQVC	TGTDMKLRPL	ASPETHLDML	RHLYQGCQVV	QGNLELTYP

	51				100
Erb3	HNADLSFLQW	IREVTGYVLV	AMNEFSTLPL	PNLRVVRGTQ	VYDGKFAIFV
Erb4	HNRDLSFLRS	VREVTGYVLV	ALNQFRLPL	ENLRIIRGTK	LYEDRYALAI
Egfr	RNYDLSFLKT	IQEVAGYVLI	ALNTVERIPL	ENLQIIRGNM	YYENSYALAV
Erb2	TNASLSFLQD	IQEVQGYVLI	AHNQVRQVPL	QRLRIVRGTQ	LFEDNYALAV

	101				150
Erb3	MLNYN.....TNSSHA	LRQLRLTQLT	EILSGGVYIE	KNDKLCHMDT
Erb4	FLNYR.....KDGNGF	LQELGLKNLT	EILNGGVYVD	QNKFLCYADT
Egfr	LSNYD.....ANKT.G	LKELPMRNLQ	EILHGAVRFS	NNPALCNVES
Erb2	LDNGDPLNNT	TPVTGASPGG	LRELQLRSLT	EILKGGVLIQ	RNPQLCYQDT

	151				200
Erb3	IDWRDIVRDR	...DAEIVVK	DNGRSCPPCH	EVC.KGRCWG	PGSEDCQTLT
Erb4	IHWQDIVRNP	WPSNLTIVST	NGSSGCGRCH	KSC.TGRCWG	PTENHCQTLT
Egfr	IQWRDIVSSD	FLSNMSMDFQ	NHLGSCQKCD	PSCPNGSCWG	AGEENCQKLT
Erb2	ILWKDIFHKN	NQLALTIDT	NRSRACHPCS	PMCKGSRCWG	ESSEDCQSLT

	201				250
Erb3	KTICAPQCNG	HCFGPNPNQC	CHDECAGGCS	GPQDTCDFAC	RHFNDSGACV
Erb4	RTVCAEQCDG	RCYGPVVSDC	CHRECAGGCS	GPKDTCDFAC	MNFNDSGACV
Egfr	KIICAQCSG	RCRGKSPSDC	CHNQCAAGCT	GPRESDECLVC	RKFRDEATCK
Erb2	RTVCAGGC.A	RCKGPLPTDC	CHEQCAAGCT	GPKHSDCLAC	LHFNHSGICE

	251				300
Erb3	PRCPQPLVYN	KLTFQLEPNP	HTKYQYGGVC	VASCPHNFVV	.DQTSVCVRAC
Erb4	TQCPQTFVYN	PTTFQLEHNF	NAKYTYGAF	VKKCPHNFVV	.DSSSCVRAC
Egfr	DTCPPMLLYN	PTTYQMDVNP	EGKYSFGATC	VKKCPRNYVV	TDHGSCVRAC
Erb2	LHCPALVTYN	TDTFESMPNP	EGRYTFGASC	VTACPYNVLS	TDVGSCTLVC

	301				350
Erb3	PPDKMEV.DK	NGLKMCEPCG	GLCPKACEGT	SGGSRF..QT	VDSSNIDGFV
Erb4	PSSKMEV.EE	NGIKMCKPCT	DICPKACDGI	GTGSLMSAQT	VDSSNIDKFI
Egfr	GADSYEM.EE	DGVRKCKKCE	GPCRKVCNGI	GIGEFKDSLS	INATNIKHFK
Erb2	PLHNQEVTA	DGTQRCEKCS	KPCARVCYGL	GMEHLREVRA	VTSANIQEFA

	351				400
Erb3	NCTKILGNLD	FLITGLNGDP	WHKIPALDPE	KLVNFRVTVRE	ITGYLNIQSW
Erb4	NCTKINGNLI	FLVTGIHGDP	YNAIEAIDPE	KLVNFRVTVRE	ITGFLNIQSW
Egfr	NCTSISGDLH	ILPVAFRGDS	FTHTPPLDPQ	ELDILKTVKE	ITGFLLIQAW
Erb2	GCKTIFGSLA	FLPESFDGDP	ASNTAPLQPE	QLQVFETLEE	ITGYLYISAW

	401				450
Erb3	PPMHNFVSFV	SNLTTIGGRS	LYNRGFSLLI	MKNLNVTSLG	FRSLKEISAG
Erb4	PPNMTDFS	SNLVTIGGRV	LYS.GLSLLI	LKQQGITS	LQFSLKEISAG

Egfr	PENRTDLHAF	ENLEIIRGRT	KQHGGFSLAV	VS.LNITSLG	LRSLEISDG
Erb2	PDSLPLDSVF	QNLQVIRGRI	LHNGAYSL.T	LQGLGISWLG	LRSLELGS

	451			End L2 domain>	500
Erb3	RIYISANRQL	CYHSLNWT	VLRGPTTEERL	DIKHNRPRRD CVA	EGKVCDP
Erb4	NIYITDNSNL	CYYHTINWTT	LF.STINQRI	VIRDNRKAEN CTA	EGMVCNH
Egfr	DVIISGNKNL	CYANTINWKK	LF.GTSGQKT	KIISNRGENS CKA	TGQVCHA
Erb2	LALIHNNTHL	CFVHTVPWDQ	LFRNP.HQAL	LHTANRPEDE CVG	EGLACHQ

	501				550
Erb3	LCSSGGCWGP	GPGQCLSCRN	YSRGGVCVTH	CNFLNGEPRE	FAHEAECFSC
Erb4	LCSSDGCWGP	GPDQCLSCRR	FSRGRICIES	CNLYDGEFRE	FENGSIKVEC
Egfr	LCSPEGCWGP	EPRDCVSCRN	VSRGRECVDK	CKLLEGEPRE	FVENSECIQC
Erb2	LCARGHCWGP	GPTQCVNCSQ	FLRGQECVEE	CRVLQGLPRE	YVNARHCLPC

	551				600
Erb3	HPECQPMG	TATCNGSGSD	TCAQCAHFRD	GPHCVSSCPH	GVLGA.KGP.
Erb4	DPQCEKMEDG	LLTCHGPGPD	NCTKCSHFCD	GPNCVEKCPD	GLQGA.NSF.
Egfr	HPECLPQAMN	I.TCTGRGPD	NCIQCAHYID	GPHCVKTCPA	GVMGENNTL.
Erb2	HPECQPMG	SVTCFGPEAD	QCVACAHYKD	PPFCVARCPS	GVKPDLSYMP

	601				649
Erb3	IYKYPDVQNE	CRPCHENCTQ	GCKGPELQDC	L.....GQT.
Erb4	IFKYADPDRE	CHPCHPNCTQ	GCNGPTSHDC	IYYPWTGHST	LPQHARTPL
Egfr	VWKYADAGHV	CHLCHPNCTY	GCTGPGLEGC	PTNGPKIPS.
Erb2	IWKFPDEEGA	CQPCPINCTH	SCVDLDDKGC	PAEQRASPLT	S.....

Figure 15. Classification of Cys-rich modules

C2-4 denote modules with the 1-3/2-4 double disulphide bond connections.
C1-2 for the single disulphide bonded modules and
C1-2t for stabilised beta turn.

First Cys-rich region

C2-4 modules

		1	2	3	4		
Higflr	152	CPGTMEEKPM-CEKTTIINNEYNYRCWTTNRC	QKM	184	(1st)		
Hir	159	CPGTAKGKTH-CPATVINGQEVERCATHSHC	QKV	191	(1st)		
Hirr	154	CPGV/LGAAGEPCAKTTFSGHTDYRCWTSSHC	QRV	187	(1st)		
Egfr	166	CDPSCPNG-SCWGAG-EENC	QKLTKEI	190	(1st)		
hErb2	174	CSPHCKGS-RCWGES-SEDC	QSLTRTV	198	(1st)		
hErb3	167	CHEVCKG--RCWGP-SEDC	QTLTKTI	190	(1st)		
hErb4	167	CHKSGTG--RCWGPT-ENHC	QTLTRTV	190	(1st)		
Higflr	185	CPSTCGK-RACTEN---NEC		200	(2nd)		
Hir	192	CPTICKS-HGCTAE---GLC		207	(2nd)		
Hirr	188	CP--CPHGMACTAR---GEC		202	(2nd)		
Egfr	191	CAQQCSG--RCRGKS-PSDC		207	(2nd)		
hErb2	199	CAGGCA---RCKGPL-PTDC		214	(2nd)		
hErb3	191	CAQCNG--HCGPH-PNQC		207	(2nd)		
hErb4	191	CAEQCDG--RCYGPY-VSDC		207	(2nd)		
Higflr	201	CHPECLG--SCSAPDNDTAC	VA	220	(3rd)		
Hir	208	CHSECLG--NCSQPDPTKC	VA	227	(3rd)		
Hirr	203	CHTECLG--GCSQPEDPRAC	VA	222	(3rd)		
Egfr	208	CHNQCAA--GCTGPR-ESDC	LV	226	(3rd)		
Erb2	215	CHQCAA--GCTGPK-HSDC	LA	233	(3rd)		
hErb3	208	CHDECAG--GCSGPK-DTDC	FA	226	(3rd)		
hErb4	208	CHDECAG--GCSGPK-DTDC	FA	226	(3rd)		

C1-2 modules

Higflr	231	CRHY---YAGVC	VPA	233	(4th)
Hir	238	CRHY---LDGRC	VET	240	(4th)
Hirr	233	CRHY---FQGAC	LWA	235	(4th)
Egfr	237	CRHFR---DEATC	KDT	239	(4th)
hErb2	234	CRHEN---HSGIC	ELR	246	(4th)
hErb3	237	CRHEN---DSGAC	VPR	239	(4th)
hErb4	237	CRHEN---DSGAC	VTQ	239	(4th)
Higflr	254	CPENTYREFGWRC	VDRDF	251	(5th)
Hir	241	CPRETYHEQDWRC	VNESE	259	(5th)
Hirr	236	CPSTYQYESWRC	VTAER	253	(5th)
Egfr	240	CPPLMLXNETTYQNDVHPEGKYSFGATC	VKK	270	(5th)
hErb2	247	CPALVTYHDTFESHPNPEGRYTEGASC	VTA	277	(5th)
hErb3	240	CPQPLVINKLTFQLEPNPHTKYQYGGVC	VAS	270	(5th)
hErb4	240	CPQTEVINPTTFQLEHNFNAKYTYGAF	VKK	270	(5th)
Higflr	252	CAWLSAESSDSEG....FVIHD.GEC	MQE	276	(6th)
Hir	259	CQD.LHHCKNSRRQSGHQYVIHN.NKC	IFE	287	(6th)
Hirr	254	CAS.LHSVPGRASST....FGIHQ.GSC	LAQ	276	(6th)
Egfr	271	CPENYVVDHGGSC	VRA	286	(6th)
hErb2	279	CPENYVVDHGGSC	TLV	293	(6th)
hErb3	271	CPENYVVDHGGSC	VRA	285	(6th)
hErb4	271	CPENYVVDHGGSC	VRA	285	(6th)
Higflr	277	CPSSG.FIRNGSQ-SHYC	IP	293	(7th)
Hir	280	CPSSG.VTINSSH--LLC	TP	303	(7th)
Hirr	278	CPSSG.FTRISS--SIFC	HK	293	(7th)
Egfr	297	CSADSYENE-EDGVRKC	KK	304	(7th)
hErb2	294	CPHHNQEVTAEDGTQRC	EK	312	(7th)
hErb3	296	CPFQKIEVDKH-GLKIC	EP	303	(7th)
hErb4	296	CPSSKIEVEEH-GIKIC	KP	303	(7th)

C1-2t module

Higflr	294	CEGPC	298	(8th)
Hir	304	CLGPC	308	(8th)
Hirr	294	CEGLC	298	(8th)
hEgfr	305	CEGPC	309	(8th)
hErb2	313	CSKPC	317	(8th)
hErb3	304	CGGLC	308	(8th)
hErb4	304	CTDIC	308	(8th)

Second Cys-rich region.**C2-4 modules**

hEgfr	482	CHALCSP-----EGCWGPEPRDCVS	501	(1st)
hErb2	490	CHQLCAR-----GHCWGPGPTQCVN	509	(1st)
hErb3	481	CDPLCSS-----GGCWGPGPGQCLS	500	(1st)
hErb4	481	CNHLCSS-----DGCWGPGPDQCLS	500	(1st)
Egfr	534	CHPECLPQAM-NITCTGRGPDNC IQ	557	(4th)
hErb2	542	CHPECQPQNG-SVTCFGPEADQC VA	565	(4th)
hErb3	533	CHPECQPMEG-TATCNGSGSDTC AQ	556	(4th)
hErb4	533	CDPQCEKMEDGLLTCHGPGPDNC TK	557	(4th)
hEgfr	596	CHPNCTY-----GCTGPGLEGC PTNGPKIPS/	621	(7th)
hErb2	605	CPINCTH-----SCVDLDDKGC PAEQRAQRASPLTS/	632	(7th)
hErb3	594	CHENCTQ-----GCKGPELQDC LGQT/	614	(7th)
hErb4	595	CHPNCTQ-----GCNGPTSHDC IYYPWTGHSTLPQHARTPL	630	(7th)

C1-2 modules

hEgfr	502	CRNVS---RGREC VDK	514	(2nd)
hErb2	510	CSQFL---RGQEC VEE	522	(2nd)
hErb3	501	CRNYS---RGGVC VTH	513	(2nd)
hErb4	501	CRRFS---RGRIC IES	513	(2nd)
hEgfr	515	CKLLEGEPRFVENSEC IQ	533	(3rd)
hErb2	523	CRVLQGLPREYVNRHC LP	541	(3rd)
hErb3	514	CNFLNGEPREFAHEAEC FS	532	(3rd)
hErb4	514	CNLYDGEFREFENGSIK VE	532	(3rd)
hEgfr	558	CAHYI---DGPHC VKT	570	(5th)
hErb2	566	CAHYK---DPPFC V-A	578	(5th)
hErb3	557	CAHFR---DGPHC V-S	569	(5th)
hErb4	558	CSHFK---DGPNC VEK	570	(5th)
hEgfr	571	CPAGVMGENNTL-VWKYADAGHVC HL	595	(6th)
hErb2	579	CPSGVKPDLSYMPIWKFPDEEGAC QP	604	(6th)
hErb3	570	CPHGVLGAKG--PIYKYPDVQNEC RP	593	(6th)
hErb4	571	CPDGLQGANS--FIFKYADPDREC HP	594	(6th)

See Pattern is:

IR family: C2-4, C2-4, C2-4, C1-2, C1-2, C1-2, C1-2, C1-2t
EGFR family: 1st C2-4, C2-4, C2-4, C1-2, C1-2, C1-2, C1-2, C1-2t
2nd C2-4, C1-2, C1-2,
C2-4, C1-2, C1-2,
C2-4

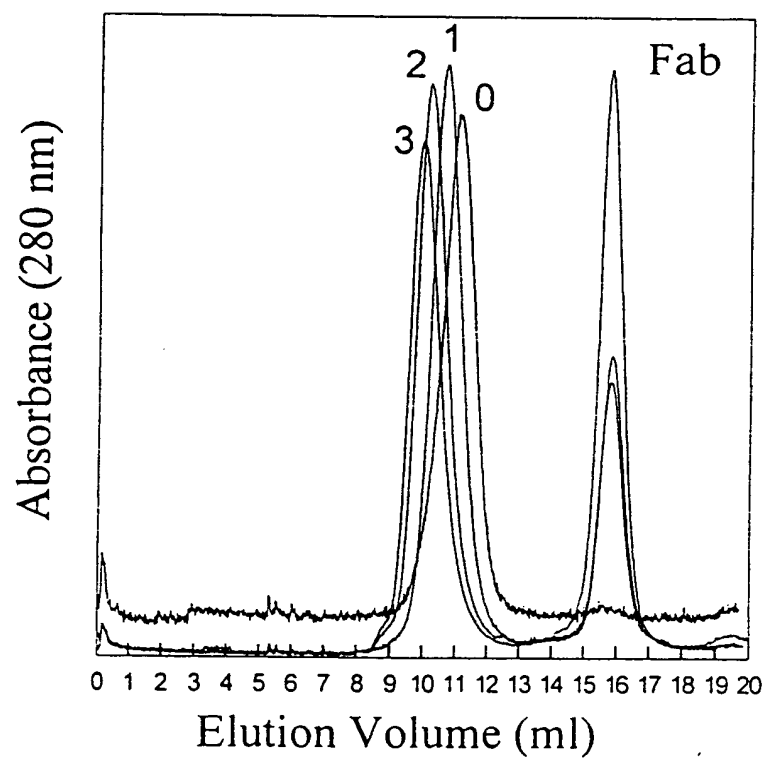
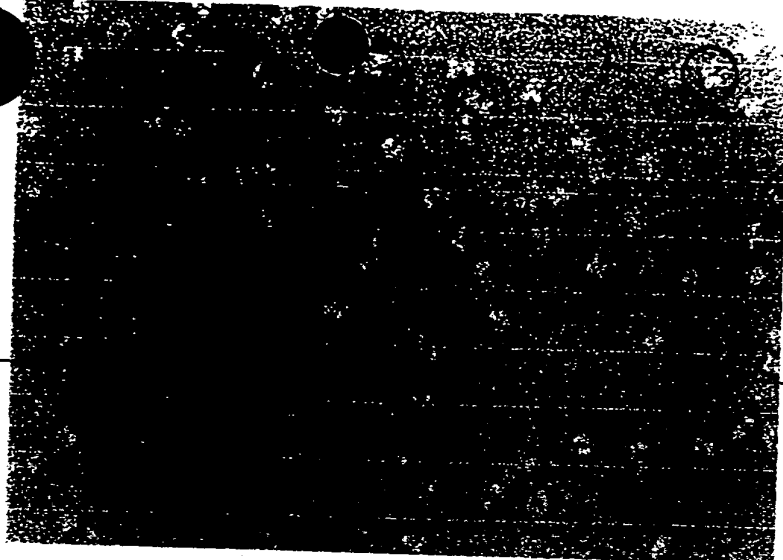
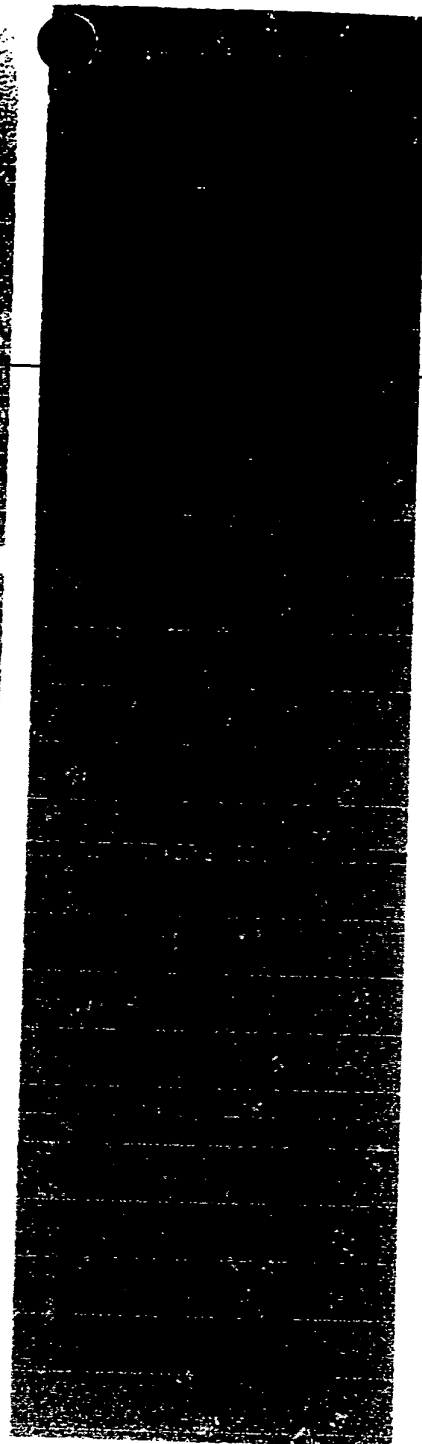


Figure 16

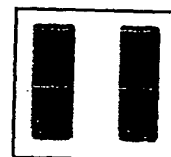


(h)



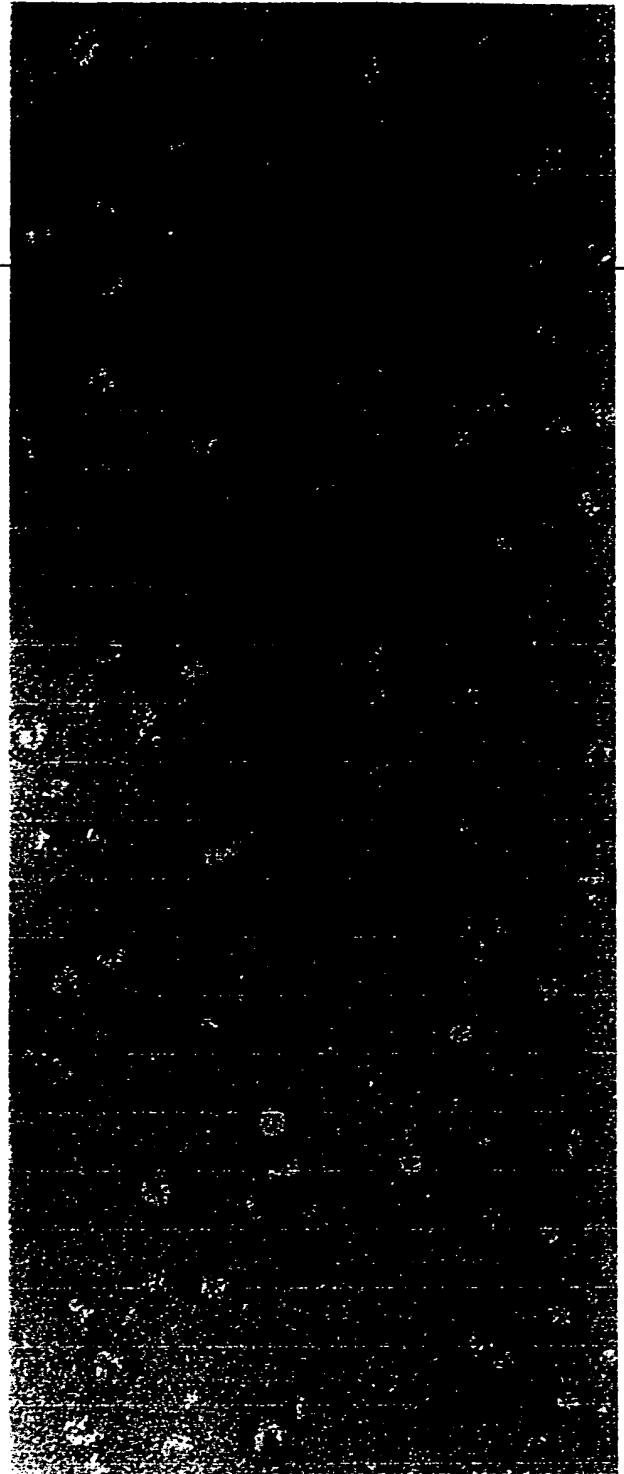
(c)

Figure 17





(a)



(b)



Figure 18

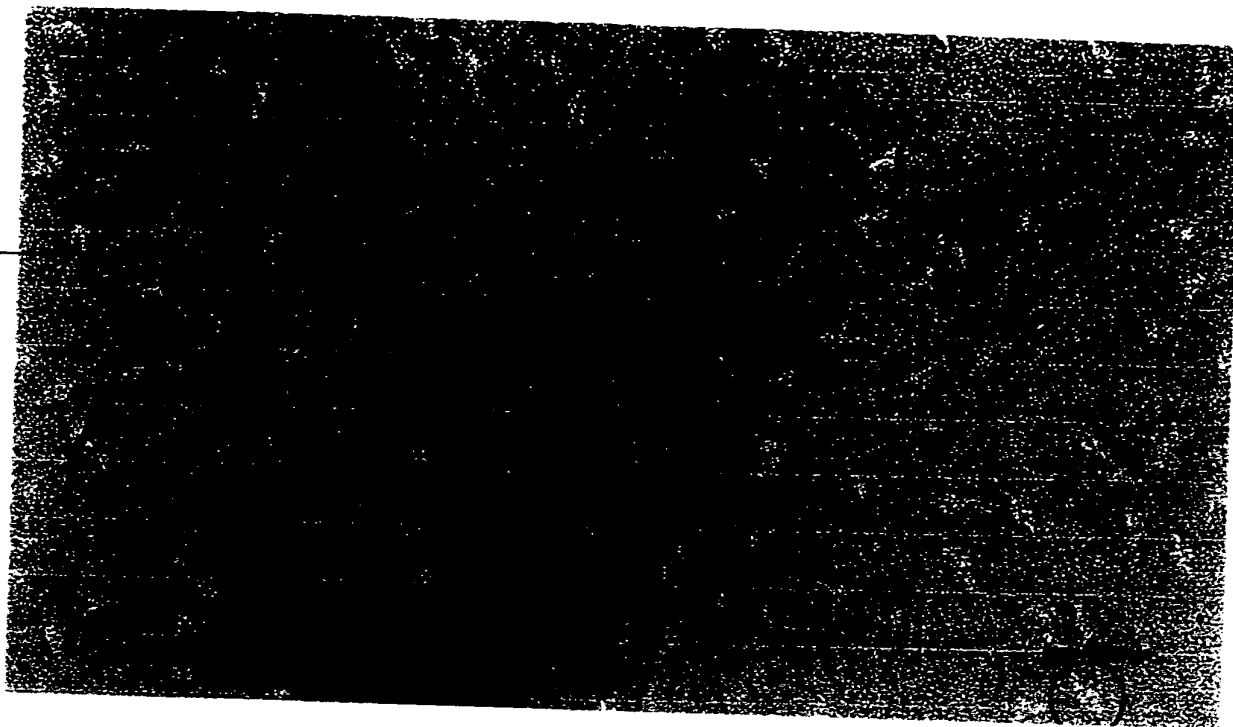


Figure 19

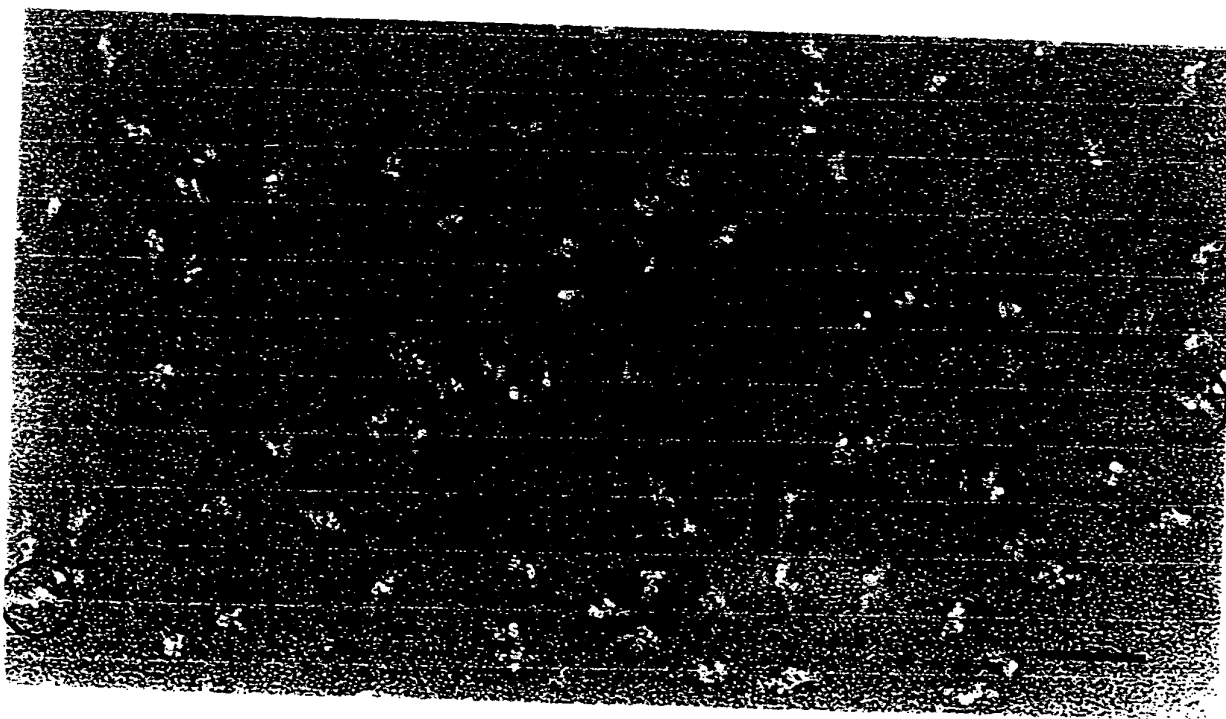


Figure 20

(a)



(b)

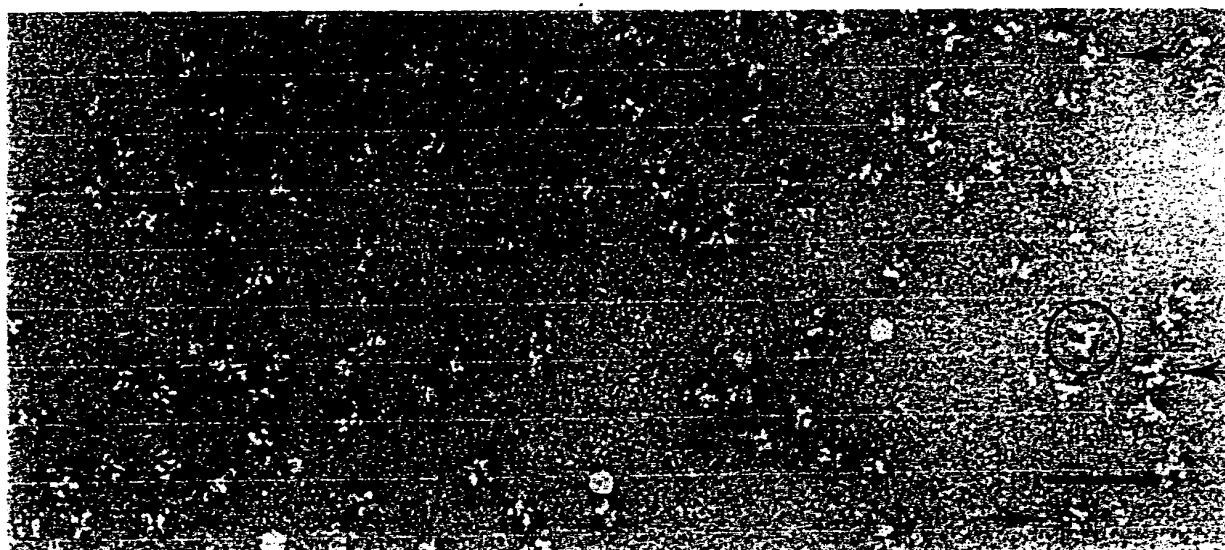


Figure 21



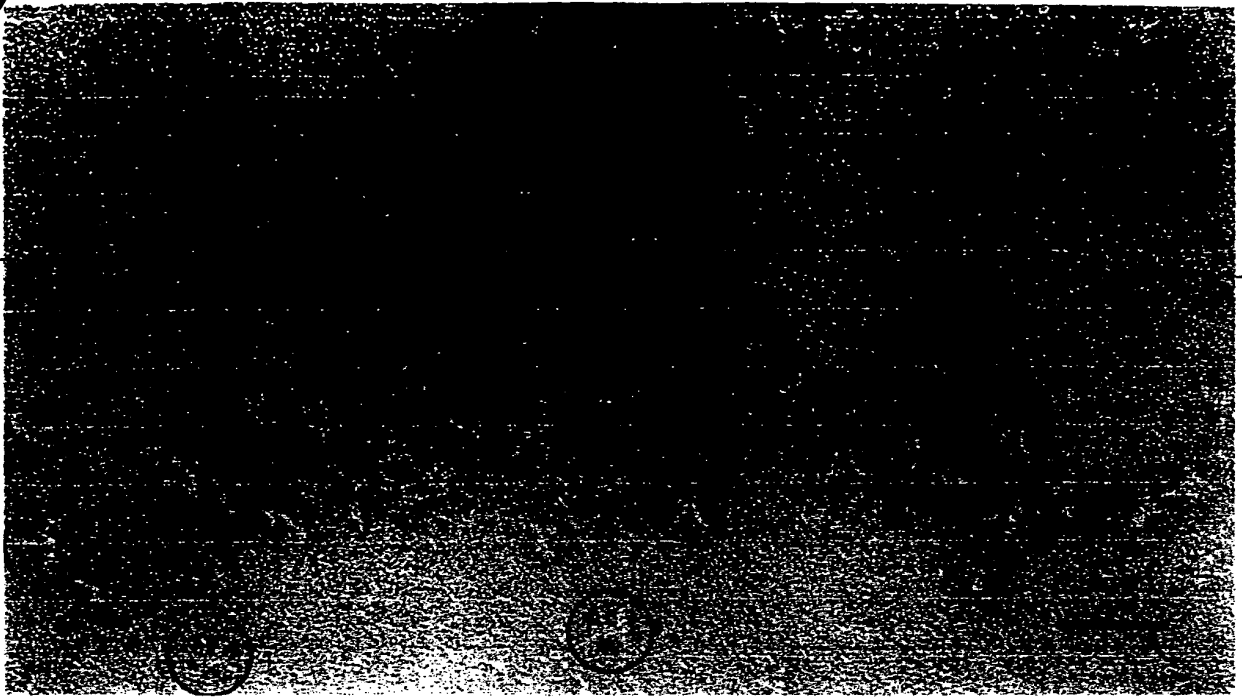


Figure 22

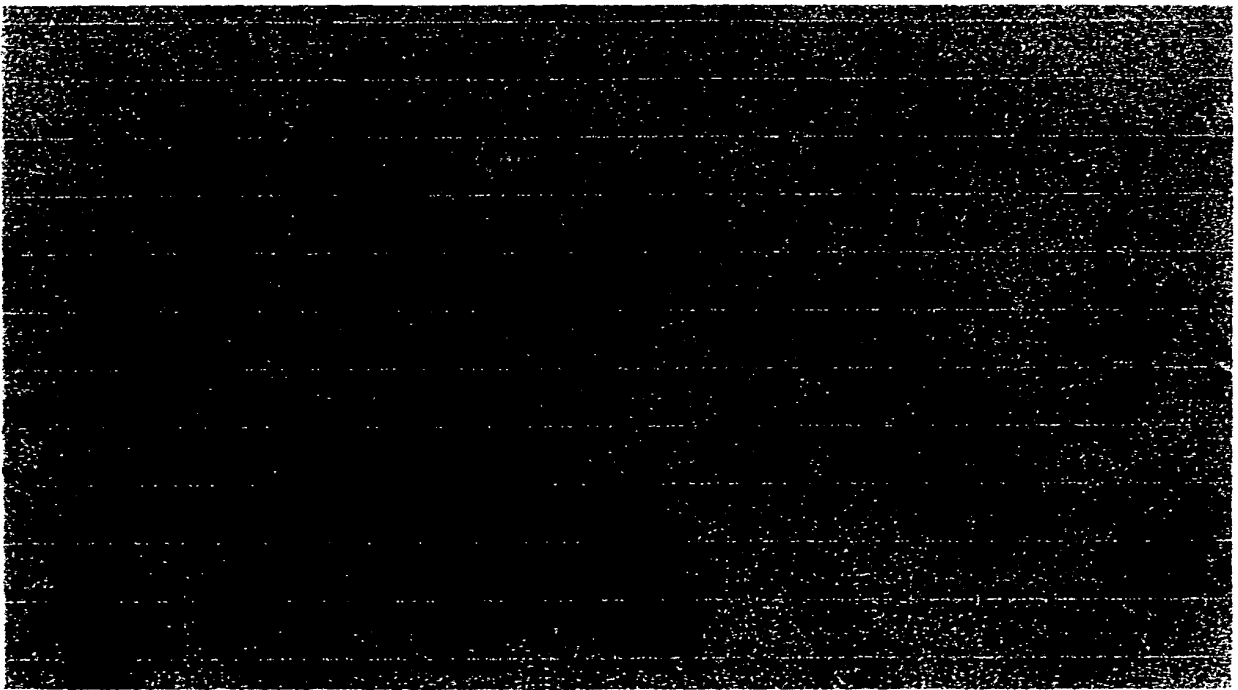
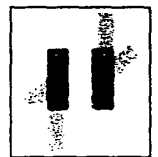


Figure 23

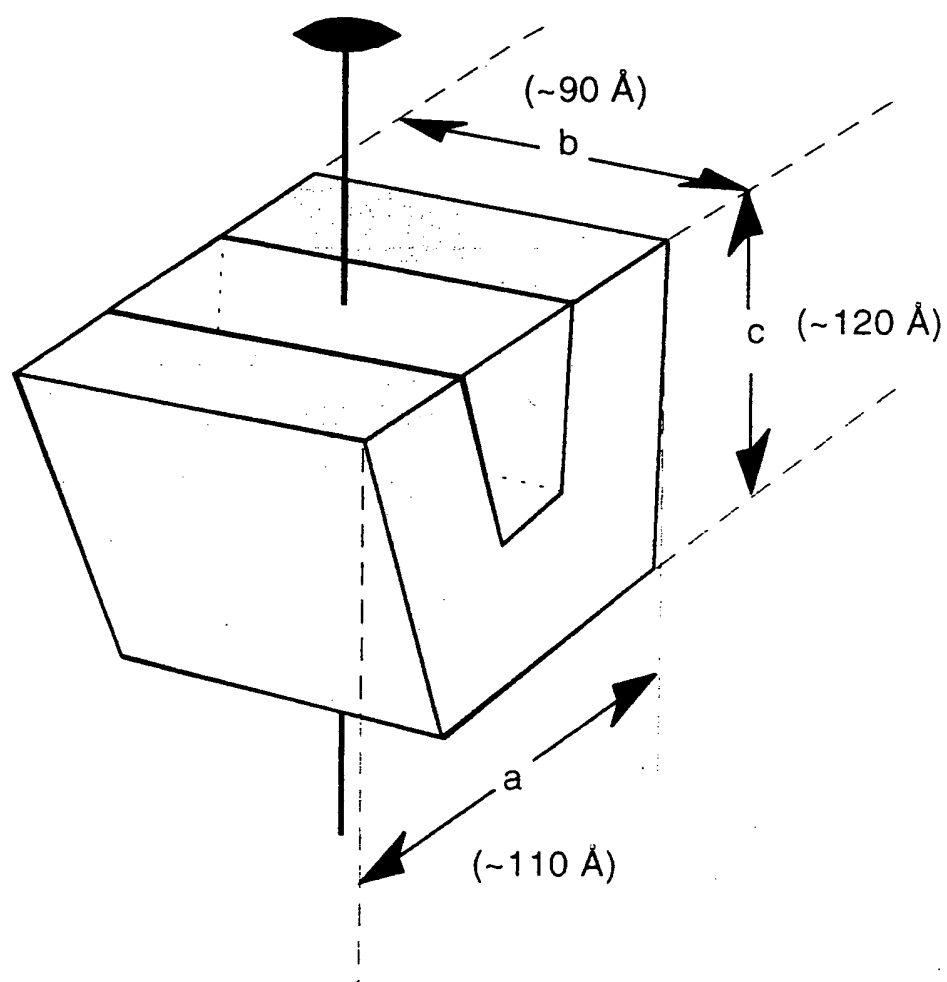


Figure 24

